

A REVIEW OF MAMMAL-ASSOCIATED PSOROPTIDIA (ACARIFORMES: ASTIGMATA)

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ABSTRACT: Mammal-associated psoroptidians belonging to the superfamily Sarcoptoidea (Acariformes: Astigmata: Psoroptidia) reviewed at the familial level. Keys to all genera and lists of their species along with verified host records are provided. The families of the superfamily Sarcoptoidea (12 families and 3 subfamilies *incertae sedis*, more than 160 genera and 1000 species) belongs to two familial complexes deserving superfamilial status: Sarcoptid complex — Atopomelidae, Chirodiscidae, Chirohynchobiidae, Gastronyssidae, Lemurnyssidae, Listrophoridae, Myocoptidae, Pneumocoptidae, Sarcoptidae, Rhyncoptidae, and 2 subfamilies *incertae sedis* deserving familial status, Dromiciocoptinae (described as subfamily of Myocoptidae) and Listropsoralginae (described as subfamily of Psoroptidae); Psoroptid complex — Lobalgidae, Psoroptidae, and the subfamily Paracoroptinae (described as a subfamily of Psoroptidae) deserving familial status. The following taxonomic changes were made — **Atopomelidae:** (i) the subfamily Centetesiinae Fain, 1976 **syn. nov.** is synonymized with Atopomelinae; (ii) the genus *Atopomelopsis* Fain, 1972 **stat. res.** is restored with one species *A. longipilis* (Fain, 1971); (iii) *Bathyergolichus cryptomys* Fain et Bochkov, 2003 **syn. nov.** synonymized with *Bathyergolichus hottentotus* Fain, 1986. **Listrophoridae:** (i) the genus *Amlistrophorus* Fain, 1981 **stat. nov.** (formerly subgenus of the genus *Afrolistrophorus*) received the full generic status; (ii) The subgenus *Spalacarus* Fain, 1980 **stat. nov.** (formerly genus) included to the genus *Afrolistrophorus*; (iii) the subgenus *Teinolistrophorus* Fain, 1981 **syn. nov.** is synonymized with the subgenus *Spalacarus*; (iv) the subgenus *Mexicochirus* Fain et Estebanes, 1996 (formerly subgenus of the genus *Asiochirus*) is included to the genus *Olistrophorus*; (v) *Amlistrophorus venezuelensis* Fain et Lukoschus, 1983 **comb. nov.** (formerly species of the subgenus *Afrolistrophorus*) is included to the genus *Amplistrophorus*. **Myocoptidae:** a new monobasic genus *Apocalypsis* Bochkov **gen. nov.** is described with the type species *Gliricoptes allactaga* Fain et Lukoschus, 1979. **Psoroptidae:** (i) the following species are considered as valid in the genus *Psoroptes*: *Psoroptes ovis* (von Hering, 1838) [= *Psoroptes equi* (von Hering, 1838), *Psoroptes cuniculi* (Delafond, 1859), and *Psoroptes cervinus* (Ward, 1915)], *Psoroptes natalensis* (Hirst, 1919), and *P. pienaar* (Fain, 1970); *Chorioptes japonensis* Takahashi et Nogami, 2001 **syn. nov.** is synonymized with *Chorioptes bovis* (von Hering, 1845).

KEY WORDS: Psoroptidia, mammals, mites, Sarcoptoidea, symbiotes, systematics

INTRODUCTION

The absolute majority of astigmatan mites (Astigmata) permanently living on vertebrates belongs to the presumably monophyletic group Psoroptidia (OConnor 1982, 2009). The Psoroptidia is a very diverse group currently including over 3100 described species arranged in about 600 genera, and, depending on taxonomy concepts, in 48–50 families (OConnor 2009). In the recently proposed classification of acariform mites by Mironov and Bochkov (2009), Psoroptidia is treated as a parvorder. Most psoroptidian species are highly host-specific being mono- or oligoxenous symbiotes and the phenomenon of parallel evolution between these mites and their hosts is often clearly observable (Fain 1994). According to the generally accepted hypothesis of psoroptidian phylogeny (OConnor 1982, 2009), bird-associated (Pterolichoidea, Analgoidea) and mammal-associated symbiotes (Sarcoptoidea) constitute three major phylogenetic branches within the Psoroptidia. This traditional view of phylogenetic relationships within Psoroptidia has been detonated by the work of Klimov and OConnor (2008a) based on evidence from three nuclear genes. These authors clearly showed that the superfamily Sarcoptoidea in traditional sense (OConnor 2009) is diphyletic (Psoroptidae-Lobalgidae and remaining

sarcoptoids) and these two sarcoptoid branches are nested in different groups of analgooid mites (Analgoidea). The recent paper of Dabert et al. (2010) devoted to the molecular phylogeny of Acariformes roughly supports these data. Thus, the superfamily Sarcoptoidea *sensu* OConnor (2009) is not monophyletic and mammal associated psoroptidians belong to two different groups (superfamilies); they are tentatively named here as Sarcoptid and Psoroprid complexes. Our recent studies based on morphological characters correspond to this hypothesis (Bochkov and OConnor 2010; Bochkov and Mironov, submitted).

In this paper we provide the family level review of mammal associated psoroptidians, keys to all genera, and lists of their species along with verified host records. Some “sarcoptoid” subfamilies deserving the full familial rank are given here as subfamilies *incertae sedis* in the respective familial complexes. Their ranks are changed in the paper by Bochkov and Mironov (submitted).

MATERIAL AND METHODS

The materials used in this work were obtained from two main sources: acarological and mammal collections housed in various museums and from hosts captured in nature. Data about sarcoptoid

families and subfamilies including numbers of recognized species and genera, examined taxa, hosts, microhabitats, and distribution are summarized in Table 1. Idiosomal and leg setation of sarcoptoid families and subfamilies represented in Tables 2–4.

Most examined materials are deposited in the following collections: Institut royal des Sciences naturelles de Belgique, Brussels, Belgium (IRSNB); Musée royal de l’Afrique Centrale, Tervuren, Belgium (MRAC); Museum of Zoology, the University of Michigan, Ann Arbor, USA (UMICH); Zoological Institute, Russian Academy of Sciences, Saint-Petersburg, Russia (ZISP).

A general review of the external morphology of free-living and symbiotic Astigmata was recently given by Klimov and OConnor (2008b) and OConnor (2009). The idiosomal chaetotaxy follows Griffiths et al. (1990) with modifications of Norton (1998) for coxal-genital setae. The leg chaetotaxy follows Grandjean (1939). Host systematics follows the checklist of Wilson and Reeder (2005).

SYSTEMATICS

Key to families and subfamilies *incertae sedis* of mammal-associated Psoroptidia (both sexes)

1. Setae *hl* present (if absent — most dorsal hysteronotal setae also absent) Sarcoptid complex 4
 - Setae *hl* absent, most hysteronotal setae present ... Psoroptid complex 2
2. Dorso-apical projection on tarsi I and II absent or scarcely distinct 3
 - Dorso-apical projection on tarsi I and II distinctly developed Psoroptidae Canestrini, 1892
3. Apodemes Ia widely separated. Setae *sI*, II hook-like. Setae *vi* present. Females without opisthosomal lobes Paracoptinae Lavoipierre, 1955
 - Apodemes Ia fused into sternum. Setae *sI*, II filiform. Setae *vi* absent. Females with opisthosomal lobes Lobalgidae Fain, 1965
4. Most hysteronotal setae present. Cheliceral hood present. Genital papillae present (absent in most Sarcoptidae). Ectosymbiotes 8
 - Most hysteronotal setae absent. Cheliceral hood absent. Genital papillae absent. Endosymbiotes 5
5. Gnathosoma not hypertrophied, having typical structure for psoroptidians. Hypostome and palps not enlarged, palps rounded apically without eupathidia. Setae *se* present 6
 - Gnathosoma hypertrophied, distantly resembling gnathosoma of ixodids. Hypostome and palps enlarged, palps pointed apically, bearing eupathidia. Setae *se* absent Chirorhynchobiidae Fain, 1967
6. Idiosoma slightly or strongly elongated or even vermiform. Propodonotal shield without longitudinal apodeme. Opisthosoma distinctly developed. Trochanters and genua III and IV not joining normally. Famulus ϵ present 7
 - Idiosoma ovoid in outline. Propodonotal shield with longitudinal apodeme. Opisthosoma weakly developed and legs IV inserted close to posterior margin of opisthosoma. Trochanters and genua III and IV joining at right angles. Famulus ϵ absent. Pneumocoptidae Baker et al., 1958
7. Palpal segments fused dorsally. Gnathosoma without subcapitular setae. Movable cheliceral digit stylet-like, fixed digit reduced. Hysteronotal shield present. In both sexes, solenidion $\omega 3$ present Lemurnyssidae Fain, 1957
 - Palpal segments separated dorsally. Gnathosoma with subcapitular setae. Chelicerae chelate. Hysteronotal shield absent. Solenidion $\omega 3$ always absent in females and present or absent in males Gastronyssidae Fain, 1956
8. Tarsi I and II with ambulacral discs and without striated clasping membranes 9
 - Tarsi I and II without ambulacral discs and bearing striated clasping membranes Chirodiscidae Trouessart, 1892
9. Striated membranes or flaps absent between coxal fields I and II 11
 - Striated membranes or flaps present between coxal fields I and II 10
10. Propodonotal shield not covering entire gnathosoma dorsally. Palpal membranes indistinct. Setae *vi* and *cI* absent. Striated membranes between coxal fields I and II not forming distinct flaps. Tibiae and tarsi III and IV fused Atopomelidae Gunther, 1942
 - Propodonotal shield covering entire gnathosoma dorsally (in Aplodontochirinae it not covering palpal extremities). Palpal membranes distinctly developed. Setae *vi* and *cI* present. Striated membranes between coxal fields I and II forming distinct flaps. Tibiae and tarsi III and IV not fused Listrophoridae Megnin et Trouessart, 1884
11. Legs III of males and legs III and IV of females not modified into attachment organs 13
 - Legs III of males and legs IV of females modified into attachment organs 12
12. Setae *scx* absent. Setae *vi* and *cI* present. Setae

baI, II present, *sIII* absent. Femora of modified legs enlarged and ventrally concave; tibia and tarsi devoid of pretarsi and shortened. Claspings apparatus formed by all leg podomeres, excluding trochanters Myocoptidae Gunther, 1942 — Setae *scx* present. Setae *vi* and *cI* absent. Setae *baI*, II absent, *sIII* present. Femora of modified legs strongly reduced and inserted into respective trochanters; tibiae flattened; tarsi fold back over ventral surface of respective tibiae and also flattened, bearing distinct pretarsi, which strongly curved ventrally, flattened and armed with sclerotized crests. Claspings apparatus formed by tibia and tarsus Dromiciocoptinae Fain, 1970 13. Setae *cI* present. Setae *c2* layered. Hysteronotal shield in females present. Tarsi I and II with distinct retrorse projections. Setae *baI*, II, *laI*, II and *waI*, II present 14 — Setae *cI* absent. Setae *c2* not layered. Hysteronotal shield in females absent. Tarsi I and II with distinct retrorse projections. Setae *baI*, II, *laI*, II and *waI*, II absent ... Listropsoralginae Fain, 1965 14. Palpal apices without corolla of retrorse projections. Genital papillae absent (present in Diaboliocoptinae). Setae *ps3* absent. Male opisthosomal lobes and paranal suckers absent. Sarcoptidae Murray, 1877 — Palpal apices with corolla of retrorse projections. Setae *ps3* present. Genital papillae present. Male opisthosomal lobes and paranal suckers present. Rhyncoptidae Lawrence, 1956

Families of Sarcoptid complex

Diagnosis. *Both sexes:* Cheliceral hood and ventral apophysis of movable digit present or absent. Dorso-apical spur on tarsi I and II absent or developed in one sex. Setae *h1* present, if absent — many other dorsal idiosomal setae also lost. Intercostal striate membranes between coxal fields I and II present or absent. Genital papillae present or absent. Anal opening situated ventrally, terminally or dorsally. Anterior part of central sclerite of ambulacral discs of legs I and II (if present and discernible) narrowed anteriorly. Females oviparous or ovoviviparous. Males with or without opisthosomal lobes and paranal suckers

Family Sarcoptidae Murray, 1877

Type genus: *Sarcoptes* Latreille, 1802

Table 5, Figs. 1–8.

Diagnosis. *Both sexes.* Cheliceral hood and ventral apophysis of movable digit present. Supracoxal sclerite and supracoxal opening indistinct, setae *scx* absent. Genital papillae present only in

Diaboliocoptinae. Intercostal attaching organs absent. Setae *c2* layered. Legs without clasping organs but sometimes with ventral processes on some segments. Setae *baI*, II spine-like. Setae *ps3* absent, setae *h3* present only in some Teinocoptinae. Setae *dI* absent in Sarcoptinae and Teinocoptinae. Solenidion σ/III present only in Diaboliocoptinae, σ/III absent; famulus ε present only in *Satanicoptes*. Ambulacral sclerites small but recognizable.

Female. Idiosoma highly variable in shape, slightly flattened dorso-ventrally, globose or subcylindrical. Hysteronotal and opisthogastral shields absent. Ovipore transversal. Epigynum weakly developed or absent. Anal opening situated terminally, dorso-terminally or dorsally. Setae *c1* and *c2* layered in most sarcoptids but not layered in Teinocoptinae. Tibia and tarsi III and IV fused (free in Diaboliocoptinae). Ambulacral stalks on tarsi I and II present or absent and absent on tarsi III and IV of most species. Oviparous.

Male. Idiosoma slightly elongated or ovoid form, slightly flattened dorso-ventrally. Opisthosomal lobes and paranal suckers absent. Legs III and IV normally developed. Tibia and tarsi III fused (free in Diaboliocoptinae). Setae *dIV* and *eIV* (if present) sucker-like.

Taxa included: 117 species and 15 genera in 3 subfamilies, Sarcoptinae Murray, 1877 — *Sarcoptes* Latreille, 1802 (1 species), *Prosarcoptes* Lavoipierre, 1960 (3 species), *Kutzerocoptes* Lavoipierre, 1970 (1 species), *Trixacarus* Sellnick, 1944 (3 species); Teinocoptinae Fain, 1959 — *Teinocoptes* Rodhain, 1923 (20 species), *Chirnyssoides* Fain, 1959 (8 species), *Chirobia* Fain, 1959 (12 species), *Chirophagoides* Fain, 1963 (1 species), *Cynopterocoptes* Klompen, 1992 (1 species), *Notoedres* Railliet, 1893 (45 species), *Nycteridocoptes* Oudemans, 1898 (15 species), *Rousettocoptes* Klompen, 1992 (1 species), *Tychosarcoptes* Fain, 1976 (3 species); Diaboliocoptinae Fain et Domrow, 1974 — *Diaboliocoptes* Fain et Domrow, 1974 (1 species), *Satanicoptes* Fain et Lawrence, 1975 (2 species).

Associations with hosts: inhabitants of the superficial skin layers of therian mammals, Sarcoptinae* — Primates (Hominidae, Cebidae, Cercopithecidae) and Rodentia (Caviidae, Cricetidae, Muridae, Nesomyidae); Teinocoptinae — Placentalia: Chiroptera (Emballonuridae, Molossidae,

* *Sarcoptes scabiei* (Linnaeus, 1758), being initially hominid-associated parasite, secondarily switched to representatives of more than 16 families belonging to ten orders of therian mammals (Fain 1968a; Klompen 1992; Domrow 1992).

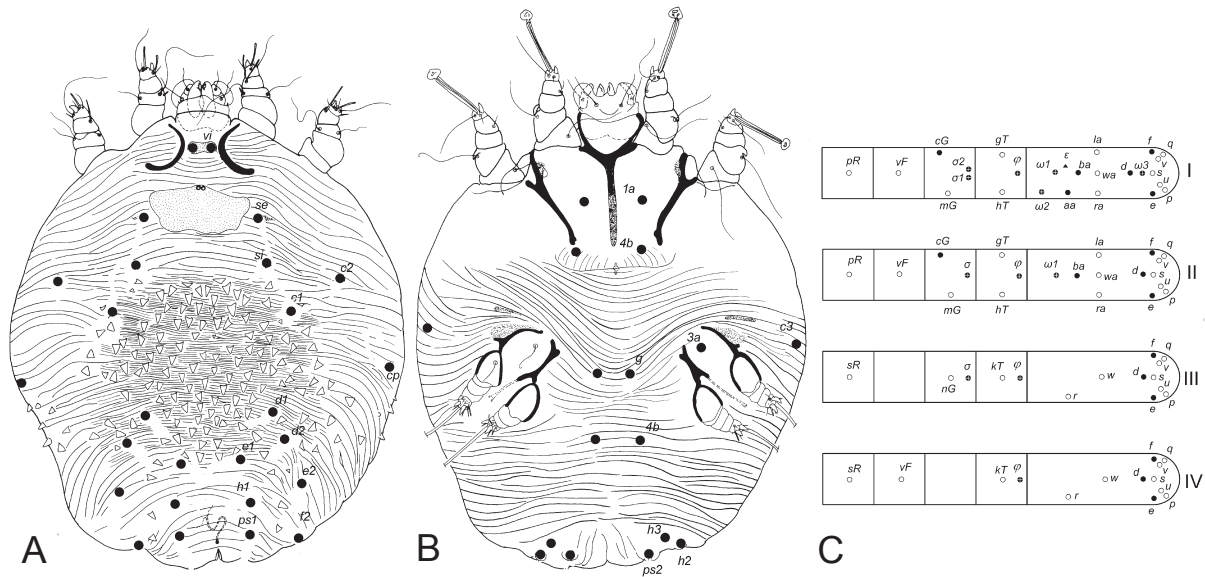


Fig. 1. Sarcoptidae, scheme of setation: A — idiosoma in dorsal view, B — same in ventral view, C — legs I-IV.

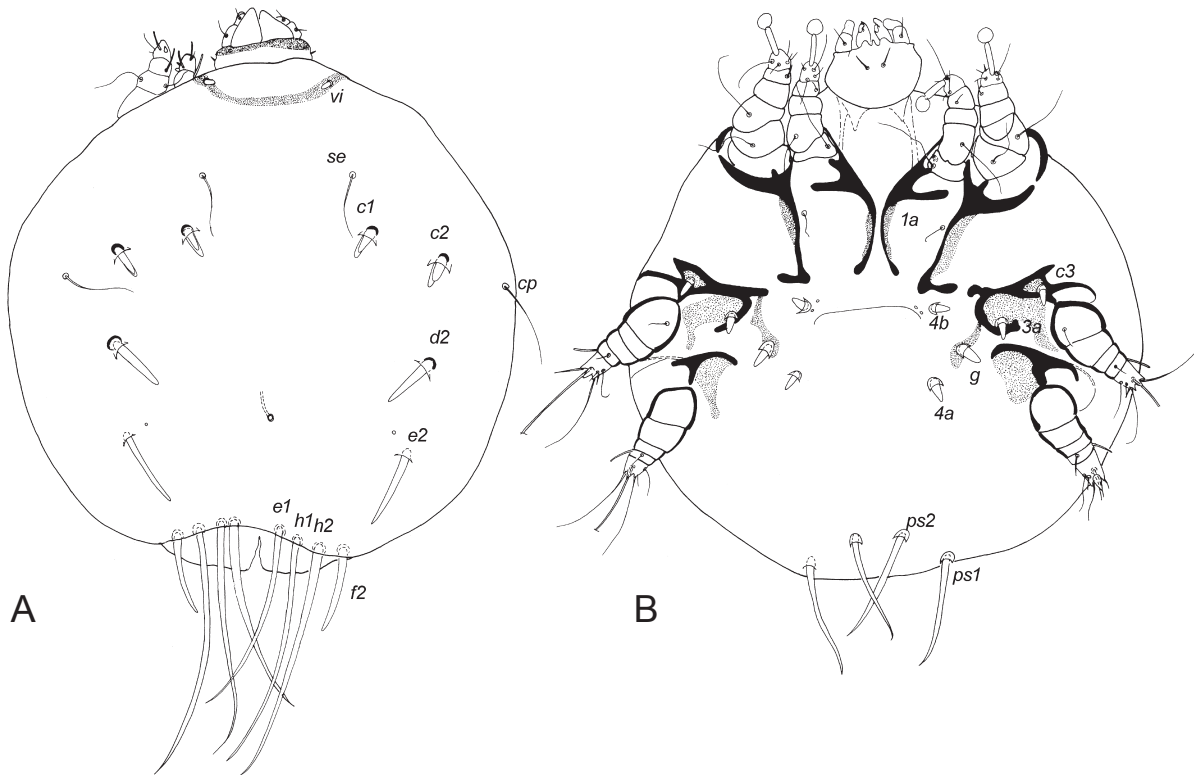


Fig. 2. *Diablicoptes sarcophilus* Fain et Domrow, 1974, female: A — dorsal view, B — ventral view.

Mystacinidae, Noctilionidae, Phyllostomidae, Pteropodidae, Rhinolophidae, Vespertilionidae), Erinaceomorpha (Erinaceidae), Primates (Galagidae), Carnivora (Canidae, Felidae, Herpestidae, Procionidae), Lagomorpha (Leporidae), Rodentia (Cricetidae, Muridae, Sciuridae), Soricomorpha (Soricidae); Marsupialia: Peramelemorpha (Perameliidae), Phalangeriformes (Phalangeridae) Vombatiformes (Phascolarctidae); Diablicoptinae — Dasyuomorpha (Dasyuridae).

Mites of the subfamily Teinocoptinae are primarily associated with chiropteran hosts and only secondarily associated (*Notoedres* spp.) with hosts belonging to other orders (Klompen 1992).

Distribution. Sarcoptinae and Teinocoptinae — cosmopolites; Diablicoptinae — Australia.

Main references. Fain (1968a) — revision of Sarcoptidae (old conception), Klompen (1992) — revision and phylogeny of Sarcoptidae (new conception), Fain and Domrow (1974) — description

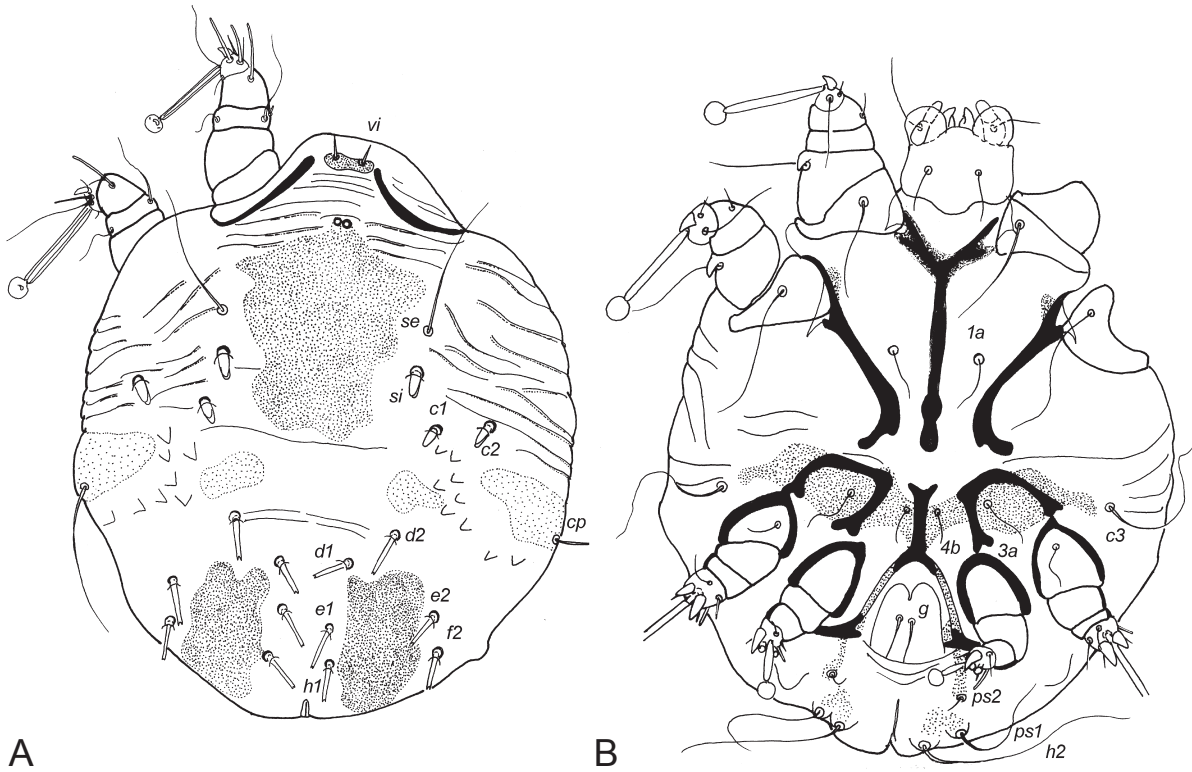


Fig. 3. *Sarcoptes scabiei* (Linnaeus, 1758), female: A — dorsal view, B — ventral view.

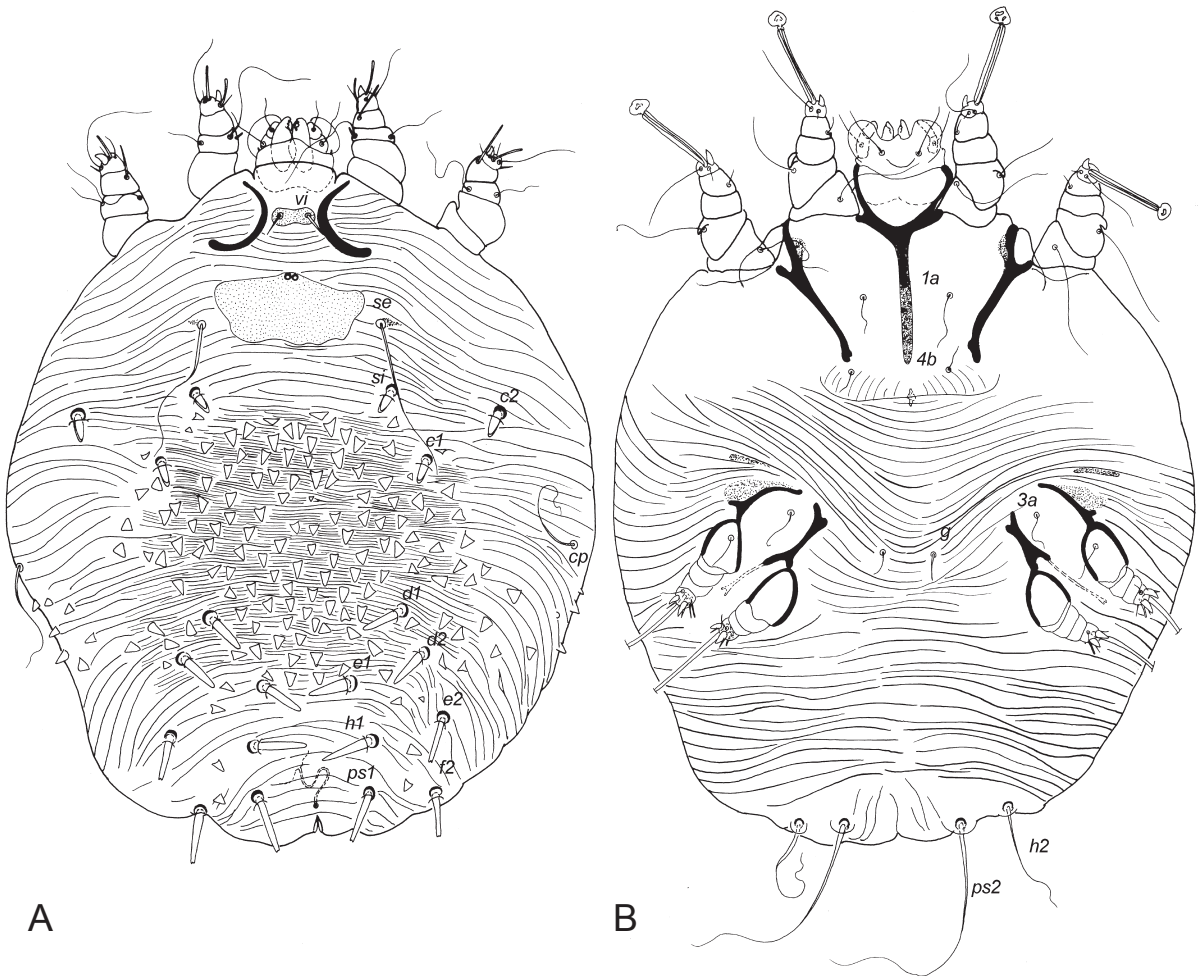


Fig. 4. *Sarcoptes scabiei* (Linnaeus, 1758), male: A — dorsal view, B — ventral view.

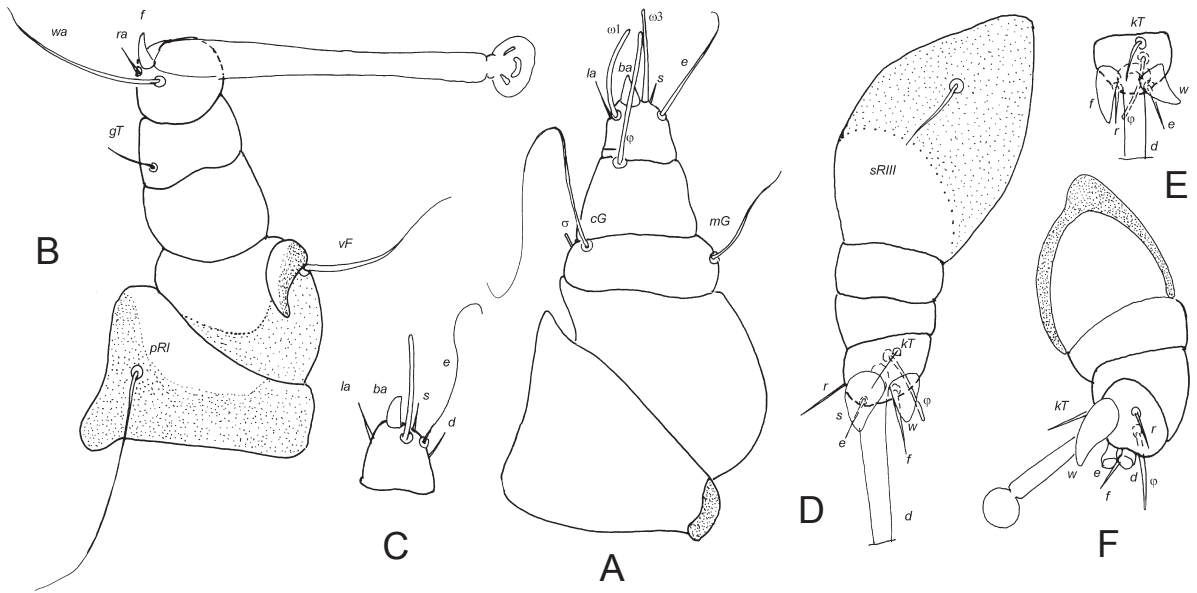


Fig. 5. *Sarcoptes scabiei* (Linnaeus, 1758), female legs (A–E): A — leg I in dorsal view, B — same in ventral view, C — tarsus II in dorsal view, D — leg III in ventral view, E — tibiotarsus IV in ventral view; F — male leg IV in ventral view.

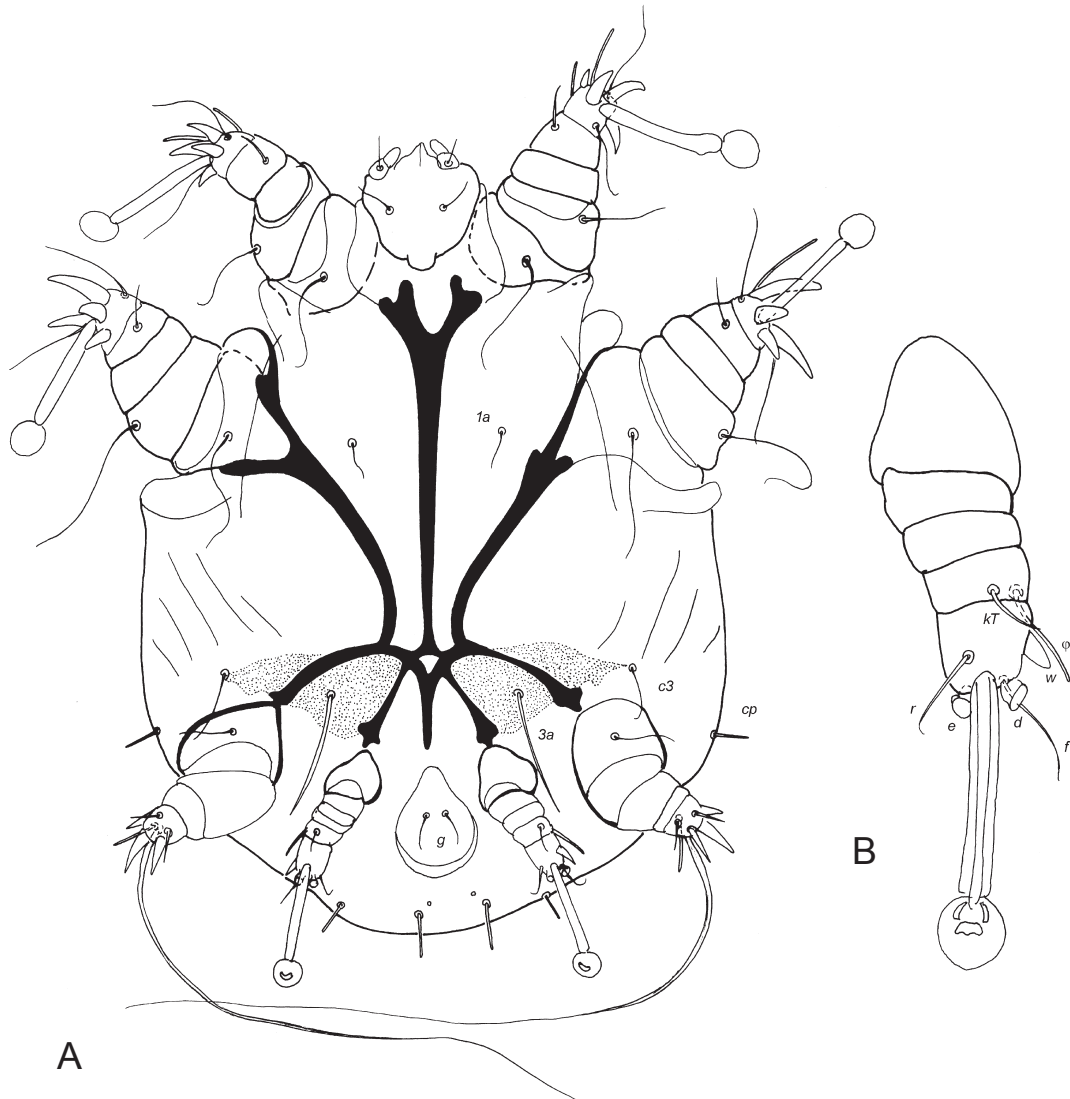


Fig. 6. *Notoedres pahangi* Klompen et al., 1983, female: A — dorsal view, B — ventral view, C — leg I in ventral view, D — tibiotarsus III in ventral view.

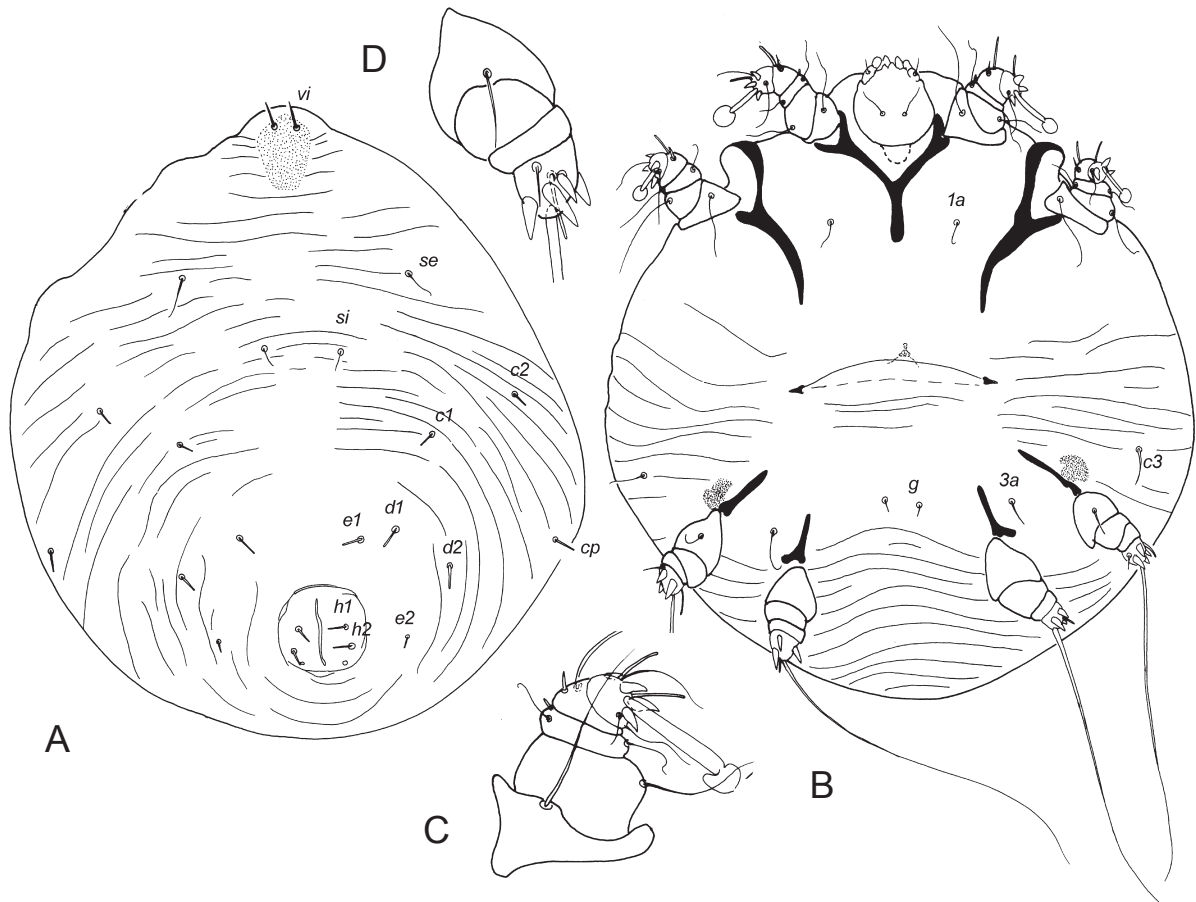


Fig. 7. *Notoedres alexfaini* Lavoipierre, 1968, male: A — ventral view, B — leg IV in ventral view, C — leg I in ventral view, D — leg III in ventral view.

of *Diablicoptes*, Fain and Laurence (1975) — description of *Satanicoptes*; OConnor (1982) — diagnoses of Sarcoptidae and references.

**Key to genera of the family Sarcoptidae
Murray, 1877**

(based on Klompen [1992])

1. *Both sexes*: Setae *si* and *dI* present, *dI* and σ III absent. *Female*: tibia and tarsi III fused. *Male*: Propodonal shield moderately developed 3
— *Both sexes*: Setae *si* and *dI* absent, *dI* and σ III present. *Female*: tibia and tarsi III free. *Male*: Propodonal shield reduced to narrow transverse band *Diablicoptinae* Fain et Domrow, 1974 2
2. *Both sexes*: Coxal apodemes Ia parallel to each other. Bifurcate spines on trochanter II absent. Tarsi I with incomplete set of setae (*laI* and famulus ϵ absent) *Diablicoptes* Fain et Domrow, 1974 — *Both sexes*: Coxal apodemes Ia fused to each other. Bifurcate spine on trochanter II present. Tarsi I with complete set of setae (*laI* and famulus ϵ present) *Satanicoptes* Fain et Laurence, 1975
3. *Both sexes*: Setae *sI*, II spur-like; setae *dII*, *laI*, II, and *eIV* absent. *Male*: Tibia and tarsus IV fused

or free. Setae *eIII* absent *Teinocoptinae* Fain, 1959 7
— *Both sexes*: Setae *sI*, II filiform; setae *dII*, *laI*, II, and *eIV* present. *Male*: Tibia and tarsus IV fused. Setae *eIII* present *Sarcoptinae* Murray, 1877 4
4. *Both sexes*: Setae *si*, *cI*, and *c2* thickened, much shorter than 20 μ m. Setae *4a* absent 5
— *Both sexes*: Setae *si*, *cI*, and *c2* filiform, longer than 20 μ m. Setae *4a* present *Trixacarus* Sellnick, 1944
5. *Female*: Propodonal shield present (sometimes strongly reduced). *Male*: Ventral median apodeme (situated between coxal fields III and IV) fused with genital sclerites 6
— *Female*: Propodonal shield absent. *Male*: Ventral median apodeme not fused with genital sclerites *Kutzerocoptes* Lavoipierre, 1970
6. *Female*: Posterior ventral tubercles of opisthosoma poorly developed. *Male*: Hysteronotal shield paired *Sarcoptes* Latreille, 1802
— *Female*: Posterior ventral tubercles of opisthosoma distinctly developed. *Male*: Hysteronotal shield unpaired *Prosarcoptes* Lavoipierre, 1960

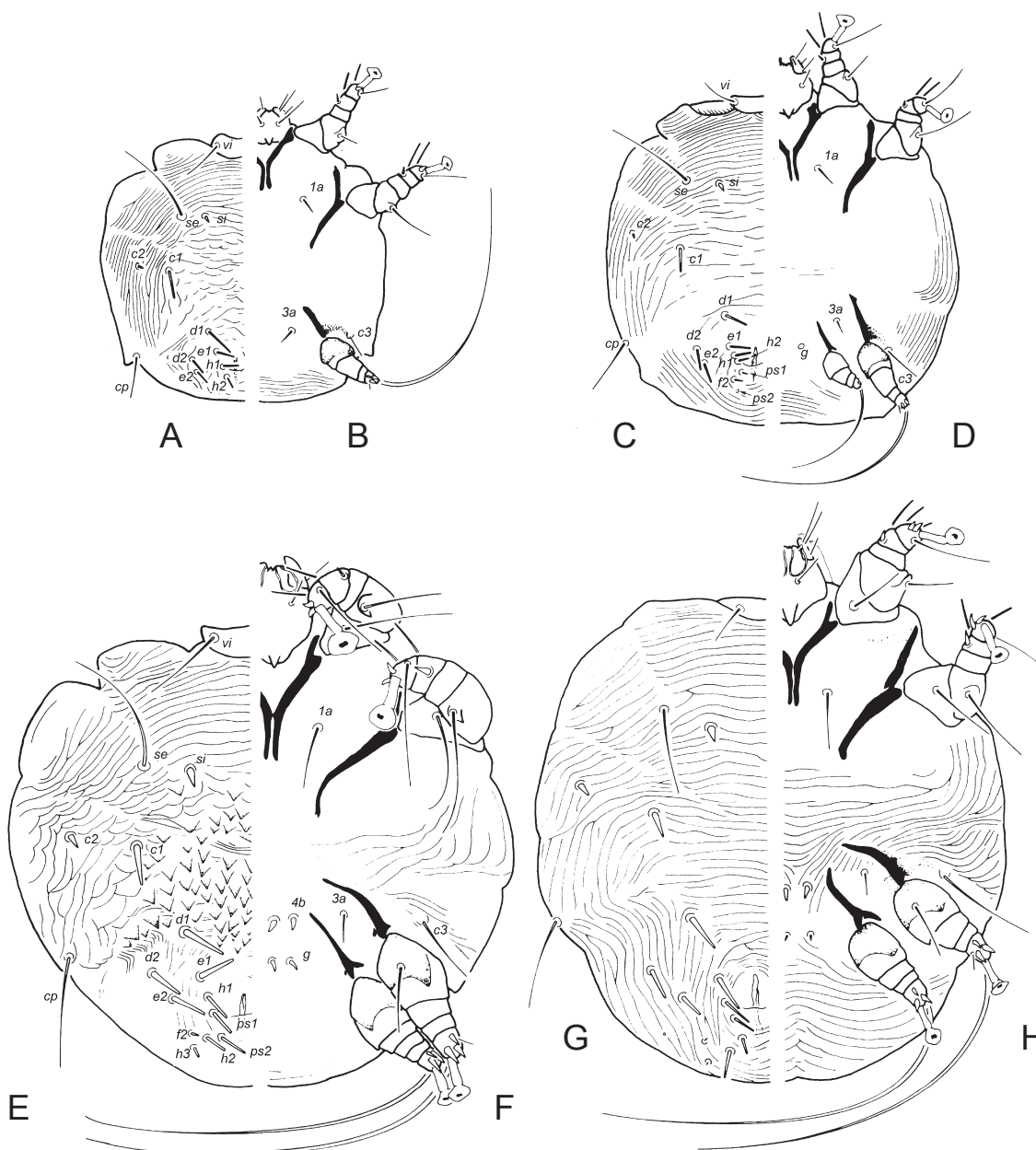


Fig. 8. *Nycteridocoptes heidemanni* Klompen, 1992, immature instars: A — larva in dorsal view, B — same in ventral view, C — protonymph in dorsal view, D — same in ventral view, E — female tritonymph in dorsal view, F — same in ventral view, G — male tritonymph in dorsal view, H — same in ventral view (after Klompen (1992), with minor modifications).

7. *Both sexes*: Setae *si*, *c1*, and *c2* thin spines or filiform. Setae *pRI*, II present or absent, filiform or simple spine-like. *Female*: Propodonotal shield absent 8
 — *Both sexes*: Setae *si*, *c1*, and *c2* broad spines. Setae *pRI*, II present, with bulbous proximal and filiform distal parts. *Female*: Propodonotal shield present *Chirophagoides* Fain, 1963
 8. *Female*: ambulacral discs of legs I and II absent. *Male*: Setae *4a* and *4b* present 10
 — *Female*: ambulacral discs of legs I and II present. *Male*: Setae *4a* and *4b* absent 9
 9. *Both sexes*: Setae *ps1* and *ps2* present. *Male*:

Aedeagus elongate *Chirnyssoides* Fain, 1959
 — *Both sexes*: Setae *ps1* and *ps2* absent. *Male*: Aedeagus short *Notoedres* Railliet, 1893
 10. *Female*: Unpaired dorsal expansion (tube) absent. *Male*: ambulacra absent 11
 — *Female*: Unpaired dorsal expansion (tube) present, weakly developed in a few species. *Male*: ambulacra present
 *Nycteridocoptes* Oudemans, 1898
 11. *Both sexes*: Setae *f2* and *h3* absent. *Female*: Legs IV consisting of 1–2 articulated segments ...
 12
 — *Both sexes*: Setae *f2* and *h3* present. *Female*:

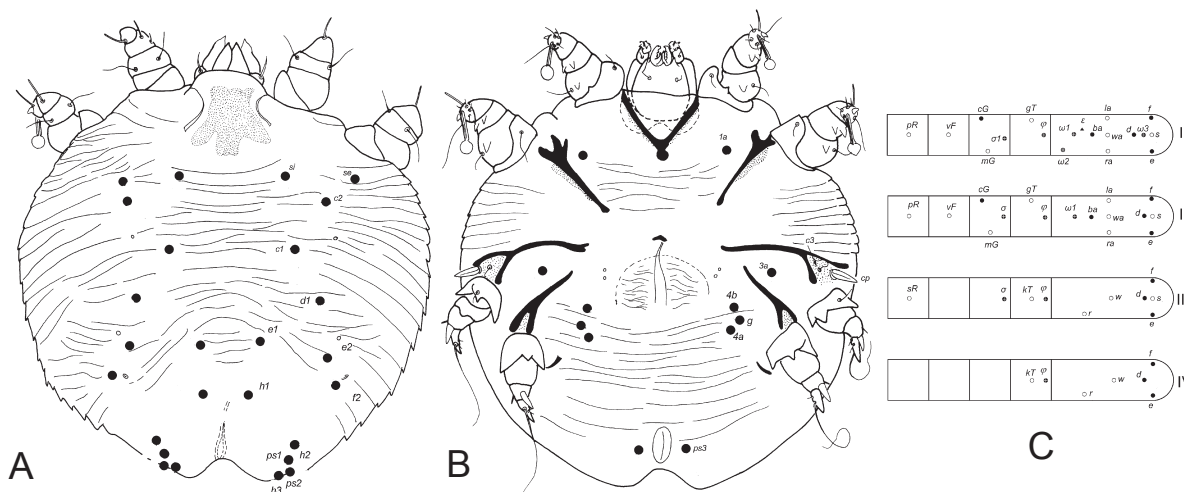


Fig. 9. Rhyncoptidae, scheme of setation: A — idiosoma in dorsal view, B — same in ventral view, C — legs I–IV.

Legs IV consisting of 4 articulated segments
 *Cynoptercoptes* Klompen, 1992
 12. *Female*: Setae *c1* shorter than 5 μm. Trochanter III without internal spur 13
 — *Female*: Setae *c1* longer than 5 μm. Trochanter III with distinct internal spur
 *Rousettocoptes* Klompen, 1992
 13. *Female*: Most posterior idiosomal spines in lateral spine group small. *Male*: Coxal field II with posterior retrorse spurs 14
 — *Female*: Most posterior idiosomal spines in lateral spine group very large (longer than 10 μm). *Male*: Coxal field II without posterior retrorse spurs *Tychosarcoptes* Fain, 1976
 14. *Female* (vigorous): Body distinctly longer than wide. Legs III with 4 articulated segments, femur and genu III separated. Legs IV present, sometimes strongly reduced. *Male*: In most species, apodemes IIIa and IVa fused to each other by their distal apices *Teinocoptes* Rodhain 1923
 — *Female* (vigorous): Body length and width subequal. Legs III with 3 articulated segments, femur and genu III fused. Legs IV absent. *Male*: Apodemes IIIa and IVa not fused
 *Chirobia* Fain, 1959

Family Rhyncoptidae Lawrence, 1956

Type genus: *Rhyncoptes* Lawrence, 1956
 Table 6, Figs. 9–17.

Diagnosis. *Both sexes.* Cheliceral hood and ventral apophysis of movable digit present. Apices of palpal tibiae with corolla of retrorse processes. Setae *dT* of palpal tarsi absent. Idiosoma slightly elongated or ovoid in outlines. Supracoxal sclerite and setae *scx* absent, supracoxal opening indistinct. Genital papillae present. Intercostal at-

taching organs absent. Setae *d1* and *c2* layered, setae *vi* and *d2* absent. Abambulacra on legs III and IV absent. Legs without clasping organs but with ventral processes on some segments. Femora and genua of legs III and IV fused. Setae *baI*, II spine-like; setae *kTIII*, IV with deep root each (without root in *Rhyncoptes*). Setae *sIII*, *eIII*, IV and *fIII*, IV absent. Solenidion σII –III absent. Ambulacral sclerites small but recognizable.

Female. Hysteronotal and opisthogastral shields absent. Ovipore in shape of inverted Y or transversal. Epigynum weakly developed or absent. Bursa copulatrix with thickened walls. Anal opening situated ventrally or ventro-terminally. Ambulacral stalks on tarsi I and II strongly elongated. Oviparous.

Male. Opisthosomal lobes and paranal suckers present. Legs III and IV normally developed (reduced in *Rhyncoptes*). Setae *dIV* setiform or stick-like, setae *eIV* absent.

Taxa included: 12 species and 4 genera, *Audycoptes* Lavoipierre, 1964 (2 species), *Caenolestocoptes* Fain et Lukoschus, 1976 (1 species), *Rhyncoptes* Lawrence, 1956 (5 species), *Saimiriocoptes* Fain, 1968 (2 species), *Ursicoptes* Fain et Johnston, 1970 (2 species).

Associations with hosts: inhabitants of hair follicles of therian mammals, Marsupialia: Paucituberculata (Caenolestidae); Placentalia: Carnivora (Ursidae, Procionidae), Primates (Cebidae, Cercopithecidae), and Rodentia (Hystricidae).

The parasitism of *Rhyncoptes recurvidens* Lawrence, 1956 on *Hystrix africae australis* Peter, 1852 (Hystricidae) is probable a consequence of host shift from cebids, because other four species of the genus are associated with primates of the

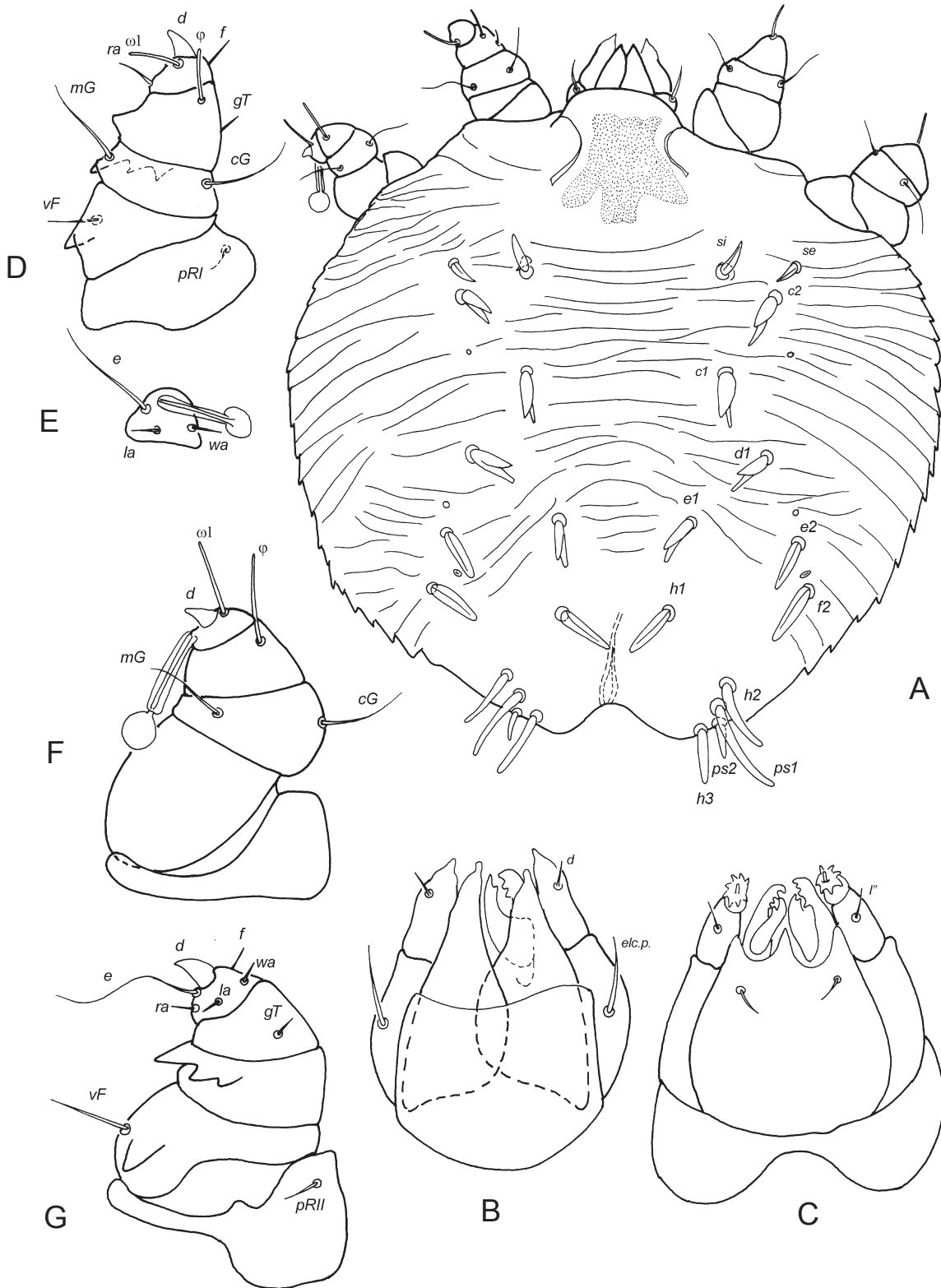


Fig. 10. *Caenolestocoptes inca* Fain et Lukoschus, 1976, female: A — dorsal view, B — gnathosoma in dorsal view, C — same in ventral view, D — leg I in dorsal view, E — tarsus I in ventral view, F — leg II in dorsal view, G — same in ventral view.

family Cebidae and other representatives of the family are not associated with rodents.

Distribution. Eurasia, Africa (not recorded from Madagascar), North and South America.

Main references. Fain (1965a) — revision of *Rhyncoptes*, Fain and Lukoschus (1976) — description of *Caenolestocoptes*, OConnor (1982, 1987) — diagnoses of Rhyncoptidae and Audy-

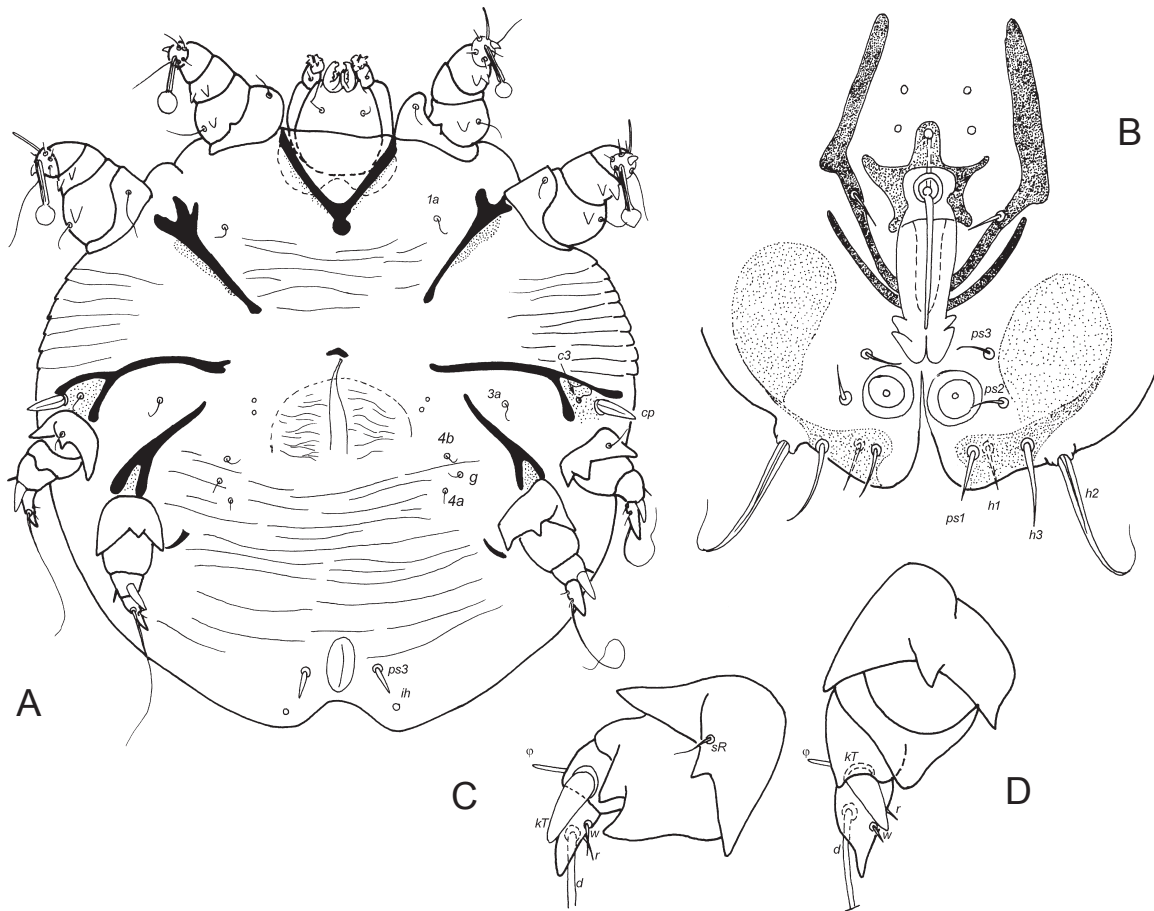


Fig. 11. *Caenolestocoptes inca* Fain et Lukoschus, 1976: A — female in ventral view, B — male opisthosoma in ventral view, C — leg III of female in ventral view, D — same, leg IV.

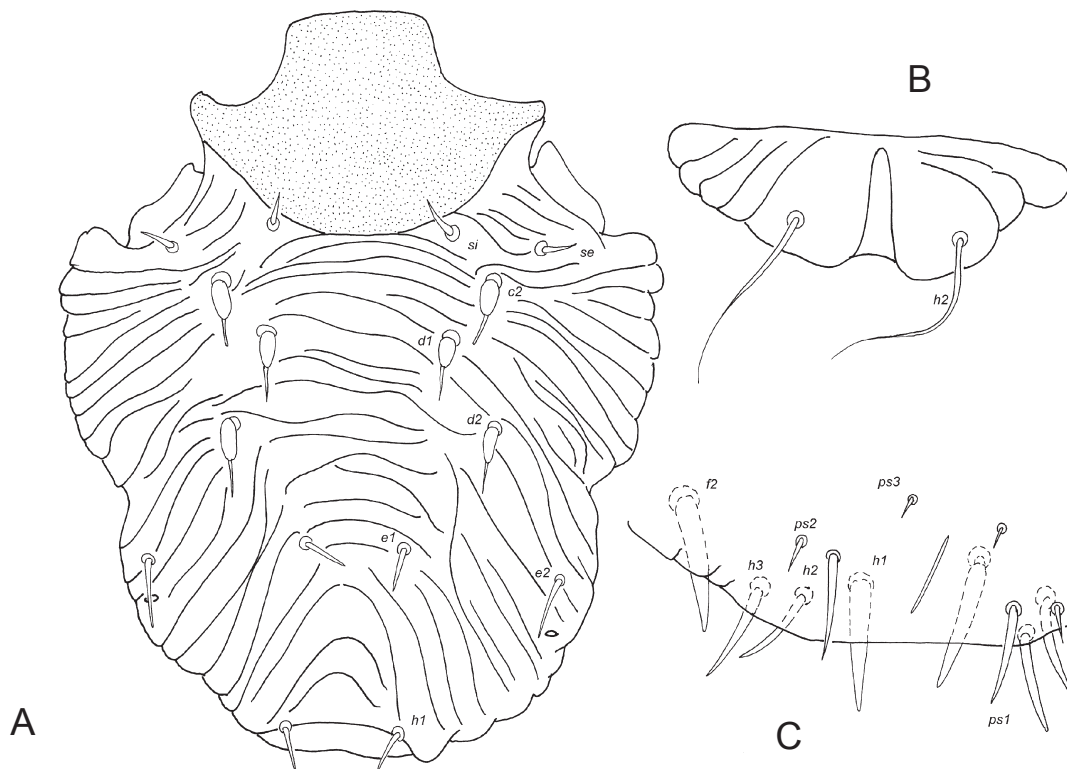


Fig. 12. *Caenolestocoptes inca* Fain et Lukoschus, 1976, immature instars: A — larva in dorsal view, B — opisthosoma of larva in ventral view, C — opisthosoma of protonymph in ventral view.

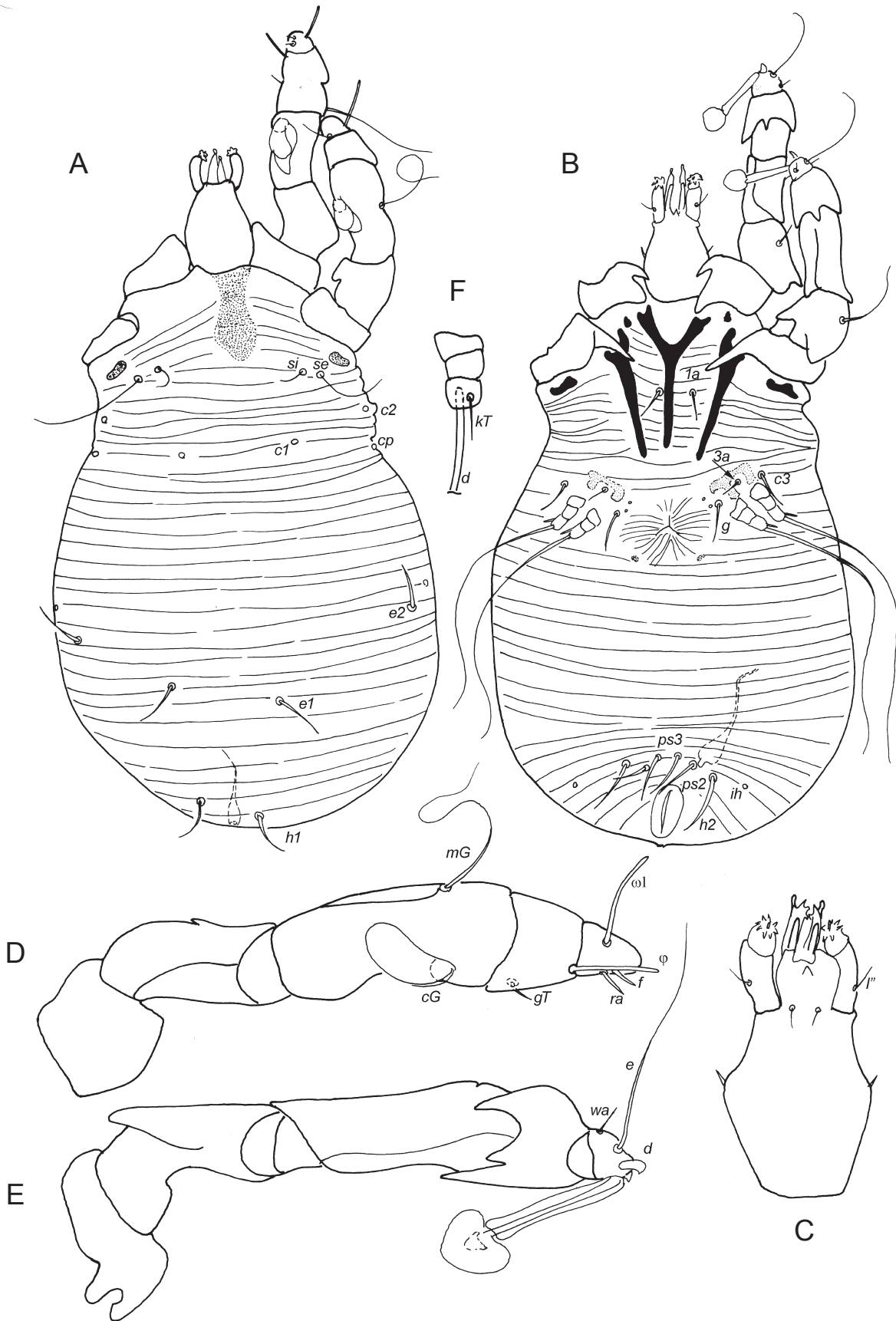


Fig. 13. *Rhyncoptes grabberi* Klompen, 1989, female: A — dorsal view, B — ventral view, C — gnathosoma in ventral view, D — leg I in dorsal view, E — leg I in ventral view, F — leg III in ventral view.

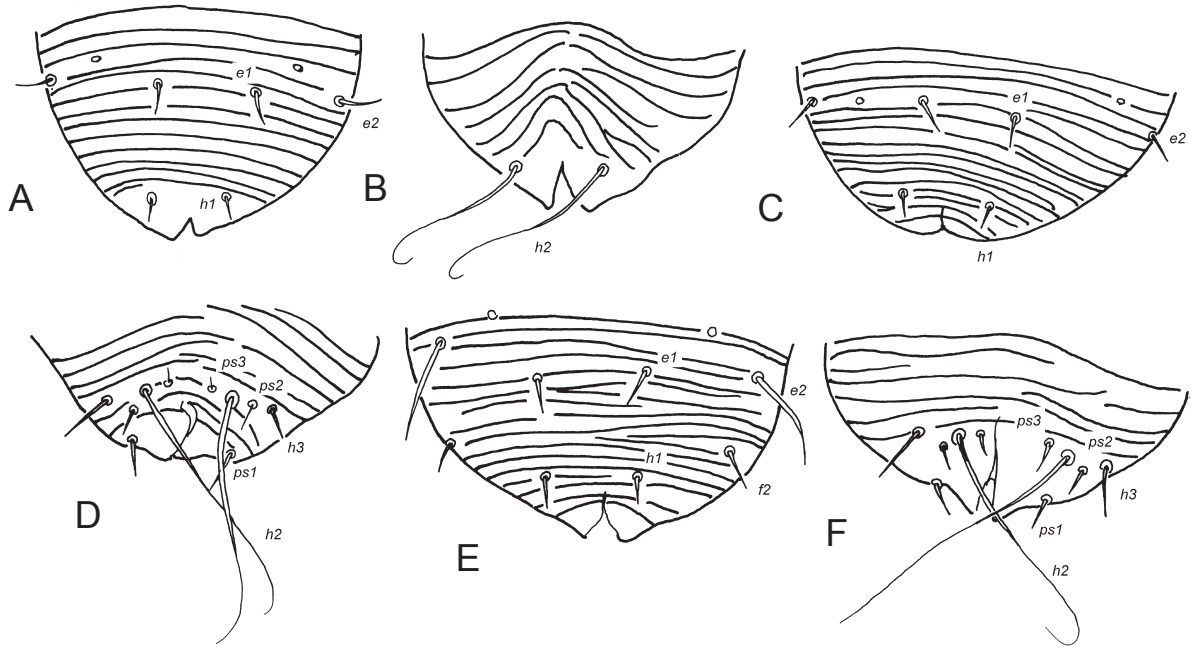


Fig. 14. *Rhyncoptes grabberi* Klompen, 1989, opisthosoma of immature instars: A — larva in dorsal view, B — same in ventral view, C — protonymph in dorsal view, D — same in ventral view, E — tritonymph in dorsal view, F — same in ventral view.

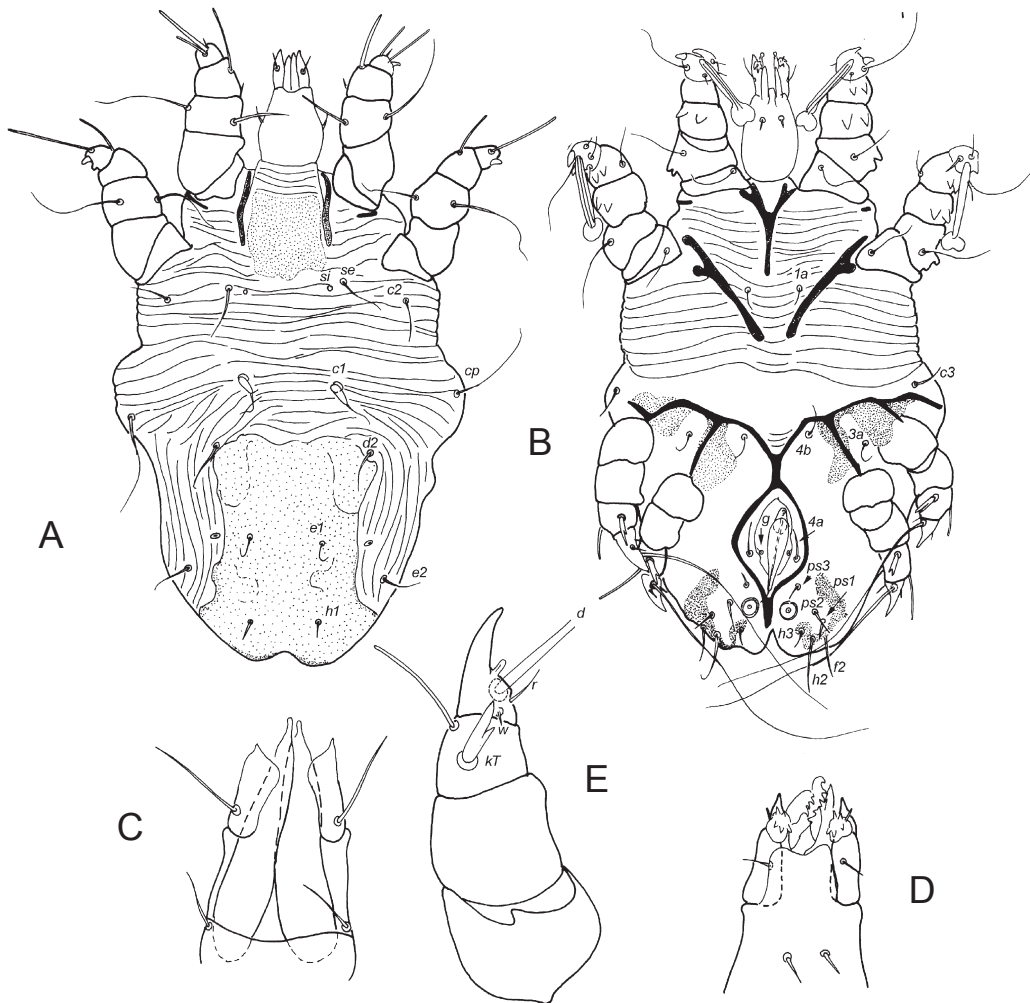


Fig. 15. *Uрсicoptes procioni* Fain et Wilson, 1979, male: A — dorsal view, B — ventral view, C — gnathosoma in dorsal view, D — same in ventral view, E — leg III in ventral view.

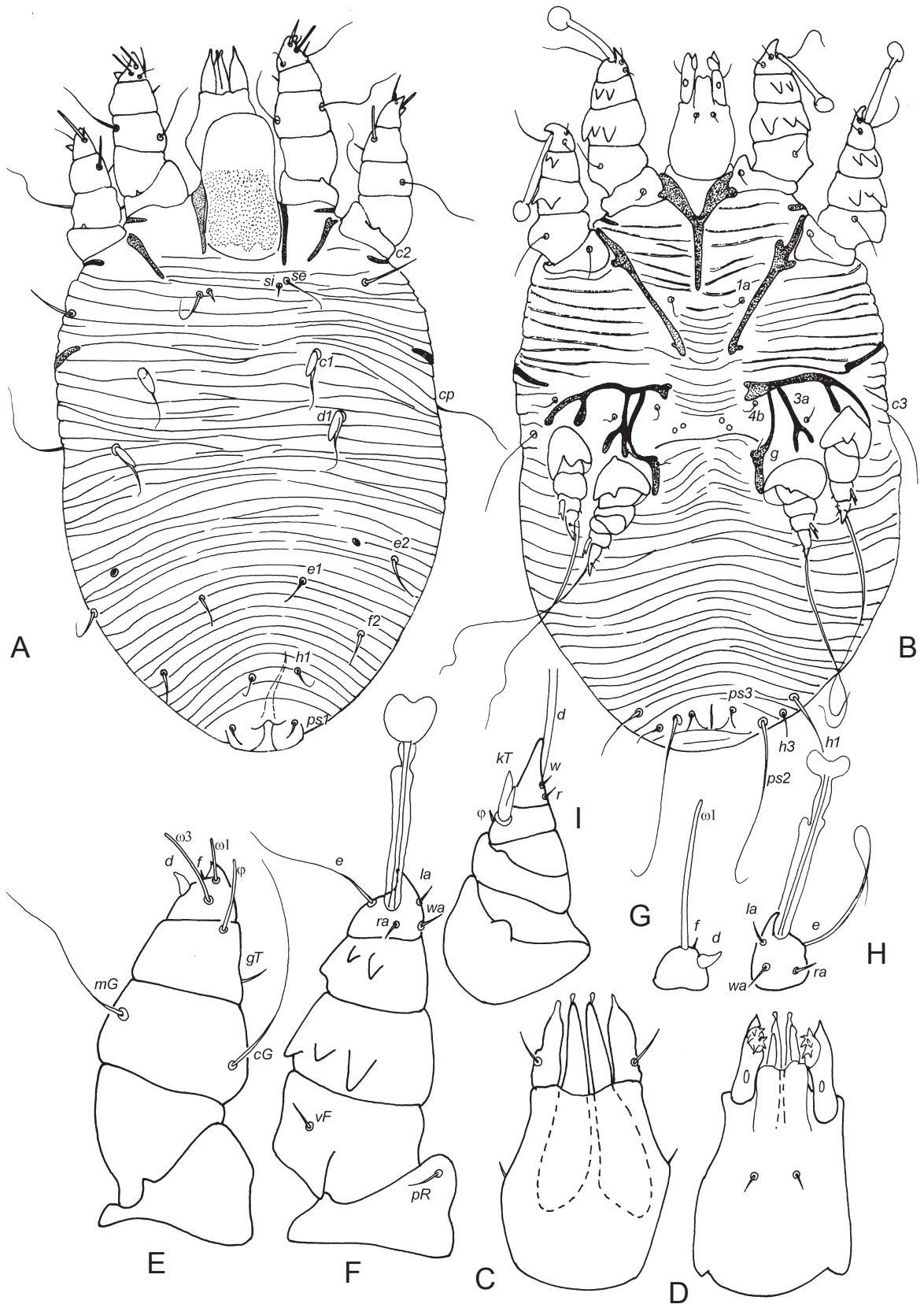


Fig. 16. *Ursicoptes procioni* Fain et Wilson, 1979, female: A — dorsal view, B — ventral view, C — gnathosoma in dorsal view, D — same in ventral view, E — leg I in dorsal view, F — leg I in ventral view, G — tarsus II in dorsal view, H — tarsus II in ventral view, I — tarsus III in ventral view.

coptidae (synonym of Rhyncoptidae), references, and morphology of *Audycytes*, Klompen (1989,

1993) — ontogeny of *Rhyncoptes* and phylogeny of Rhyncoptidae.

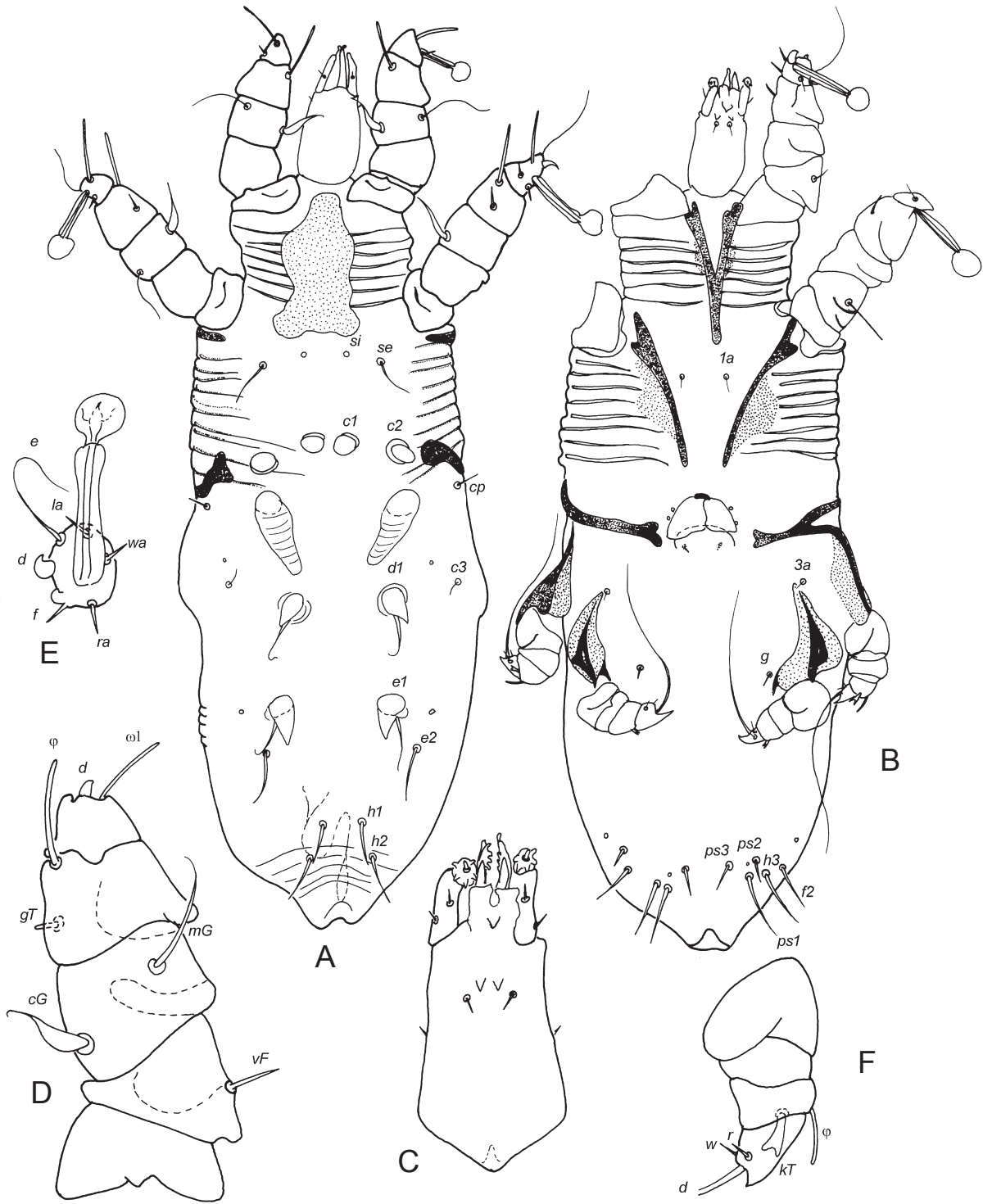


Fig. 17. *Saimirioptes herskovitzi* OConnor, 1987, female: A — dorsal view, B — ventral view, C — gnathosoma in ventral view, D — leg I in dorsal view, E — tarsus I in ventral view, F — leg III in dorsal view.

**Key to genera of the family Rhyncoptidae
Lawrence, 1956**

1. *Both sexes*: Idiosoma longer than wide. Subcapitulum elongated. Setae 4a absent. Setae si filiform. Setae cp filiform or represent by alveoli. Solenidion $\omega 3$ present. *Female*: Setae sRIII absent 2

— *Both sexes*: Length and width of idiosoma subequal. Subcapitulum rounded. Setae 4a present. Setae si and cp spine-like. Solenidion $\omega 3$ absent. *Female*: Setae sRIII present *Caenolestocptes* Fain et Lukoschus, 1976

2. *Both sexes*: Ventral ridges or spines on legs I–II present. Setae pRI–II absent. Setae cGI–II with

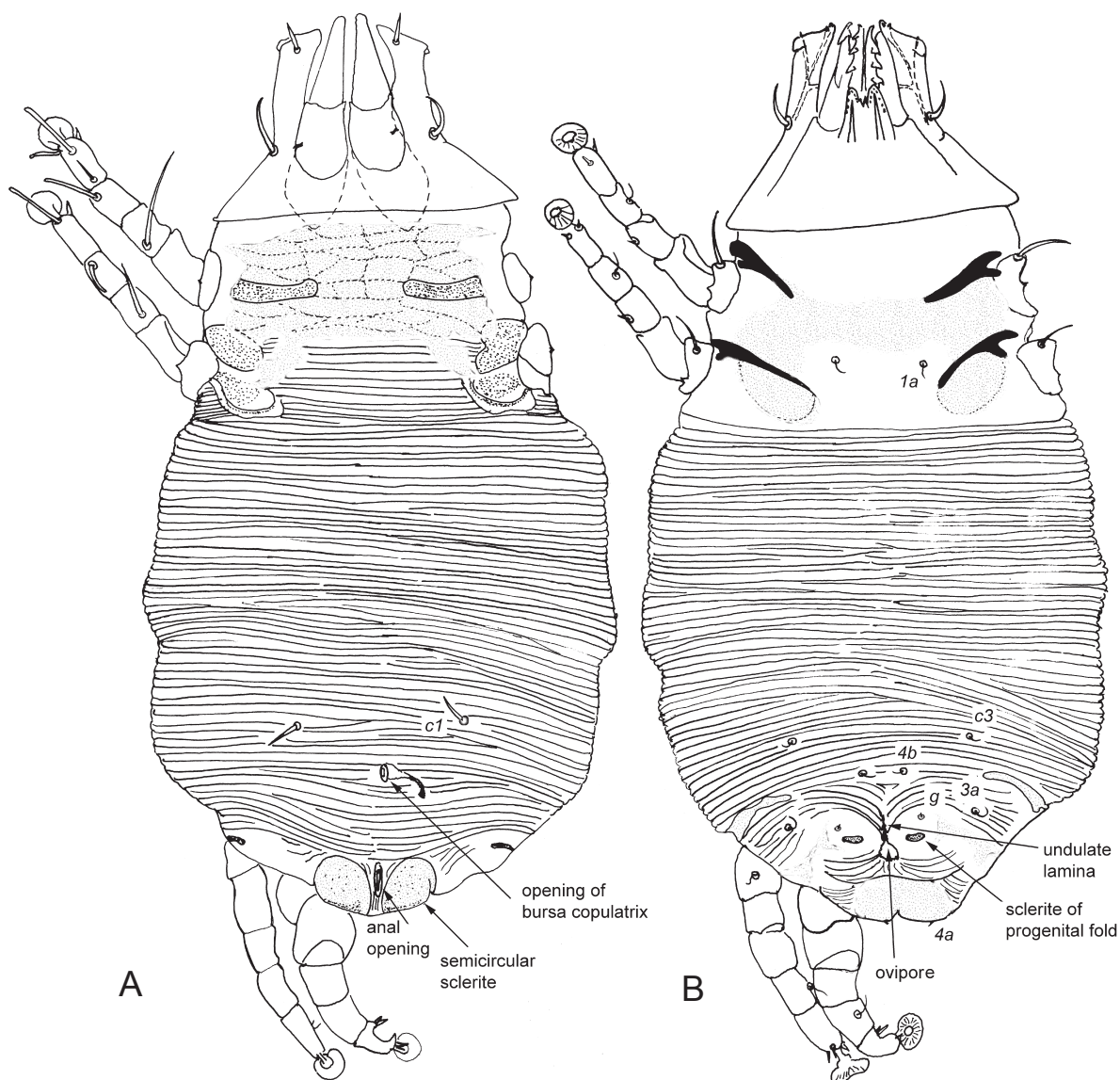


Fig. 18. *Chirorhynchobia matsoni* Yunker, 1970, female: A — dorsal view, B — ventral view (from Bochkov et al. 2008 with minor modifications).

strongly inflated base. *Female*: Genital opening triangular. Setae *c2* layered. Setae *3a* and ϕ IV absent 3
 — *Both sexes*: Ventral ridges or spines on legs I–II absent. Setae *pRI*–II present. Setae *cGI*–II without inflated base. *Female*: Genital opening transverse. Setae *c2* not layered. Setae *3a* and ϕ IV present *Ursicoptes* Fain et Johnston, 1970
 3. *Both sexes*: Setae *cp* represented by alveoli or absent. *Female*: Dorsal cuticular lobes absent ... 4
 — *Both sexes*: Setae *cp* setiform. *Female*: Dorsal cuticular lobes present ... *Saimirioptes* Fain, 1968
 4. *Both sexes*: Setae *l''* of palpal tibia and solenidion ϕ III absent. *Female*: Setae *c1*, *c2*, and *d1* represented by alveoli or absent. Tarsus and tibia III and IV fused *Rhyncoptes* Lawrence, 1956

— *Both sexes*: Setae *l''* of palpal tibia and solenidion ϕ III present. *Female*: Setae *c1*, *c2*, and *d1* setiform. Tarsus and tibia III and IV not fused *Audycoptes* Lavoipierre, 1964

Family Chirorhynchobiidae Fain, 1968

Type genus: *Chirorhynchobia* Fain, 1968

Table 7, Figs. 18–20

Diagnosis. *Both sexes*. Cheliceral hood and ventral apophysis of movable digit not observable. Palps distinctly enlarged, longer than hypostome. Palpal podomeres completely fused to each other and basal podomere (tibia) fused with subcapitulum, bearing full set of setae (eupathidia and solenidion very short). Subcapitulum with trapezoid shape without ventral subcapitular setae *subc*. Id-

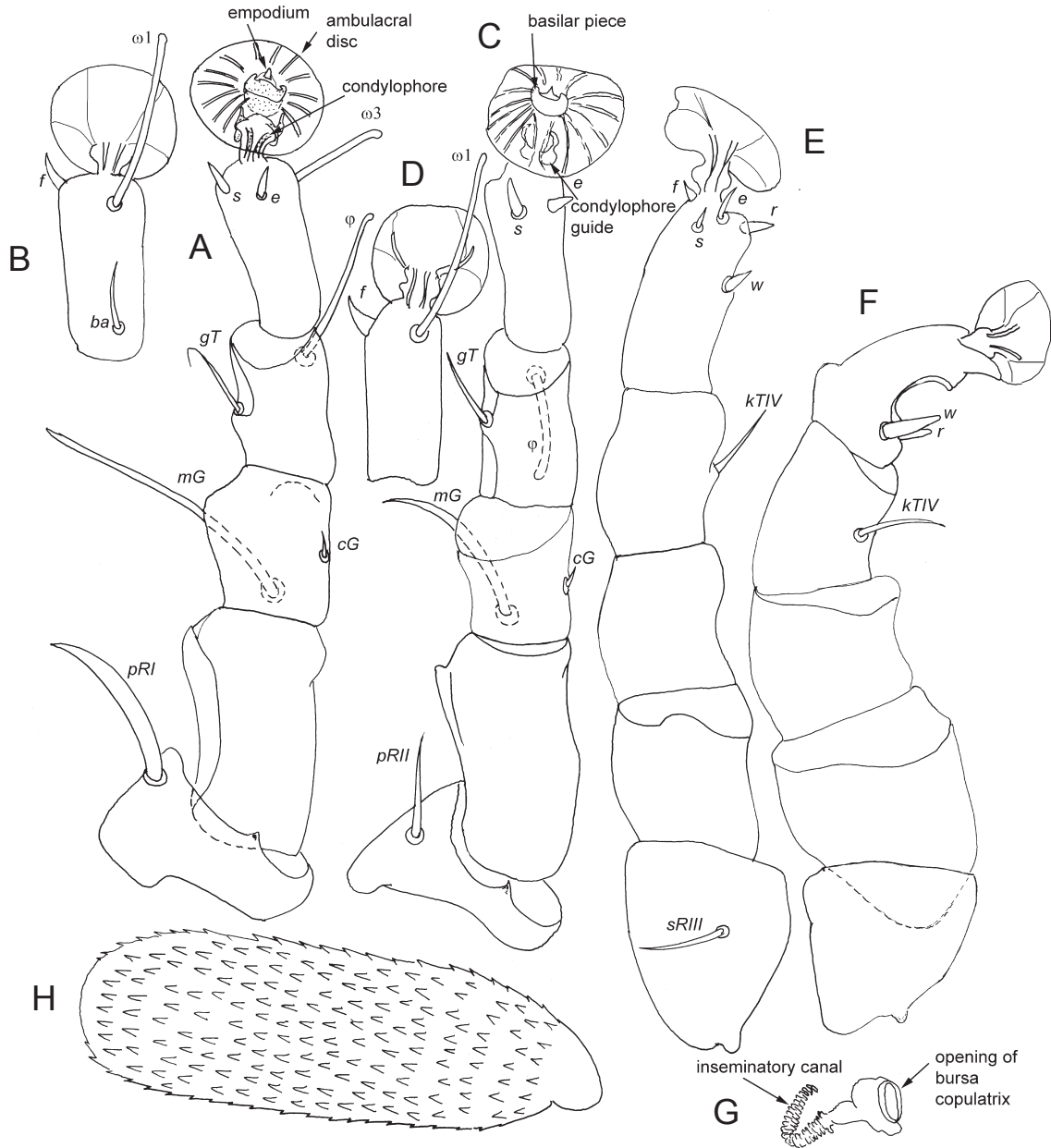


Fig. 19. *Chiorhynchobia matsoni* Yunker, 1970, details of female: A — leg I in ventral view, B — tarsus I in dorsal view, C — leg II in ventral view, D — tarsus II in dorsal view, E — leg III in ventral view, F — leg IV in ventral view, G — bursa copulatrix, H — egg (from Bochkov et al. 2008 with minor modifications).

iosoma slightly elongated. Supracoxal sclerite and setae *scx* absent, supracoxal opening indistinct. Hysteronotal and opisthogastral shields absent. Space between coxal fields I–II and III–IV greatly enlarged. Opisthosoma strongly reduced, and legs IV situated almost at posterior end of idiosoma. Intercoxal membranes or flaps absent. Genital papillae absent. Idiosomal and leg setation incomplete. Ambulacral sclerites distinctly developed. Solenidia ω I–II situated in apical part of respective tarsi.

Female. One pair of semicircular sclerites situated on posterior end of female opisthosoma. Anal opening situated dorsally, posterior to opening of bursa copulatrix. Vulvar lobes absent. Ovi-

pore in shape of inverted Y. Epigynum absent. Tarsi IV distinctly curved. Oviparous. Chorion of egg completely covered with short denticles.

Male. Setae *4a* are displaced dorsally. Opisthosomal lobes and paranal suckers absent. Setae *eIV* setiform or stick-like.

Taxa included: type genus with 3 species.

Associations with hosts: inhabitants of wing membranes of chiropteran hosts belonging to the family Phyllostomidae (Chiroptera).

Distribution. South America.

Main references. Fain (1968b) — original diagnosis of the family, Yunker (1970) — description of *C. matsoni* Yunker, 1970, Bochkov et al.

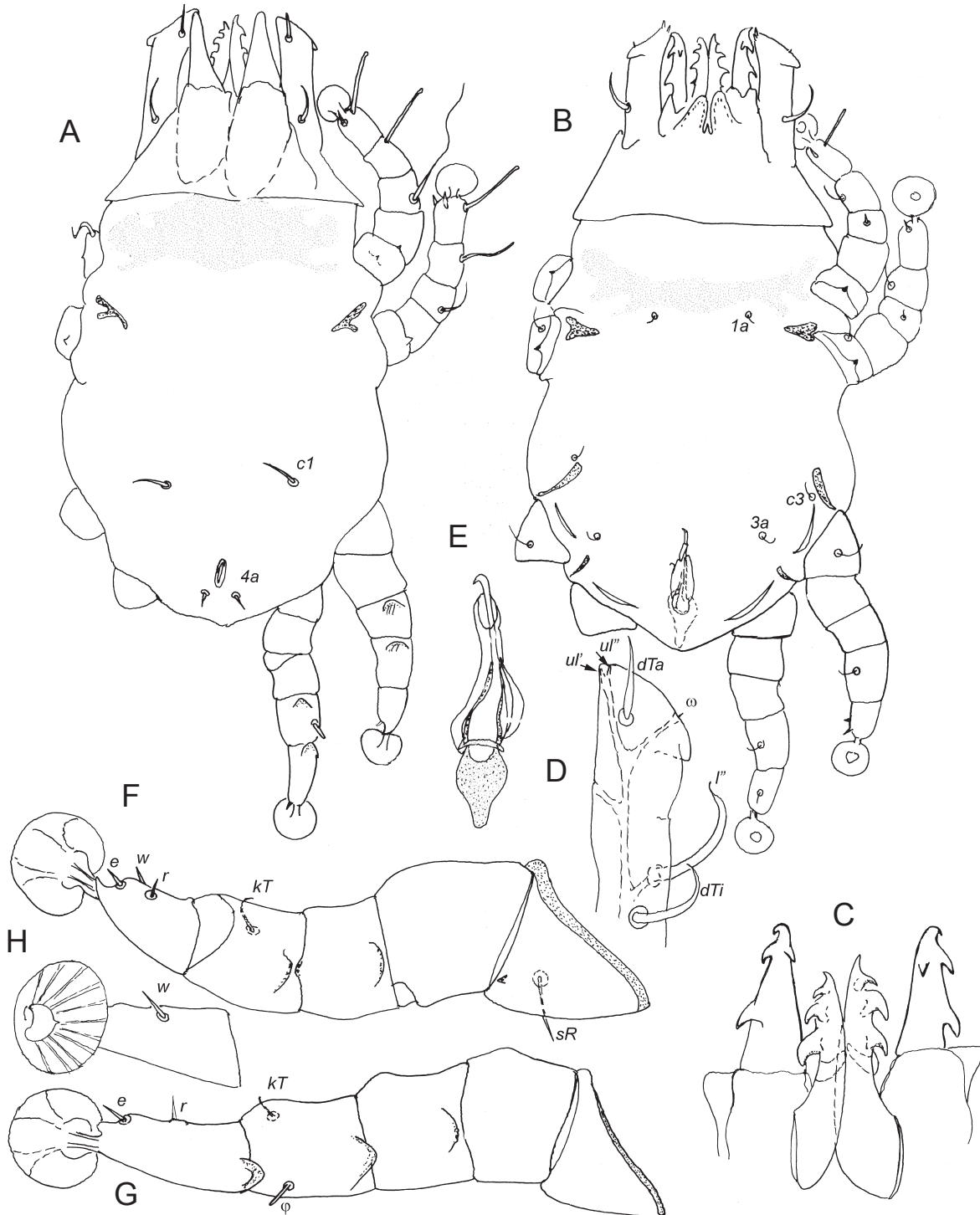


Fig. 20. *Chirorhynchobia glossophaga* Bochkov et al., 2008, male: A — dorsal view, B — ventral view, C — chelicerae in ventral view, D — palp in dorsal view, E — aedeagus, F — leg III in dorsal view, G — leg IV in dorsal view, H — tarsus IV in ventral view (from Bochkov et al. 2008 with minor modifications).

(2008a) — external morphology of chirorhynchobiids.

Family Atopomelidae Lawrence, 1956

Type genus: *Atopomelus* Lawrence, 1956

Table 8, Figs. 21–30

Diagnosis. Both sexes. Cheliceral hood and ventral apophysis of movable digit present. Mem-

branes of palpal tarsi weakly developed. Idiosoma elongated, subcylindrical or dorso-ventrally flattened. Supracoxal sclerite present; setae *scx* present or absent, supracoxal opening distinct. Propodonal shield subdivided onto prescapular and postscapular shields or postscapular shield absent. Opisthonotal shield present or absent. Genital papillae present. Intercoxal membranes dis-

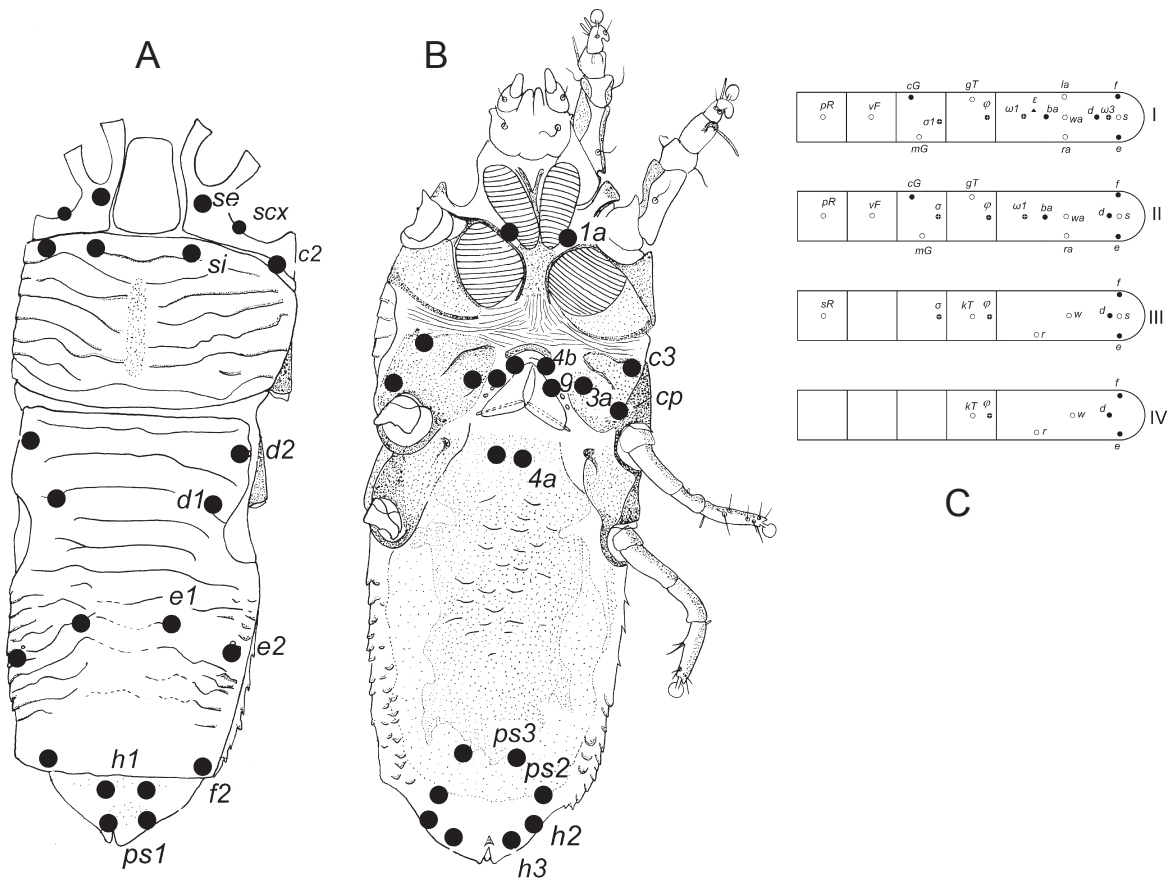


Fig. 21. Atopomelidae, scheme of setation: A — idiosoma in dorsal view, B — same in ventral view, C — legs I-IV.

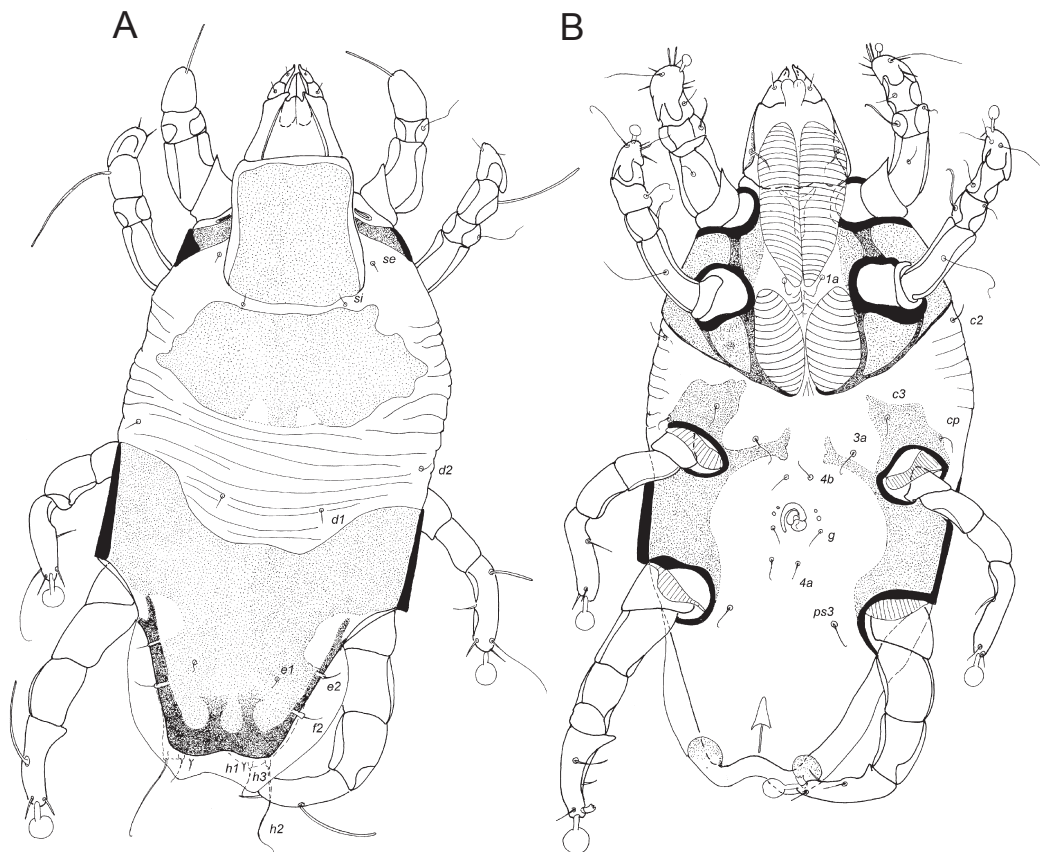


Fig. 22. *Tenrecobia tenrec* Fain, 1970, male: A — dorsal view, B — ventral view.

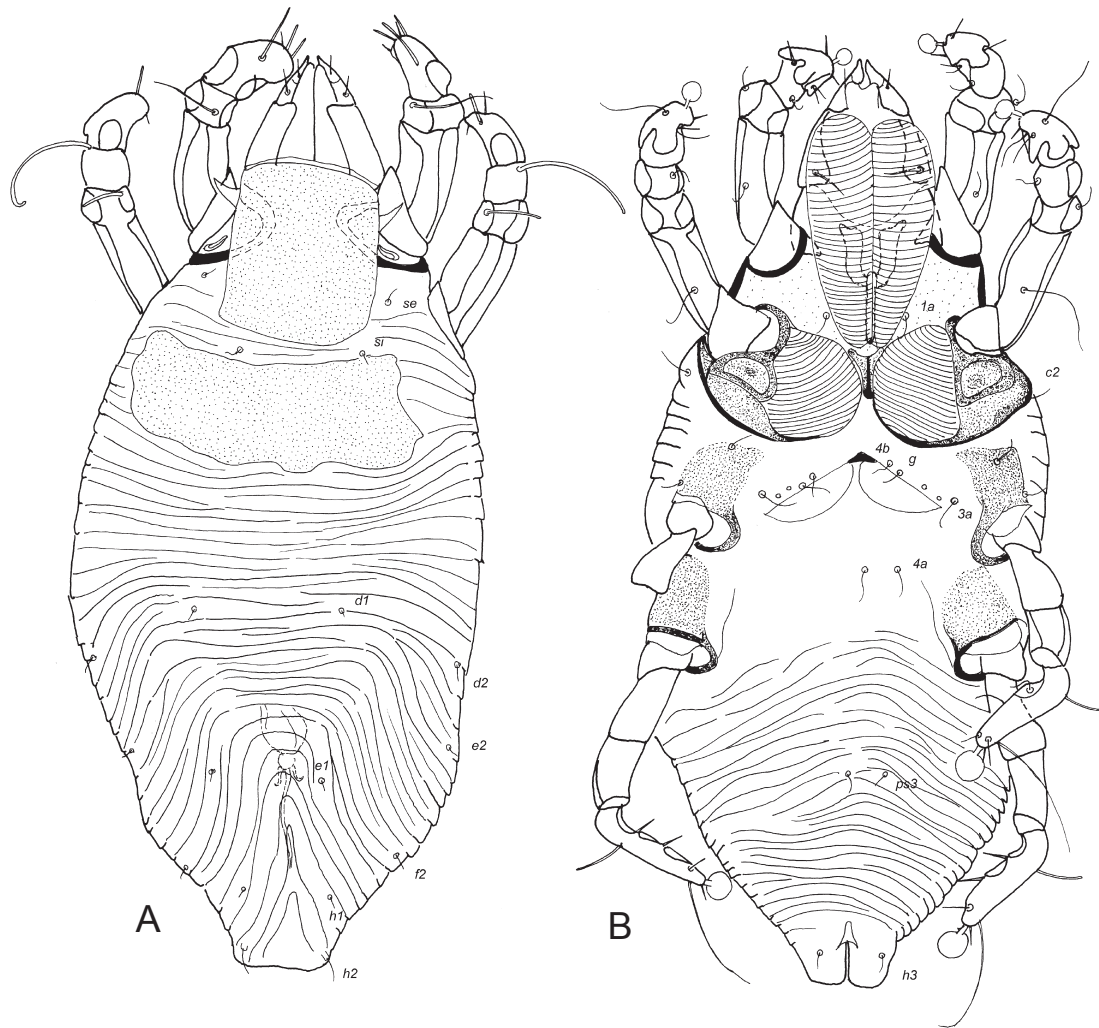


Fig. 23. *Tenrecobia tenrec* Fain, 1970, female: A — dorsal view, B — ventral view.

tinctly developed between coxal fields I and II or even occupying coxal field surfaces but not forming curved flaps. Idiosomal setae *vi* and *c1* absent; setae *ps1* and *ps2* present or absent. Tibia and tarsi of legs I and II partially fused. Tibia and tarsi III and IV completely fused. In many taxa, legs I and II with different expanding projections. Setae *s* and *laI*, II and genual solenidion σ II absent in most species. Ambulacral sclerites small but recognizable in tarsi I–II and distinctly developed in tarsi III and IV.

Female. Ovipore in shape of inverted Y. Epigynum distinctly developed or reduced. Anal opening situated ventrally or ventro-terminally. Oviparous.

Male. Opisthosomal lobes and paranal suckers present or absent. Legs III and IV normally developed. Setae *dIV* setiform or stick-like.

Taxa included: 419 species and 46 genera, *Atellana* Domrow, 1968 (1 species), *Atopomelus* Trouessart, 1918 (5 species), *Atopomelopsis* Fain,

1972 stat. res. (1 species), *Austrobisus* Fain, 1971 (1 species), *Austrochirus* Womersley, 1943 (9 species), *Bathyergolichus* Fain, 1970 (4 species), *Caenolestolichus* Fain et Bochkov, 2003 (1 species), *Campylochiropsis* Fain, 1972 (3 species), *Campylochirus* Trouessart, 1893 (8 species), *Capromylichus* Fain, 1970 (1 species), *Capromysia* Fain, 1979 (1 species), *Centetesia* Lawrence, 1966 (2 species), *Chirodiscoides* Hirst, 1917 (11 species), *Cubanochirus* Fain, 1970 (1 species), *Cytostethum* Domrow, 1956 (38 species), *Dasyurochirus* Fain, 1970 (15 species), *Didelphilichus* Fain, 1970 (2 species), *Didelphoecius* Fain, 1970 (20 species), *Distoechurobia* Fain, 1972 (1 species), *Domingoecius* Fain et Lukoschus, 1979 (1 species), *Dromiciolichus* Fain, 1970 (1 species), *Euryzygomysia* Fain, 1979 (1 species), *Isothricola* Fain, 1970 (5 species), *Koalachirus* Fain, 1970 (1 species), *Labidopygus* Fain et Domrow, 1973 (2 species), *Lemuroptes* Lawrence, 1958 (5 species), *Listrocarpus* Fain, 1967 (11 species), *Listrophor-*

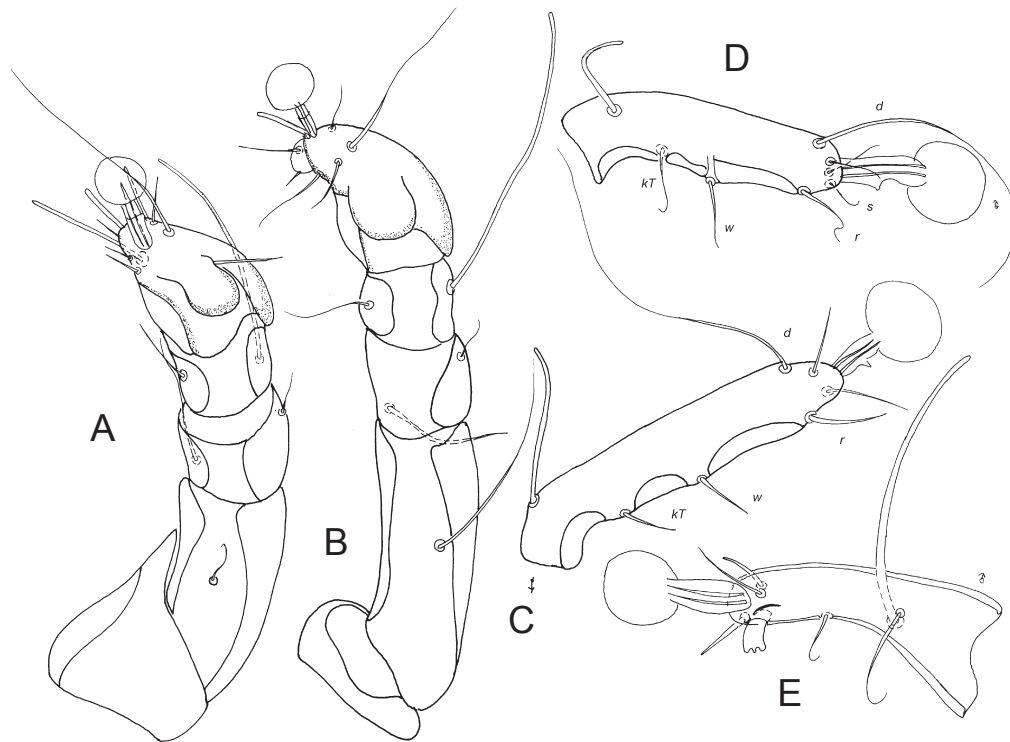


Fig. 24. *Tenrecobia tenrec* Fain, 1970, legs: A — female leg I in ventral view, B — female leg II in ventral view, C — female tarsus IV in dorsal view, D — male tarsus III in dorsal view, E — male tarsus IV in ventral view.

oides Hirst, 1923 (206 species), *Metachiroecius* Fain, Zanatta-Coutinho et Fonseca, 1996 (1 species), *Metadidelphoecius* Fain, 1979 (1 species), *Micropotamogalichus* Fain, 1970 (1 species), *Murichirus* Fain, 1971 (26 species), *Myocastorobia* Fain, 1975 (1 species), *Neolabidocarpus* Gunther, 1940 (1 species), *Notoryctobia* Fain et Lukoschus, 1981 (1 species), *Oryzomysia* Fain, 1970 (11 species), *Petaurobia* Fain, 1971 (3 species), *Petrogalochirus* Fain, 1970 (3 species), *Plagiodontochirus* Fain, 1976 (1 species), *Phalangerobia* Fain, 1972 (1 species), *Procytostethum* Fain, 1971 (1 species), *Prodidelphoecius* Fain, 1976 (1 species), *Sclerochiroides* Fain, 1970 (1 species), *Sclerochiroopsis* Fain, 1972 (1 species), *Scolonoticus* Fain, 1971 (4 species), *Tenrecobia* Lawrence, 1955 (2 species).

Associations with hosts: inhabitants of fur of therian mammals, Marsupialia: Dasyuromorphia (Dasyuridae, Myrmecobiidae), Didelphimorphia (Didelphidae), Diprotodontia (Acrobatidae, Hypsiprymnodontidae, Macropodidae, Petauridae, Phalangeridae, Phascolarctidae, Potoridae, Pseudocheiridae), Microbiotheria (Microbiotheriidae), Notoryctomorphia (Notoryctidae), Paucituberculata (Caenolestidae), Peramelimorphia (Peramelidae, Thylacomyidae); Placentalia: Afrosoricida (Tenrecida), Carnivora (Canidae, Eu-

pleridae), Erinaceomorpha (Erinaceidae), Macroscelidea (Macroscelididae), Primates (Atelidae, Cebidae, Cheirogaleicidae, Galagidae, Lemuridae, Lepilemuridae, Lorisidae, Pitheciidae), Rodentia (Anomaluridae, Bathyergidae, Capromyidae, Caviidae, Cricetidae, Echimyidae, Gliridae, Muridae, Myocastoridae, Nesomyidae, Sciuridae, Pedetidae), Soricomorpha (Soledontidae, Soricidae, Talpidae).

Distribution. Africa (including Madagascar), Asia, Australia, and South America; two species are known from North America (*Myocastorobia myocastor* from *Myocastor coypus* and *Didelphichus serrifer* from *Didelphis marsupialis*); one species is known from Europe, Spain (*Atopomelus talpae* from *Talpa romana*); one species has cosmopolite distribution being associated with domesticated guinea-pig (*Chirodiscoides caviae* from *Cavia porcellus*).

Remarks. (i) The subfamily Centetesiinae Fain, 1976 was established for two genera, *Centetesia* and *Tenrecobia*, associated with Malagasy tenrecs (Fain 1976). According to Fain (1976), the main difference of Centetesiinae from other atopomelids (Atopomelinae) is in the striate membranes between coxal fields I fused with the gnathosoma. In atopomelins, these membranes are free. Other principal distinctions between ato-

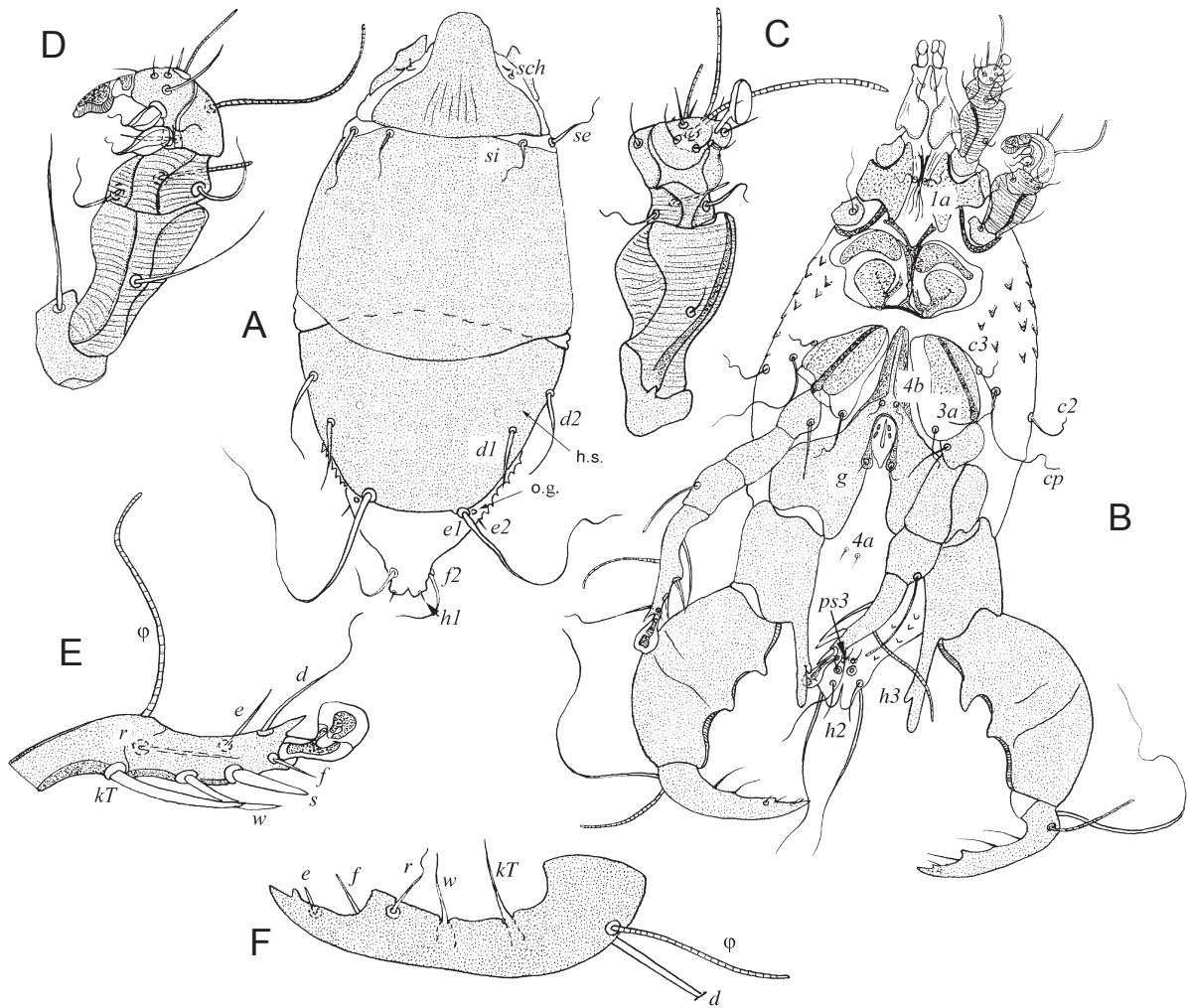


Fig. 25. *Atopomelus crociduræ* Fain et Lukoschus, 1977, male: A — dorsal view., B — ventral view, C — leg I in ventral view, D — leg II in ventral view, E — tibiotarsus III in lateral view, F — tibiotarsus IV in lateral view (after Bochkov and OConnor 2006, with minor modifications).

pomelid subfamilies were not mentioned. Our examinations of all four species of the subfamily Centetesiinae did not confirm the Fain's observations. These membranes are enlarged in the anterior part and, therefore, cover the posterior part of the subcapitulum ventrally. They, however, are not fused with the gnathosoma (examined on specially dissected specimens). Thus, *Centetesia* and *Tenrecobia* despite of some peculiarities, i.e. the enlarged membranes between coxal fields I and the absence of solenidion σ I on genua I do not possess any principal morphological differences allowing to separate them to their own subfamily. For this reason I synonymize the subfamily Centetesiinae syn. nov. with Atopomelinae.

(ii) The genus *Atopomelopsis* Fain, 1972 was created for one species *A. gibberosa* Fain, 1972 known from a single male at that time (Fain 1972a). Domrow (1992) based on his own material synonymized this species with *Dasyurochirus*

longipilis Fain, 1971 originally described from females (see Fain 1972a). *Dasyurochirus longipilis* differs from other species of the genus *Dasyurochirus* in both sexes, by the not separated postscapular shield and in males, by solenidion ϕ III situated in the middle part of the segment. In both sexes of other *Dasyurochirus* spp., the postscapular shield is paired and in males, solenidion ϕ III situated in posterior part of the segment. I, therefore, restore the genus *Atopomelopsis* Fain, 1972 stat. res. with one species, *A. longipilis* (Fain, 1971) (= *A. gibberosa*, type species).

(iii) The genus *Neolabidocarpus* Gunther, 1942 has uncertain status. It was described by Gunther (1942) for a single species *Labidocarpus buloloensis* Gunther, 1940 and originally placed in the subfamily Labidocarpinae (subfamily of the family Chirodoscidae now). Domrow (1958) examined the holotype of *N. buloloensis* and concluded that it is a tritonymph belonging to the fam-

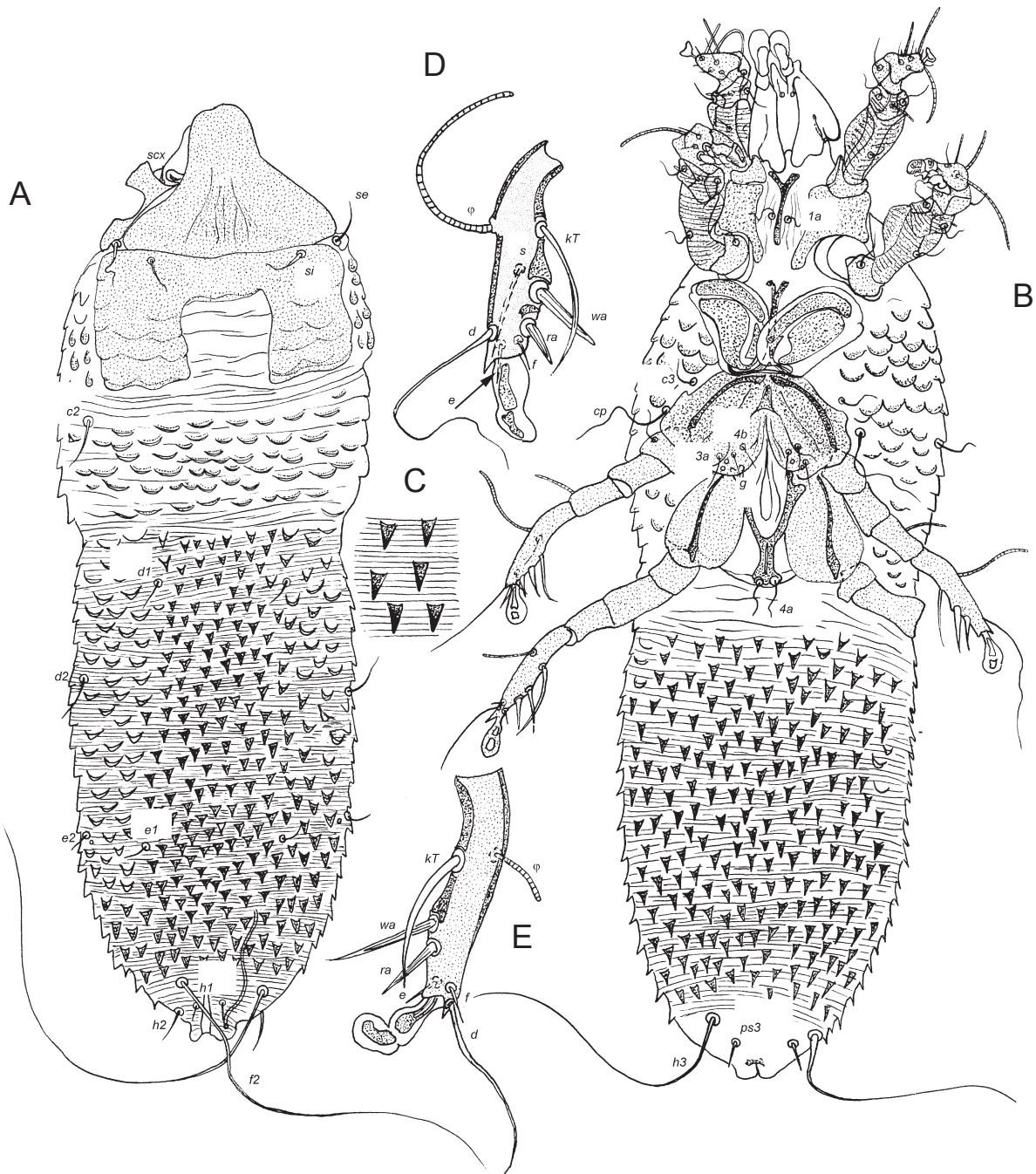


Fig. 26. *Atopomelus crocidurae* Fain et Lukoschus, 1977, female: A — dorsal view, B — ventral view, C — dorsal scales of idiosoma, D — tibiotarsus III in lateral view, E — tibiotarsus IV in lateral view (after Bochkov and OConnor 2006, with minor modifications).

ily Atopomelidae rather than the Chirodiscidae. He also pointed that “P. 45: Since the holotype is a nymph, and the remainder of the material was destroyed during the war (Gunther, in litt.), it appears best to keep this genus and species apart until fresh adult material proves them valid or otherwise”. Later on, Domrow (1962) recorded specimens which were tentatively determined as *Neolabidocarpus* sp. In his annotated list of parasitic mites of Australian vertebrates, he (Domrow 1992) attributed females of this species to *Cy-*

tostethum euphallus Fain et Domrow, 1975 and males to *Petrogalochirus* sp. I believe that a nymph of *N. buloloensis* actually belongs by the same way to one from these two above mentioned genera, but new atopomelid specimens should be collected from the type host of this species, *Thylogale coxenii* (Diprodontia: Macropodidae) before the final decision.

(iv) The comparison of holotypes of *Bathyer-golichus cryptomys* Fain et Bochkov, 2003 syn. nov. described by Fain and Bochkov (2003) and

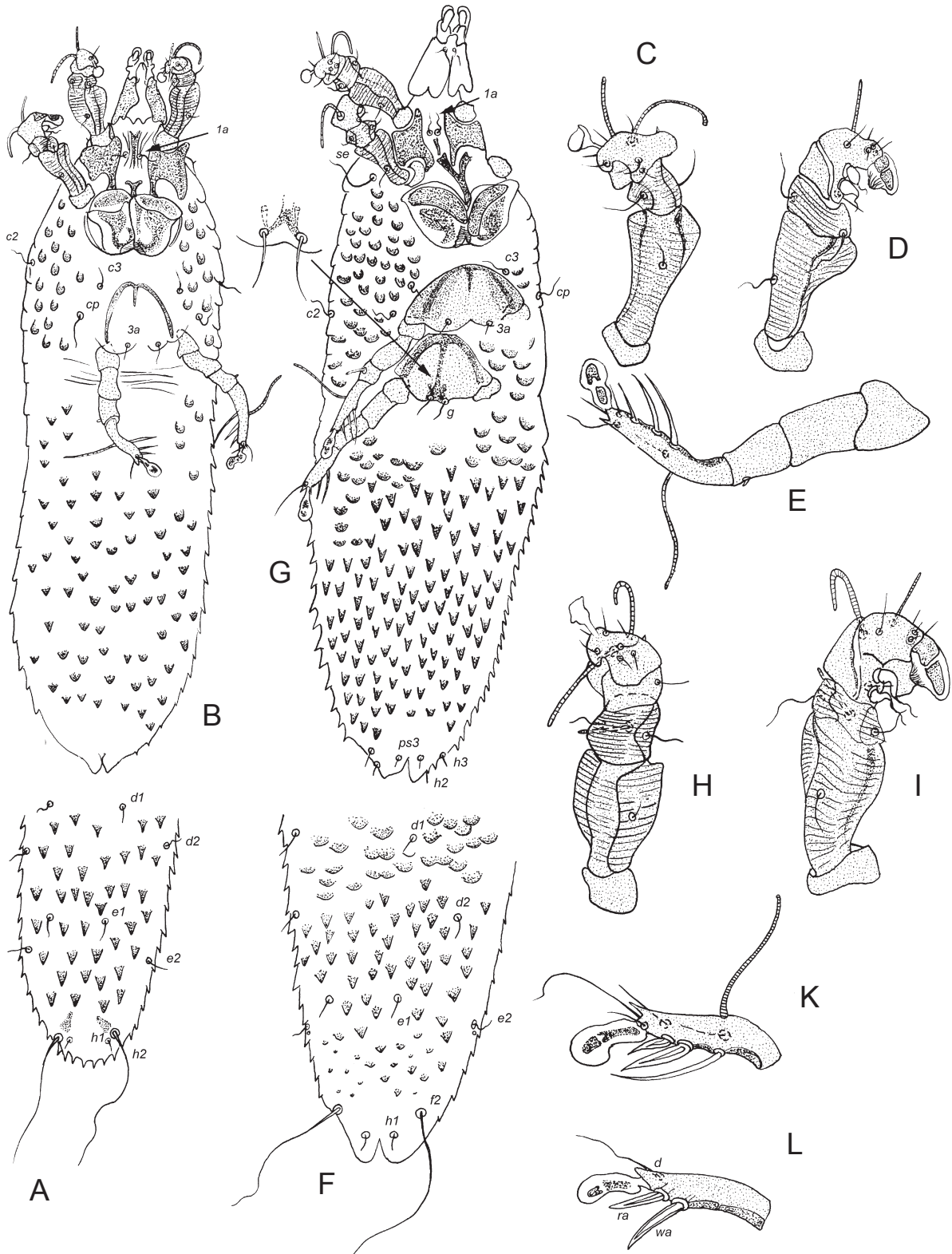


Fig. 27. *Atopomelus crociduræ* Fain et Lukoschus, 1977, immature instars: larva (A—E), A — hysterosoma, dorsal view, B — ventral view, C — leg I in ventral view, D — leg II in ventral view, E — tarsus III in lateral view. Protonymph (F—L), F — hysterosoma in dorsal view, G — ventral view, H — leg I in ventral view, I — leg II in ventral view, K — tibiotarsus III in lateral view, L — tibiotarsus IV in lateral view (after Bochkov and OConnor 2006, with minor modifications).

B. hottentotus Fain, 1986 revealed their full character similarity. Therefore I consider *B. cryptotys* syn. nov. as a junior synonym of *B. hottentotus*.

Main references. Fain (1972a) — revision of Australian and new Guinean Atopomelidae, Fain (1972b) — revision of African Atopomelidae;

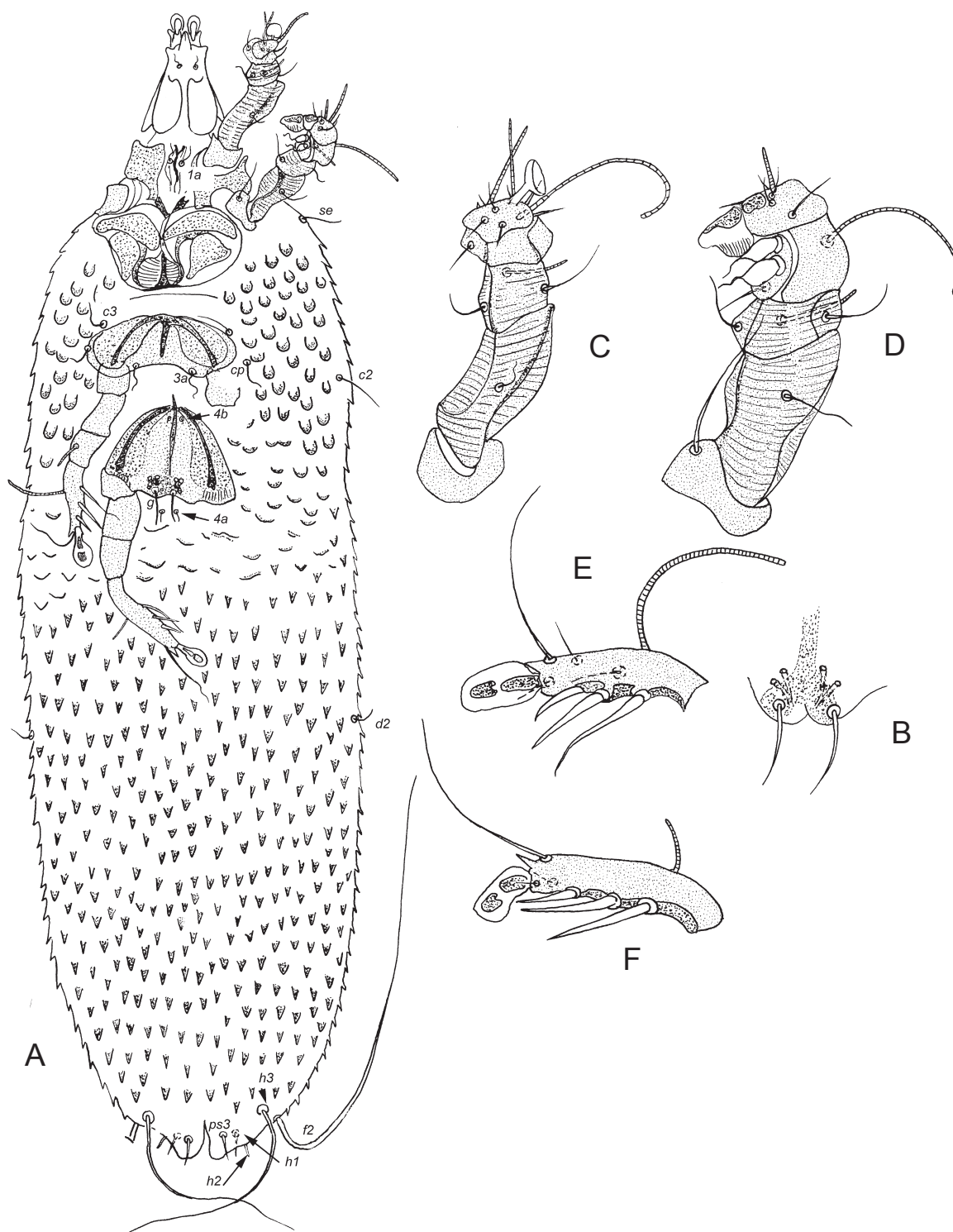


Fig. 28. *Atopomelus crocidurae* Fain et Lukoschus, 1977, tritonymph: A — ventral view, B — genital papillae, C — leg I in ventral view, D — leg II in ventral view, E — tibiotarsus III in lateral view, F — tibiotarsus IV in lateral view (after Bochkov and OConnor 2006, with minor modifications).

Fain (1976) — revision of Malagasy Atopomelidae, Fain (1979a) — revision of South American Atopomelidae, OConnor (1982) — diagnosis of Atopomelidae and references, Domrow (1992) — annotated checklist and keys to Australian Atopomelidae, Bochkov and OConnor (2005a) — review of the subgenus *Listrophoroides* s.str.,

Bochkov and OConnor (2006) — revision of Philippine Atopomelidae.

**Key to genera of the family Atopomelidae
Gunther, 1942**

(i) The genus *Neolabidocarpus* Gunther, 1942 known only from a tritonymph is not included.

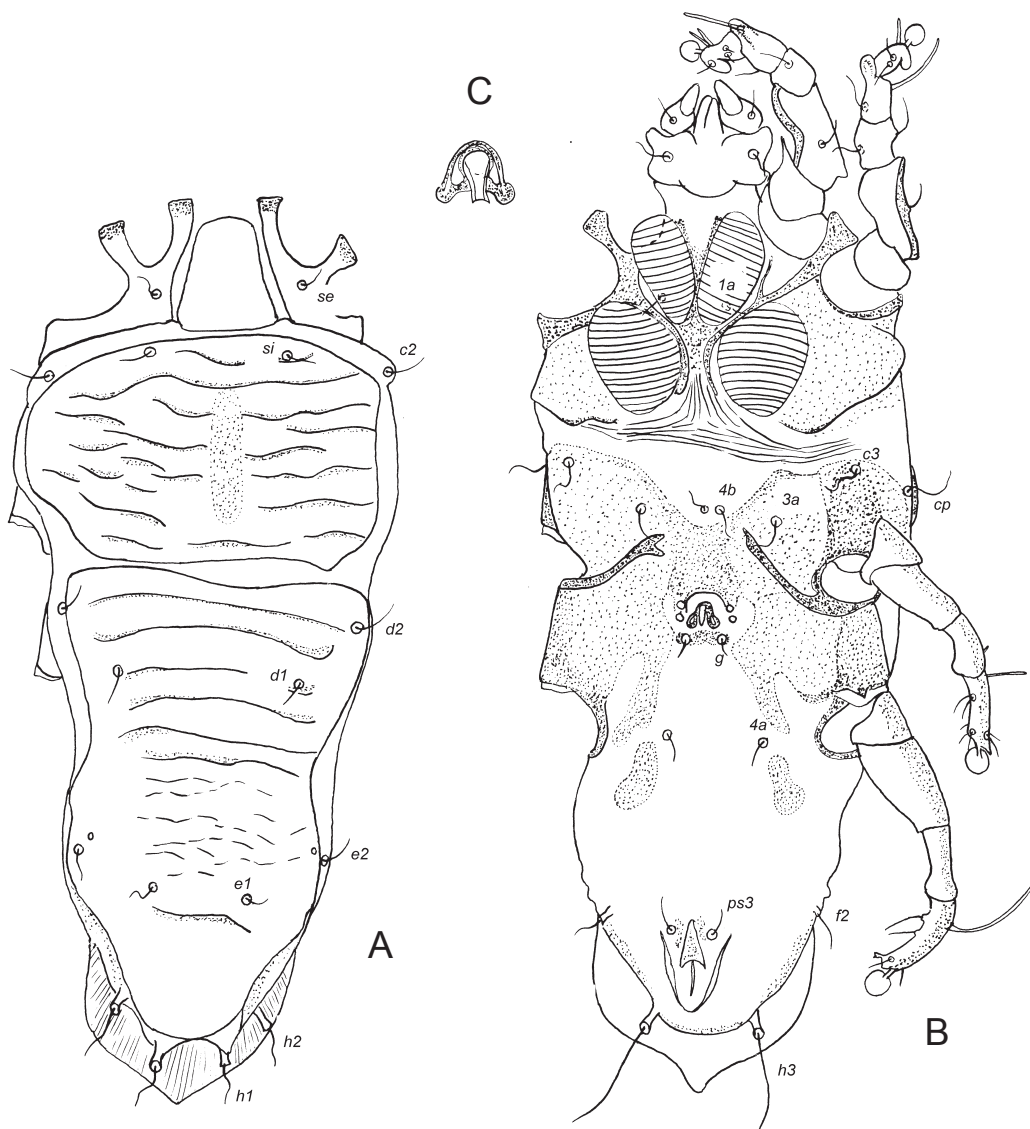


Fig. 29. *Listrophoroides mindanensis* Fain et Lukoschus, 1981, male. A — dorsal view, B — ventral view, C — aedeagus (after Bochkov and OConnor 2006, with minor modifications).

(ii) The genus *Didelphoecius* Fain, 1970 is highly polymorphic and probably not monophyletic. In most *Didelphoecius* spp., the prescapular and postscapular shields are paired and anterior shields are fused with the corresponding posterior shields; in females, the hysteronotal shield is paired and each piece is transversely separated into a pair of the anterior and posterior hysteronotal shields; in males, the hysteronotal shield with a deep and wide antero-median incision. However, in some species of the genus, this combination of characters is different from above described.

- 1. Both sexes: Solenidion ϕ III situated in basal or median part of tibiotarsus 8
- Both sexes: Solenidion ϕ III situated in apical part of tibiotarsus 2
- 2. Both sexes: Postscapular shield present 4
- Both sexes: Postscapular shield absent 3

- 3. Both sexes: Prescapular shield with distinctly sclerotized lateral and posterior margins and weakly sclerotized internal part (U-shaped sclerotization). Male: Hysteronotal shield paired. Adanal shields present. Opisthosomal lobe with small terminal cleft. Tarsus IV with ambulacral disc *Koalachirus* Fain, 1970
- Both sexes: Prescapular shield with distinctly sclerotized lateral and posterior margins and only slightly less sclerotized internal part. Male: Hysteronotal shield unpaired. Adanal shields absent. Opisthosomal lobe without terminal cleft. Tarsus IV without ambulacral disc *Petrogalochirus* Fain, 1970
- 4. Both sexes: Anterior part of propodonotum without apophyses. Tibioarsus III not more than with 1 conical seta 5
- Both sexes: Anterior part of propodonotum

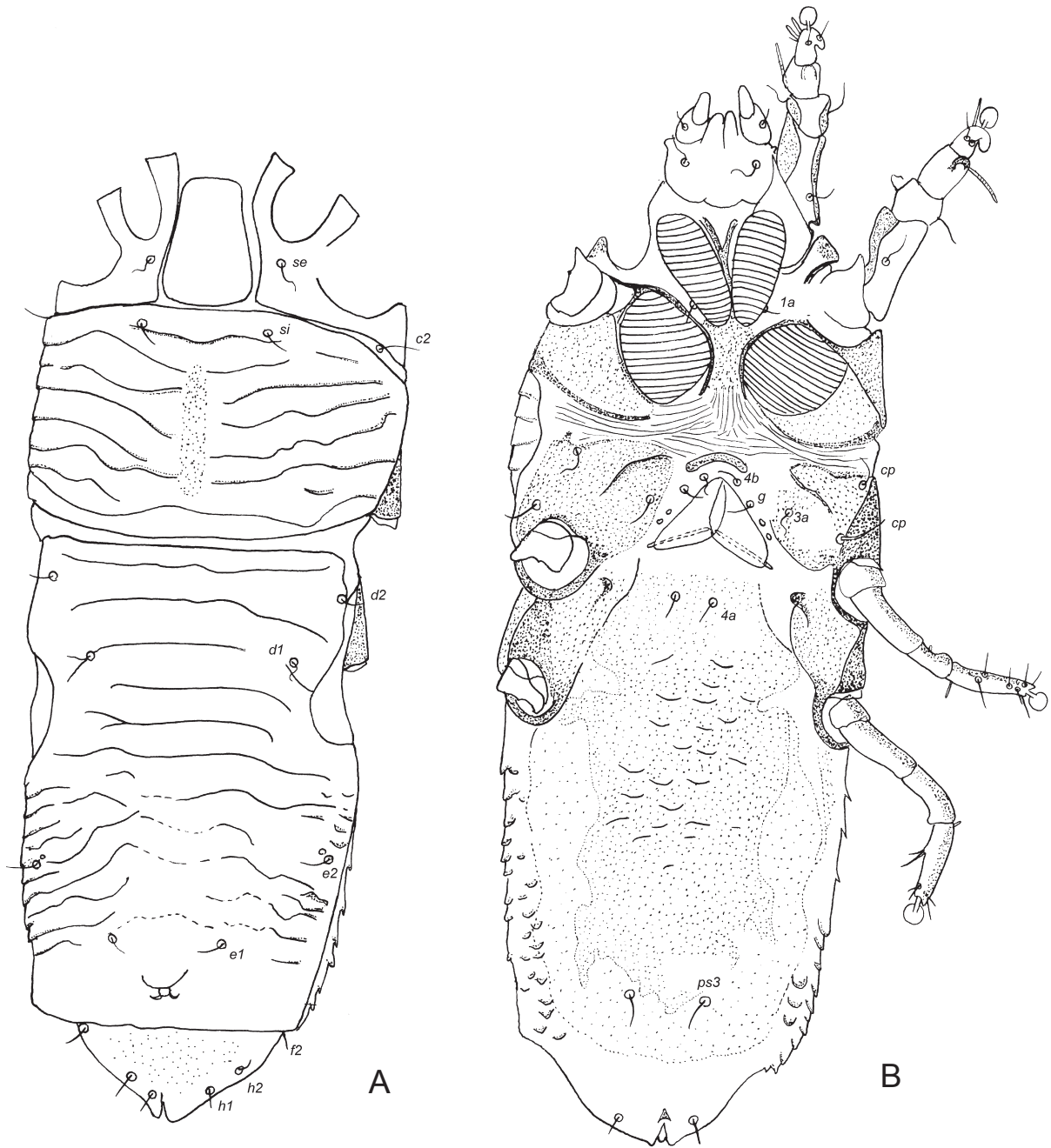


Fig. 30. *Listrophoroides mindanensis* Fain et Lukoschus, 1981, female: A — dorsal view, B — ventral view (after Bochkov and OConnor 2006, with minor modifications).

with 2 pairs of short apophyses. Tibiotarsus III with 2 conical setae *Petaurobia* Fain, 1971
 5. Both sexes: Idiosoma subcylindrical 7
 — Both sexes: Idiosoma dorso-ventrally flattened 6
 6. Both sexes: Prescapular shield present *Campylochirus* Trouessart, 1893
 — Male (Female unknown): Prescapular shield absent *Distoechurobia* Fain, 1972
 7. Male (Female unknown): Tarsus IV with ambulacral disc *Procytostethum* Fain, 1971

— Male: Tarsus IV without ambulacral disc *Cytostethum* Domrow, 1956
 8. Both sexes: Intercoxal membranes of fields I not extending to infrasubcapitulum 10
 — Both sexes: Intercoxal membranes of fields I covering most part of infrasubcapitulum 9
 9. Both sexes: Postscapular shield absent. Striated membranes of coxal fields II widely separated from each other. Female: Setae 4a situated distinctly anterior to insertion of legs IV. Male: Opisthosomal lobe widely rounded, without ter-

- minal cleft *Centetesia* Lawrence, 1955
 — *Both sexes*: Postscapular shield present. Striated membranes of coxal fields II situated close to each other. *Female*: Setae 4a situated distinctly posterior to insertion of legs IV. *Male*: Opisthosomal lobe with terminal cleft
 *Tenrecobia* Lawrence, 1955
 10. *Both sexes*: Postscapular shield present (in *Micropotamogalichus* spp. prescapular shield represented by pair of small sclerotized patches situated ventro-laterally) 20
 — *Both sexes*: Postscapular shield absent (in *Listrocarpus* spp., narrow longitudinal prescapular shield separated transversally in two unequal parts — short anterior and long posterior shields which could be confused with postscapular shield; in *Austrochirus* s.str., postscapular shield reduced to Y-shaped sclerite) 11
 11. *Both sexes*: Prescapular shield present. *Female*: Hysteronotal shield absent. *Male*: Opisthosomal lobe weakly developed without or with small terminal cleft 12
 — *Both sexes*: Prescapular shield absent. *Female*: Hysteronotal shield present. *Male*: Opisthosomal lobe well developed with distinct terminal cleft
 *Caenolestolichus* Fain et Bochkov, 2003
 12. *Female*: Dorsal surface of idiosoma striated or covered with pointed scales (*Metadidelphoecius* spp.). *Male*: Paranal suckers absent. Tarsus IV without ambulacral disc 15
 — *Female*: Dorsal surface of idiosoma covered with widely rounded scales. *Male*: Paranal suckers present. Tarsus IV with ambulacral disc 13
 13. *Both sexes*: Tibia and tarsus of legs III and IV completely fused to each other 14
 — *Both sexes*: Tibia and tarsus of legs III and IV not completely fused to each other
 *Notoryctobia* Fain et Lukoschus, 1981
 14. *Both sexes*: Prescapular shield sclerotized in lateral and anterior part (arch-like) or paired. Y-shaped propodonotal sclerite present (*Austrochirus* s.str.) or absent (*Austrochiroides*). *Male*: ventro-opisthosomal copulatory projections absent
 *Austrochirus* Womersley, 1943
 — *Male (Female unknown)*: Prescapular shield monotonously sclerotized, unpaired. Y-shaped sclerite absent. 1 pair of large, striated, conical copulatory projections flanking paranal suckers
 *Labidopygus* Fain et Domrow, 1973
 15. *Both sexes*: Idiosoma dorsally without scales. Posterior margin of prescapular shield without extensions 16
 — *Female (Male unknown)*: Idiosoma dorsally with pointed scales. Posterior margin of prescapular shield with 3–4 short triangular extensions
 *Metadidelphoecius* Fain, 1979
 16. *Both sexes*: Prescapular shield entire 17
 — *Both sexes*: Prescapular shield transverse separated in to small anterior and narrow elongated posterior parts
 *Listrocarpus* Fain, 1967
 17. *Both sexes*: Parasitizing Soledontidae or capromyid rodents other than *Plagiodontia*. *Female*: Copulatory tube much shorter than opisthosoma ..
 18
 — *Both sexes*: Parasitizing *Plagiodontia* spp (Capromyidae). *Female*: Copulatory tube and opisthosoma subequal in length
 *Plagiodontochirus* Fain, 1976
 18. *Both sexes*: Idiosoma subcylindrical and slightly dorso-ventrally flattened. *Male*: Hysteronotal shield present. Genu IV with distinct invagination and apophysis ventrally 19
 — *Both sexes*: Idiosoma egg-shaped, slightly elongated. *Male*: Hysteronotal shield absent. Genu IV without invagination and apophysis
 *Capromylichus* Fain, 1970
 19. *Both sexes*: Prescapular shield with distinctly sclerotized lateral and posterior margins (U-shaped). Dorsal striation of idiosoma not forming transverse folds. *Female*: Epigynum distinct. Copulatory tube shorter 10 μm . *Male*: Hysteronotal shield with very deep antero-median concavity; setae *dI* situated off this shield
 *Cubanochirus* Fain, 1970
 — *Both sexes*: Prescapular shield monotonously sclerotized. Dorsal striation of idiosoma forming transverse folds. *Female*: Epigynum indistinct or absent. Copulatory tube about 15 μm long. *Male*: Hysteronotal shield without antero-median concavity; setae *dI* situated on this shield
 *Capromysia* Fain, 1979
 20. *Both sexes*: Postscapular shield(s) situated dorsally or latero-dorsally, distinct 21
 — *Both sexes*: Postscapular shields situated ventro-laterally, represented by pair of small sclerotized patches ... *Micropotamogalichus* Fain, 1970
 21. *Both sexes*: Setae *pRII* absent 23
 — *Both sexes*: Setae *pRII* present 22
 22. *Both sexes*: Idiosoma subcylindrical. Seta *sRIII* present. *Male*: Paranal suckers present. Legs IV enlarged, much longer than legs III. Femur IV with spurs
 *Atopomelus* Trouessart 1918
 — *Both sexes*: Idiosoma flattened dorso-ventrally. Seta *sRIII* absent. *Male*: Paranal suckers absent. Legs IV only slightly longer than legs III. Femur IV without spurs
 *Bathyergolichus* Fain, 1970

23. *Both sexes*: Postscapular shield unpaired (in some *Murichirus* spp., postscapular shield separated by narrow non-sclerotized median line) 27
 — *Both sexes*: Postscapular shield paired (in some *Didelphoecius* spp., postscapular shield unpaired; in male *Metachiroecius* sp., postscapular shields connected to each other by narrow transverse band) 24
24. *Both sexes*: Prescapular and postscapular shields not fused to each other. *Female*: Hysteronotal shield absent, present unpaired (*Dasyurochirus* (*Dasyurochiroides* spp.)) or small paired (*Dasyurochirus biscutatus*), but not transverse separated onto anterior and posterior parts 26
 — *Both sexes*: Prescapular and postscapular shields fused to each other (in some *Didelphoecius* spp. these shields not fused). *Female*: Hysteronotal shield paired and in addition transverse separated onto small anterior end elongated posterior parts (in some *Didelphoecius* spp., hysteronotal shield unpaired or paired but not transversely subdivided) 25
25. *Female*: Prescapular and postscapular shields not completely fused to each other. *Male*: Postscapular shields separated in most species. Coxal fields III not completely fused to each other
 *Didelphoecius* Fain, 1970
 — *Both sexes*: Prescapular and postscapular shields completely fused to each other. *Male*: Postscapular shields connected by narrow transverse band. Coxal fields III completely fused to each other..... *Metachiroecius* Fain et al., 1996
26. *Both sexes*: Anterior part of hysteronotum striated or with scales, but not covered by cellular-like pattern. *Male*: Hysteronotal shield paired. Femur and genu III not fused. Ambulacral disc of tarsus IV present *Dasyurochirus* Fain, 1970
 — *Both sexes*: Anterior part of hysteronotum covered by cellular-like pattern. *Male*: Hysteronotal shield unpaired. Femur and genu III fused. Ambulacral disc of tarsus IV absent
 *Didelphilichus* Fain, 1970
27. *Both sexes*: Prescapular and postscapular shields separated from each other (in some *Oryzomysia* spp., prescapular and postscapular shields partially fused, but in male opisthosoma distinctly bilobed) 31
 — *Both sexes*: Prescapular and postscapular shields partly or fully fused to each other. *Male*: Opisthosoma not distinctly bilobed 28
28. *Both sexes*: Row of scales posterior to coxal fields II absent. *Male*: Hysteronotal shield unpaired. Ambulacral disc present on tarsus IV. Paranal suckers strongly reduced. Opisthosomal lobe moderately developed and posterior margin of idiosoma scalloped 29
 — *Both sexes*: Row of scales present ventrally immediately posterior to coxal fields II. *Male*: Hysteronotal shield paired. Ambulacral disc absent on tarsus IV. Paranal suckers moderately developed. Opisthosomal lobe strongly reduced and posterior margin of idiosoma widely rounded
 *Atellana* Domrow, 1958
29. *Female*: Setae *4a* widely separated. *Male*: Opisthosomal venter without scales. Bases of setae *ps3* jointed with paranal suckers. Legs III and IV distinctly thickened 30
 — *Female*: Bases of setae *4a* almost joined. *Male*: Opisthosomal venter with lateral scales. Setae *ps3* situated far anterior to paranal suckers. Legs III and IV moderately developed
 *Austrobius* Fain, 1971
30. *Female*: Hysteronotal shield present. *Male*: Most dorsal setae of idiosoma short, distinctly shorter than prodorsal shield
 *Sclerochiroides* Fain, 1970
 — *Female*: Hysteronotal shield absent. *Male*: Most dorsal setae of idiosoma long, distinctly longer than prodorsal shield
 *Sclerochiroopsis* Fain, 1972
31. *Both sexes*: Setae *sRIII* present 33
 — *Both sexes*: Setae *sRIII* absent 32
32. *Both sexes*: Setae *ps1* and *ps2* present. *Male*: Paranal suckers present ... *Isothricola* Fain, 1970
 — *Both sexes*: Setae *ps1* and *ps2* absent. *Male*: Paranal suckers absent
 *Listrophoroides* Hirst, 1923
33. *Both sexes*: Postscapular shield vary in shape but not reduced to narrow transverse band or pair of plates connected by narrow transverse bridge ..
 36
 — *Both sexes*: Postscapular shield reduced to narrow transverse band or narrowed only in its median part 34
34. *Both sexes*: Idiosoma subcylindrical. *Female*: Hysteronotal shield absent 35
 — *Both sexes*: Idiosoma slightly flattened dorsoventrally. *Female*: Hysteronotal shield present
 *Dromiciolichus* Fain, 1970
35. *Female*: Setae *h1* short, subequal to setae *h2* and *h3*. *Male*: Paranal suckers absent. Tarsus IV without ambulacral disc
 *Lemuroptes* Lawrence, 1958
 — *Female*: Setae *h1* long, much longer than setae *h2* and *h3*. *Male*: Paranal suckers present. Tarsus IV with ambulacral disc

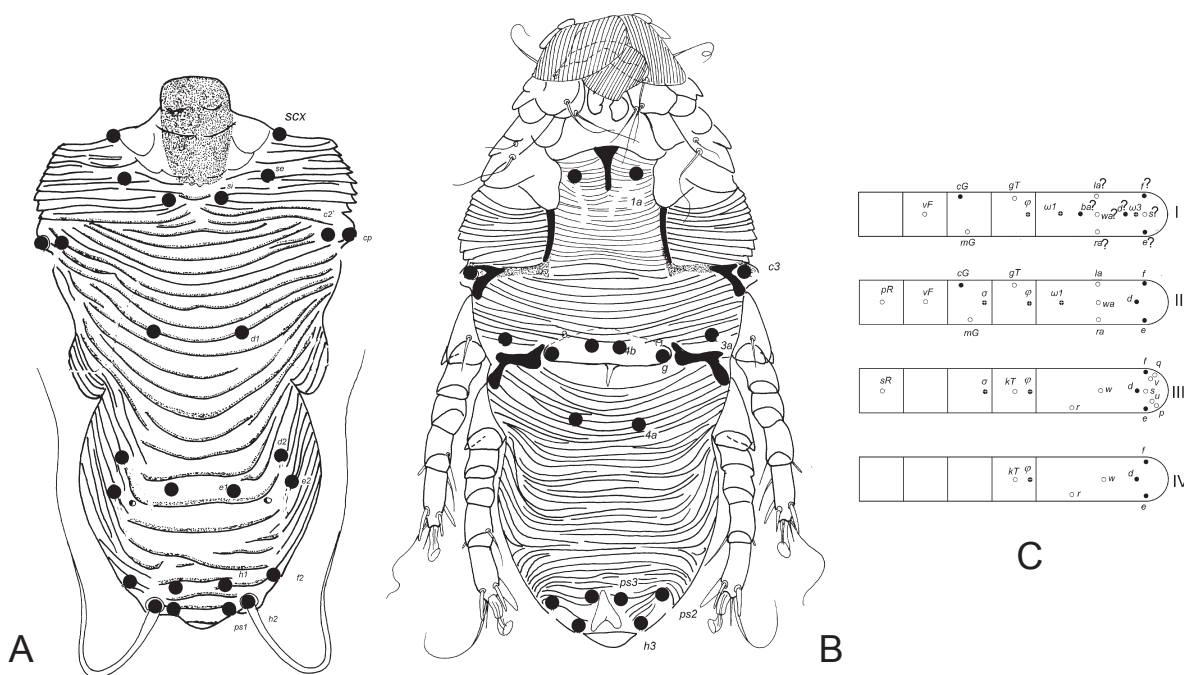


Fig. 31. Chirodiscidae, scheme of setation: A — idiosoma in dorsal view, B — same in ventral view, C — legs I-IV.

..... *Atopomelopsis* Fain, 1972 stat. res.
 36. *Both sexes*: Tibiotarsi III with not more than 2 thickened setae. Propodonotum without anterior apophyses 36
 — *Both sexes*: Tibiotarsi III with 3 thickened setae. Propodonotum with 2 pair of short lateral apophyses *Scolonoticus* Fain, 1971
 37. *Female*: Hysteronotal shield present, covering most part of hysteronotum. *Male*: Paranal suckers present 40
 — *Female*: Hysteronotal shield absent or, if present, covering less than anterior third of hysteronotum. *Male*: Paranal suckers absent 38
 38. *Both sexes*: Idiosoma subcylindrical or egg-shaped. *Male*: Apodemes IVa fused to each other. Tarsus IV with ambulacral disc 39
 — *Both sexes*: Idiosoma flattened dorso-ventrally. *Male*: Apodemes IVa widely separated from each other. Tarsus IV without ambulacral disc
 *Domingoecius* Fain et Lukoschus, 1979
 39. *Both sexes*: Idiosoma strongly elongated, subcylindrical. Hysteronotum distinctly striated
 *Murichirus* Fain, 1971
 — *Male (Female unknown)*: Idiosoma egg-shaped. Hysteronotum without striations
 *Phalangerobia* Fain, 1972
 40. *Both sexes*: Posterior half of intercoxal apodeme II not bifurcate 41
 — *Both sexes*: Posterior half of intercoxal apodeme II bifurcate *Chirodiscoides* Hirst, 1917
 41. *Both sexes*: Prescapular shield unpaired 43

— *Both sexes*: Prescapular shield paired 42
 42. *Both sexes*: Tibiotarsus III with 2 thickened setae. *Male*: Paranal suckers moderately developed. Penis short, subequal in length to setae 4a ..
 *Myocastorobia* Fain, 1976
 — *Male (Female unknown)*: Tibiotarsus III without thickened setae. *Male*: Paranal suckers very large. Penis very long, subequal in length to legs III
 *Prodidelphoecius* Fain, 1976
 43. *Both sexes*: Hysteronotal shield unpaired
 *Oryzomysia* Fain, 1970
 — *Female (Male unknown)*: Hysteronotal shield paired
 *Euryzygomysia* Fain, 1979

Family Chirodiscidae Trouessart, 1892

Type genus: *Chirodiscus* Trouessart et Neumann, 1890

Table 9, Figs. 31–42

Diagnosis. *Both sexes*. Cheliceral hood and ventral apophysis of movable digit present. Apices of palpal tibiae with indistinct membranes. Idiosoma subcylindrical or flattened dorso-ventrally. Supracoxal sclerite present, setae *scx* present or absent, supracoxal opening distinct. In some Labidocarpinae, propodonotal shield subdivided on prescapular and postscapular shields. Genital papillae usually present (absent in many Labidocarpinae). Intercoxal membranes striated, distinctly developed between coxal fields I and II or even occupying coxal field surfaces, not forming curved flaps. Idiosomal setae *vi* and *c1* absent (*c1* present

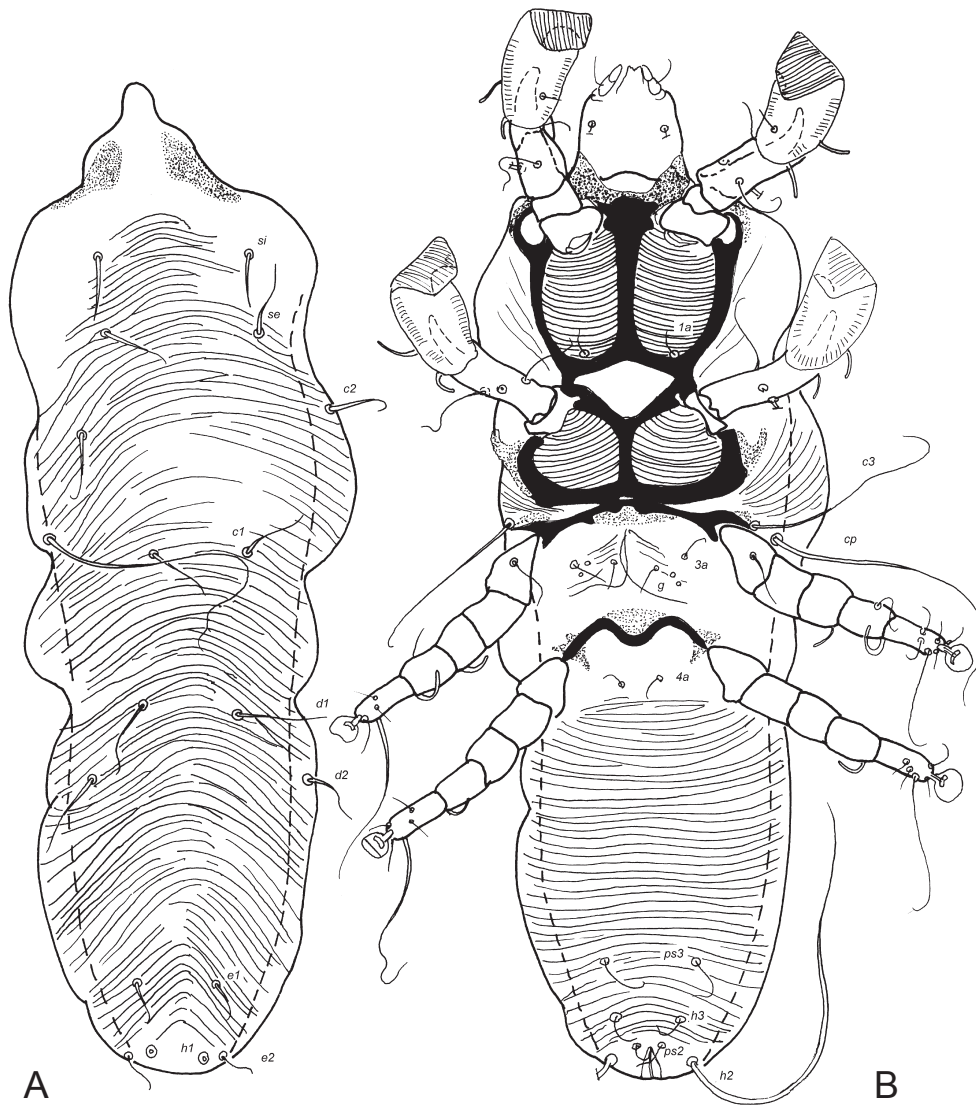


Fig. 32. *Chirodiscus amplexans* Trouessart et Neumann, 1889, female: A — dorsal view, B — ventral view (after Fain (1972a), with minor modifications).

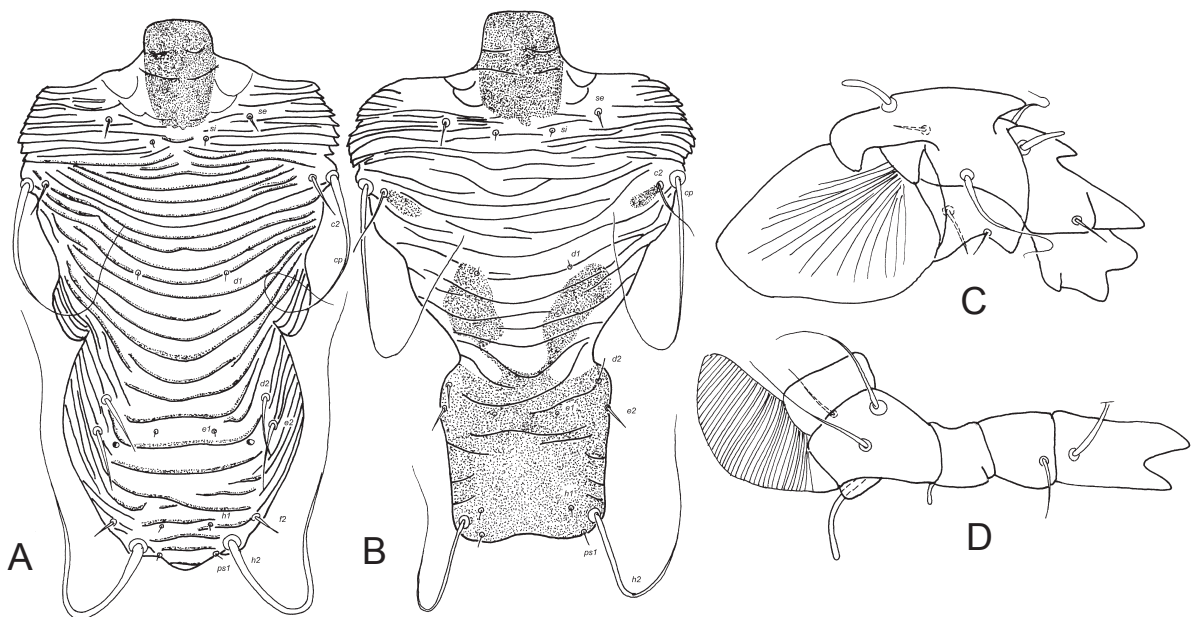


Fig. 33. *Lemuroecius cheirogalei* Fain, 1968: A — female in dorsal view, B — male in dorsal view, C — leg I in ventral view, D — leg II in ventral view (after Fain (1968b), with minor modifications).

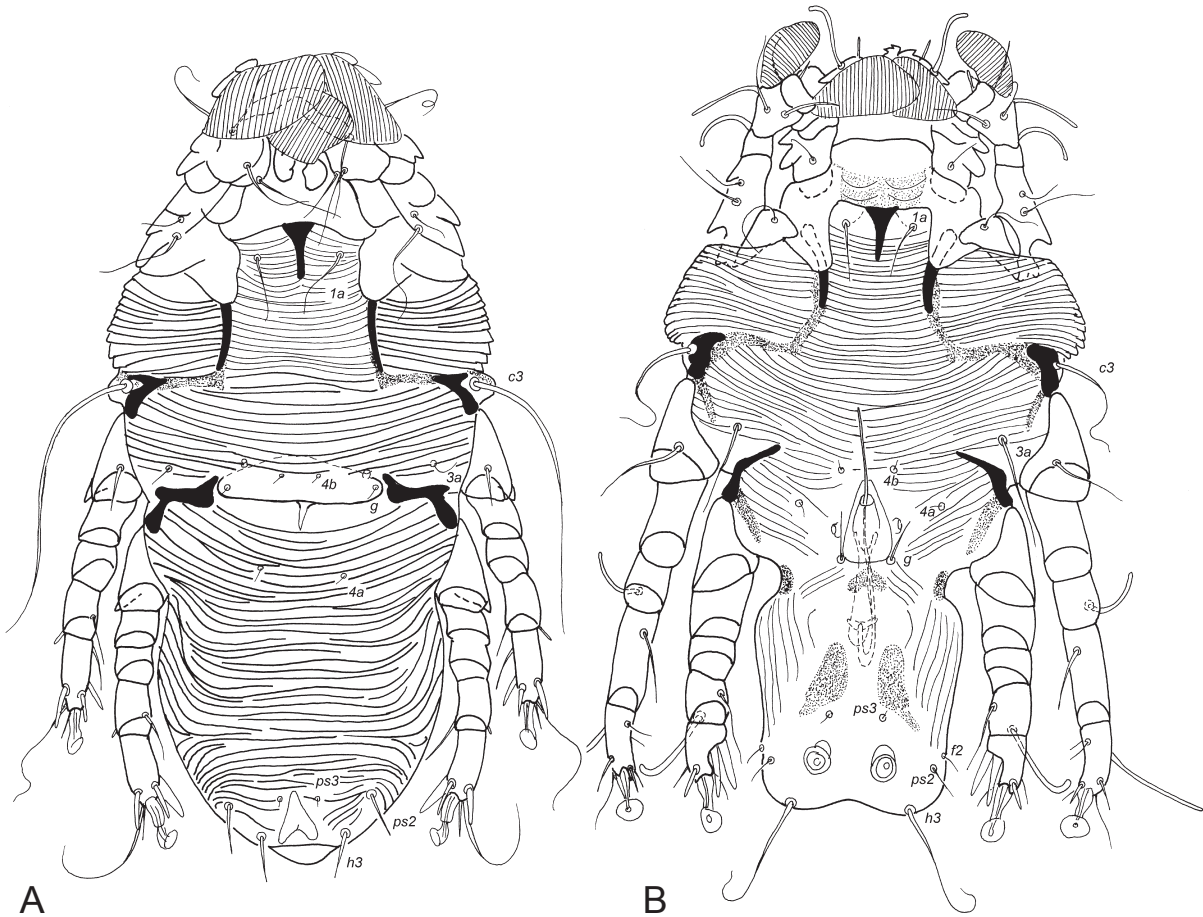


Fig. 34. *Lemuroecius cheirogalei* Fain, 1968: A — female in ventral view, B — male in ventral view (after Fain (1968b), with minor modifications).

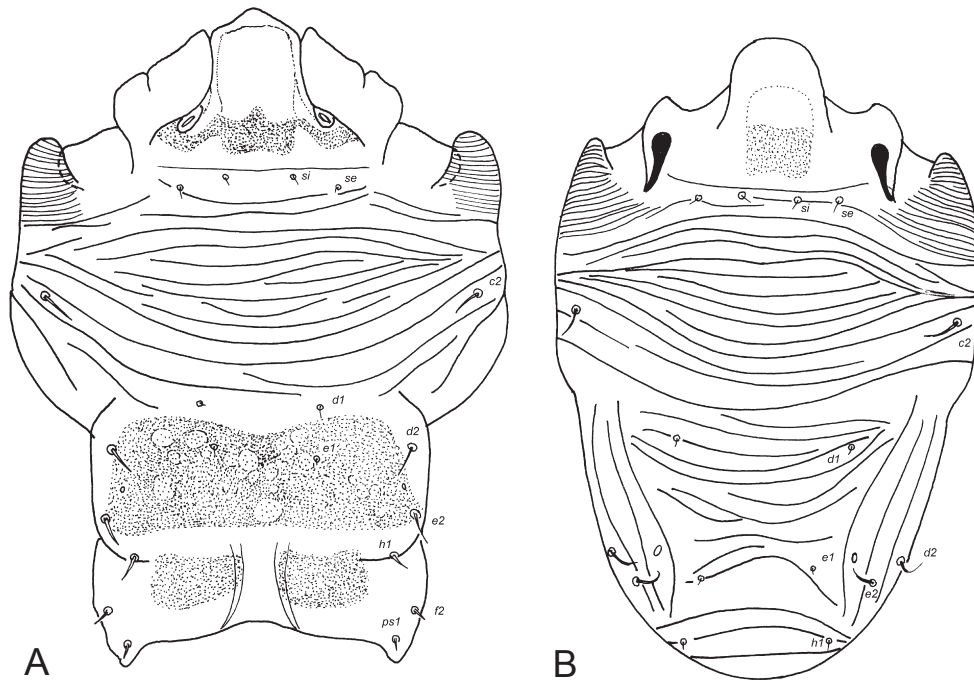


Fig. 35. *Schizocoptes conjugatus* Lawrence, 1944: A — male in dorsal view, B — female in dorsal view (after Fain (1971), with minor modifications).

in some Labidocarpini and, probably, Chirodiscinae). In most derived Labidocarpinae many idio-

somal setae lost. Many setae of apical segments of legs I and II absent. Segments of legs I and II ar-

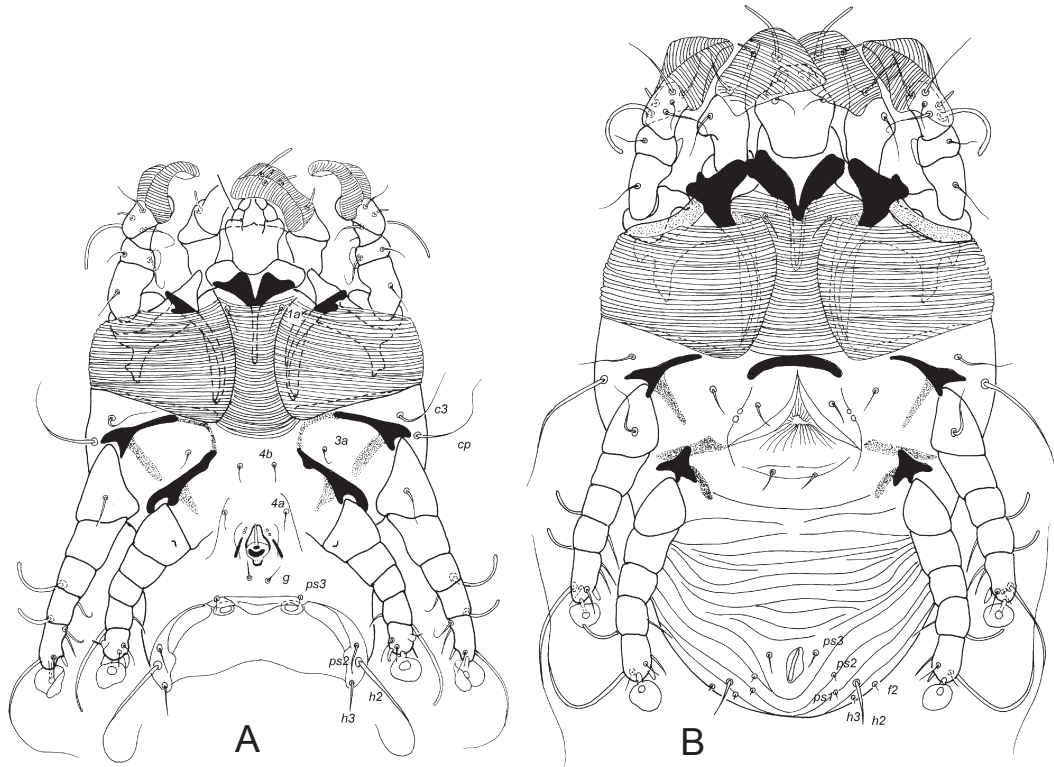


Fig. 36. *Schizocoptes conjugatus* Lawrence, 1944: A — male in ventral view, B — female in ventral view (after Fain (1971), with minor modifications).

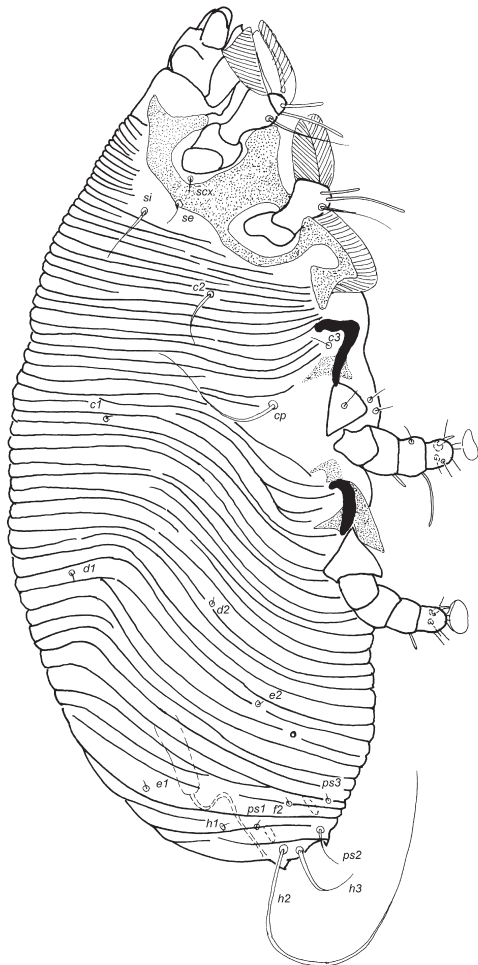


Fig. 37. *Schizocarpus mingaudi* Trouessart, 1896, female in lateral view.

ticulated in archaic genera (tibia and tarsus I always fused) and partially or completely fused in derived genera, tarsi I and II bearing striated clasping membranes and completely devoid of ambulacra. In some Labidocarpinae, ambulacra absent on tarsi III and IV and femur and genu III and IV fused. If ambulacra III and IV present, ambulacral sclerites moderately developed.

Female. Hysteronotal and opisthonal shields absent. Ovipore shaped as an inverted Y or transversal (Lemuroeciinae). Epigynum distinctly developed and often fused with apodemes IIIa (Chirodiscinae and many Labidocarpinae), it lost in Lemuroeciinae. Anal opening situated ventrally or ventro-terminally. Oviparous.

Male. Opisthosomal lobes and paranal suckers usually present. Legs III and IV normally developed. Setae *d*IV setiform or stick-like.

Taxa included: 228 species and 26 genera in 4 subfamilies, Chirodiscinae Trouessart, 1892 — *Chirodiscus* Trouessart et Neumann, 1890 (1 species); Labidocarpinae Gunther, 1942 — *Adentocarpus* Fain, 1972 (1 species), *Afrolabidocarpus* Fain, 1970 (5 species), *Alabidocarpus* Ewing, 1929 (37 species), *Asiolabidocarpus* Fain, 1972 (3 species), *Dentocarpus* Dusbabek et Cruz, 1966 (16 species), *Eulabidocarpus* Lawrence, 1948 (1 species), *Eurolabidocarpus* Fain et Aellen, 1994 (1 species), *Glossophagocarpus* Cruz, 1973 (1 spe-



Fig. 38. *Alabidocarpus molossicola* Fain, 1970: A — male in lateral view, B — female in lateral view (after Fain (1971), with minor modifications).

cies), *Labidocarpellus* Fain, 1976 (11 species), *Labidocarpoides* Fain, 1970 (6 species), *Labidocarpus* Trouessart, 1895 (12 species), *Lawrenceocarpus* Dusbabek et Cruz, 1966 (18 species), *Lutrilichus* Fain, 1970 (5 species), *Megadermicolus* Fain, 1971 (1 species), *Olalabidocarpus* Lawrence, 1948 (22 species), *Parakosa* McDaniel et Lawrence, 1962 (7 species), *Paralabidocarpus* Pinichpongse, 1963 (20 species), *Paralawrenceocarpus* Guerrero, 1992 (1 species), *Pseudoalabidocarpus* McDaniel, 1972 (3 species), *Pteropiella* Fain, 1970 (2 species), *Trilabidocarpus* Fain, 1970 (1), *Rynconyssus* Fain, 1967 (1 species), *Schizocarpus* Trouessart, 1896 (49 species), *Soricilichus* Fain, 1970 (2 species); Schizocoptinae Fain, 1970 — *Schizocoptes* Lawrence, 1944 (2 species); Lemuroeciinae Fain, 1968 — *Lemuroecius* Fain, 1968 (1 species).

Associations with hosts: inhabitants of hairs of therian mammals, Chirodiscinae — Marsupialia: unknown host; Labidocarpinae, Labidocarpini Fain, 1971 — Chiroptera (Emballonuridae, Furipteridae, Hipposideridae, Megadermatidae, Molossidae, Mormoopidae, Noctilionidae, Nycteridae,

Phyllostomatidae, Pteropodidae, Rhinolophidae, Vespertilionidae), Primates (Galagidae, Lorisidae), Schizocarpini Fain, 1971 — Carnivora (Mustelidae, Viverridae), Rodentia (Castoridae), Soricomorpha (Soricidae); Lemuroeciinae — Primates (Cheirogaleidae); Schizocoptinae — Afrosoricidae (Chrysochloridae).

The absolute majority of species of the tribe Labidocarpini are associated with chiropterans which are, probably, their initial hosts, and only mites of the monobasic genus *Rynconyssus* are associated with strepsirrhin primates. These associations are undoubted consequences of host shift from bats to primates.

Distribution. Chirodiscinae — Australia; Labidocarpinae, Labidocarpini — cosmopolite, Schizocarpini — Africa, Eurasia, and North America; Lemuroeciinae — Madagascar; Schizocoptinae — Africa.

Main references. Dubinina (1964) — life-cycle and biology of *Schizocarpus*, Fain (1968b) — description of *Lemuroecius*, Fain (1971) — revision of African Chirodiscidae, Fain (1972a) —

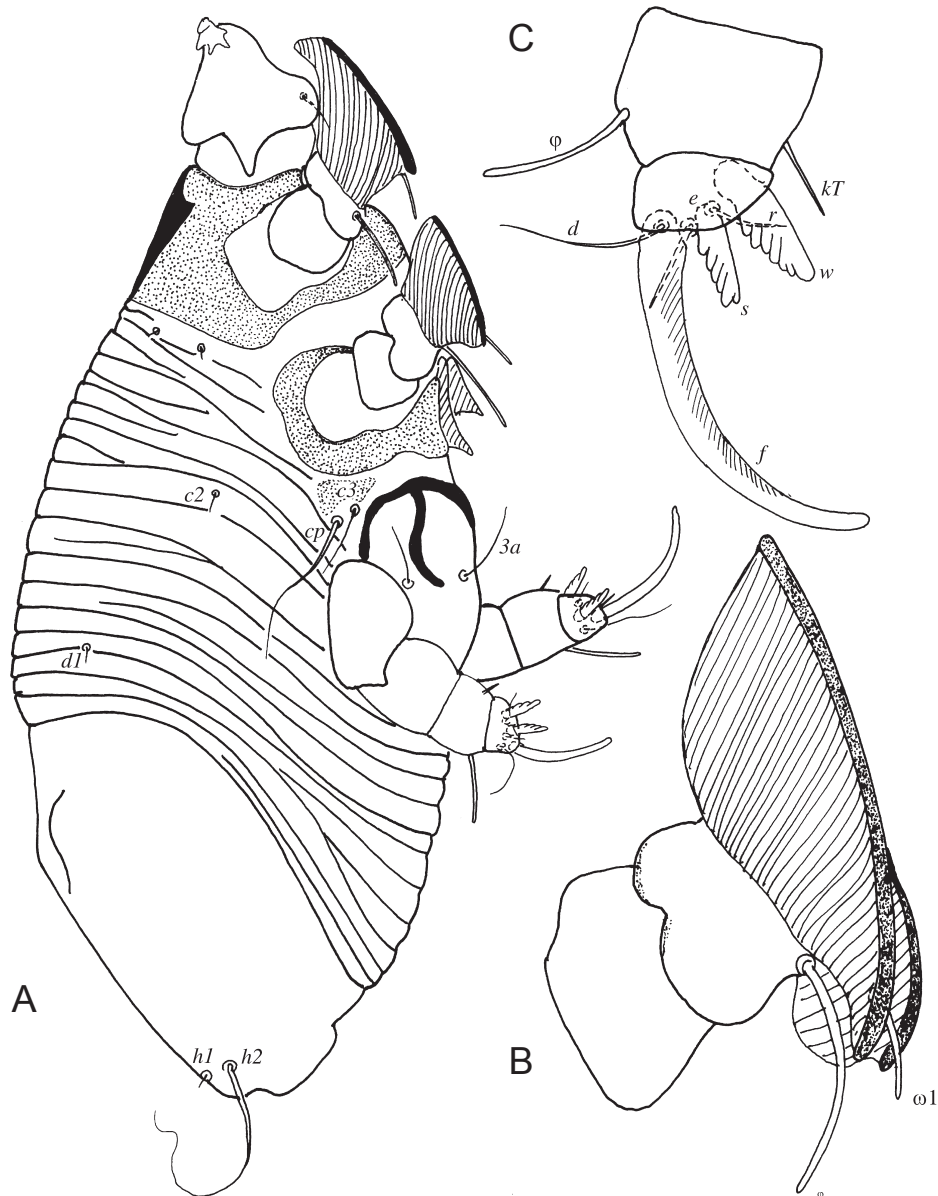


Fig. 39. *Alabidocarpus* sp., larva: A — lateral view, B — leg I in lateral view, C — tibia and tarsus III in ventral view.

redescription of *Chirodiscus amplexans* Trouessart et Neumann, 1890, Fain (1973) — revision of South American Chirodiscidae, Fain (1980a, 1981a, 1982a, b) — revision of Oriental Chirodiscidae and key to Labidocarpini, Fain and Lukoschus (1985) — revision of *Schizocarpus* from the European beaver, *Castor fiber*, Fain et al. (1984) and Fain and Whitaker (1988) — revision of *Schizocarpus* from the Canadian beaver, *Castor canadensis*, OConnor (1982) — diagnosis of Chirodiscidae and references, Guerrero (1992) — catalogue of Neotropical Labidocarpini.

**Key to genera of the family Chirodiscidae
Trouessart, 1892**

(partly based on Fain [1982b])

The monobasic genera *Eulabidocarpus* Lawrence, 1948 and *Glossophagocarus* Cruz, 1973

are insufficiently described and therefore not included in this key.

1. *Both sexes*: Legs II with 2 (trochanter and remaining fused segments) articulated segments ... 3 — *Both sexes*: Legs II with 5 or 4 (tibia and tarsus fused) articulated segments 2
2. *Both sexes*: Subcapitulum with 2 pairs of ventral apophyses. Tibia and tarsus of legs II fused. *Female*: Ovipore transverse. Epigynum absent Lemuroeciinae Fain, 1968 (*Lemuroecius* Fain, 1968) — *Both sexes*: Subcapitulum without ventral apophyses. Tibia and tarsus of legs II not fused. *Female*: Ovipore as an inverted Y. Epigynum present. Schizocoptinae Fain, 1970 (*Schizocoptes* Lawrence, 1944)
3. *Both sexes*: Apodemes Ib and IIa not fused. Se-

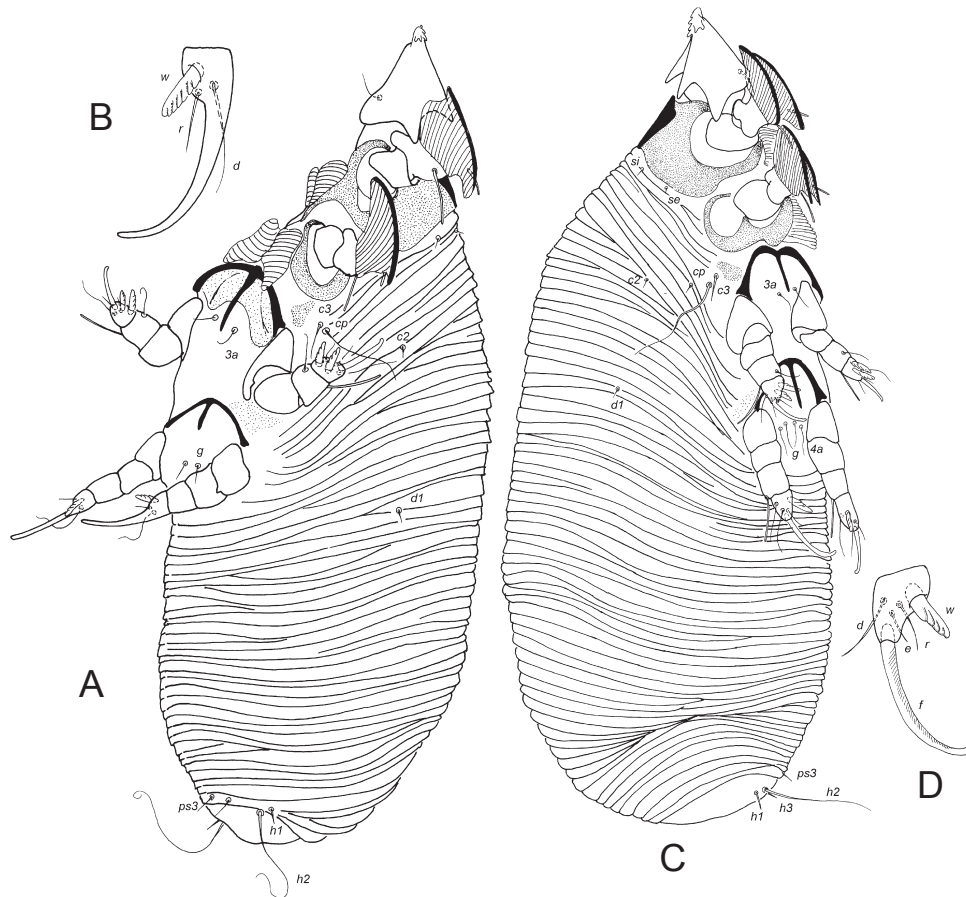


Fig. 40. *Alabidocarpus* sp., male nymphs: A — protonymph in lateral view, B — tarsus IV of protonymph in ventral view, C — tritonymph in lateral view, D — tarsus IV of tritonymph in ventral view.

tae *vFI*, II absent. Legs I and II, including flaps, distinctly shorter than legs III. Coxal fields II not striated. *Males*: Paranal suckers present or more rarely absent *Labidocarpinae* Gunther, 1942 4
 — *Both sexes*: Apodemes Ib and IIa fused. Setae *vFI*, II present. Legs I and II, including flaps, only slightly shorter than legs III. Coxal fields II striated. *Males*: Paranal suckers absent *Chirodiscinae* Trouessart, 1892 (*Chirodiscus* Trouessart et Neumann, 1890)
 4. *Both sexes*: Idiosoma compressed laterally. Ambulacra absent on tarsi III and IV if present situated in subapical position. Setae *fIII* and IV situated apically, claw-like. Legs III and IV consisting of 4 or more rarely 5 articulate segments *Labidocarpini* Fain, 1971 7
 — *Both sexes*: Idiosoma subcylindrical. Ambulacra present on tarsi III and IV, situated apically. Setae *fIII* and IV situated subapically, not claw-like. Legs III and IV consisting 5 articulate segments *Schizocarpini* Fain, 1971 5
 5. *Both sexes*: Setae *scx* absent 6
 — *Both sexes*: Setae *scx* present *Schizocarpus* Trouessart, 1896

6. *Both sexes*: Idiosoma slightly flattened dorso-ventrally. Setae *d2* present. Tarsus III with 2 ventral thickened serrate setae *Lutrilichus* Fain, 1970
 — *Both sexes*: Idiosoma spindle-shaped. Setae *d2* absent. Tarsus III with 1 ventral thickened serrate seta *Soricilichus* Fain, 1970
 7. *Both sexes*: Tarsus III with or without ambulacral disc. Tarsus IV without ambulacral disc 8
 — *Both sexes*: Tarsi III–IV with ambulacral disc in both sexes *Paralabidocarpus* Pinichpongse, 1963
 8. *Both sexes*: Tarsus III without ambulacral disc ... 9
 — *Both sexes*: Tarsus III with ambulacral disc *Labidocarpoides* Fain, 1970
 9. *Both sexes*: Ambulacral stalks absent or vestigial on tarsi III–IV or present on tarsi III–IV of females and on tarsus III of males 10
 — *Both sexes*: Ambulacral stalks present on tarsi III–IV 16
 10. *Both sexes*: Ambulacral stalk of tarsi III–IV cylindro-conical and much shorter than apical claw-like seta of tarsi 11
 — *Both sexes*: Ambulacral stalk of tarsi III–IV inflated and as long as apical claw-like seta of tarsi *Megadermicolus* Fain, 1971

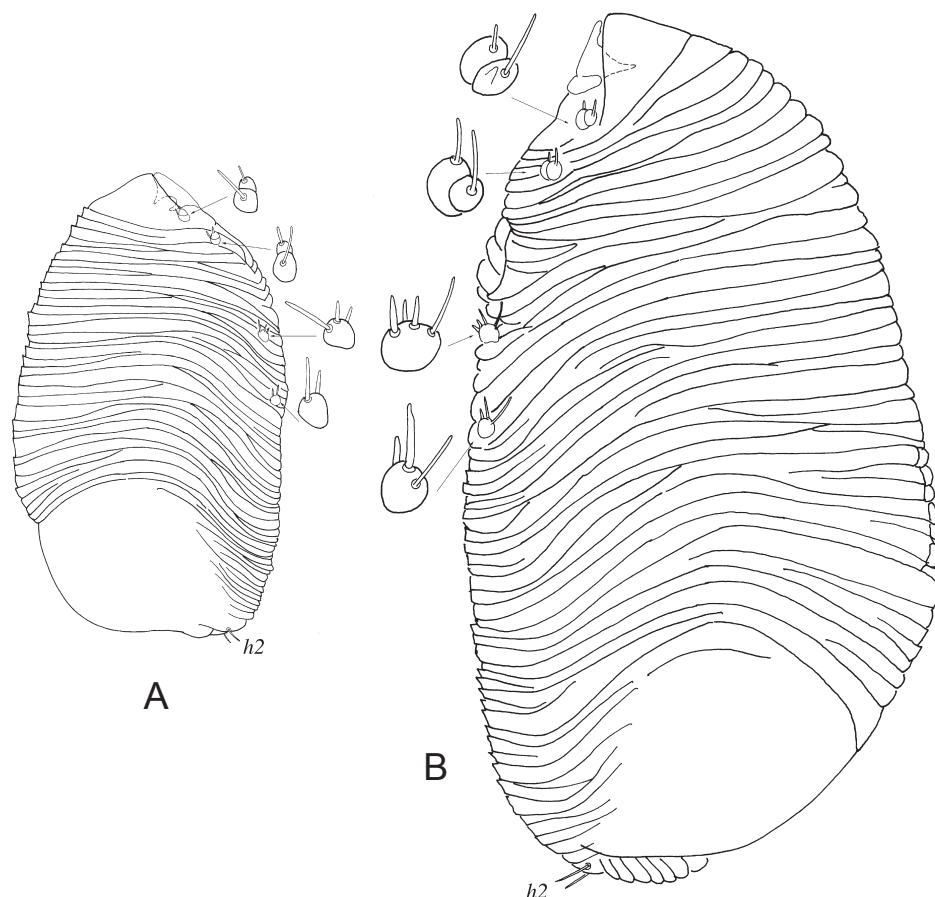


Fig. 41. *Alabidocarpus* sp., female nymphs: A — protonymph in lateral view, B — tritonymph in lateral view.

11. *Both sexes*: Cuticular crest absent posterior to prescapular shield 12
 — *Both sexes*: Cuticle behind prescapular shield sur-elevated with numerous striations close together, forming median crest
 *Olabidocarpus* Lawrence, 1948
 12. *Both sexes*: Prescapular shield without posterior paramedian lobes 13
 — *Both sexes*: Posterior margin of prescapular shield with 4 lobes: 2 paramedian triangular form and 2 lateral more rounded (in *Dentocarpus notopterus*, lateral lobes absent). Postscapular shield(s) present or absent
 *Dentocarpus* Dusbabek et Cruz, 1966
 13. *Both sexes*: Prescapular shield well developed, longer than wide 15
 — *Both sexes*: Prescapular shield poorly developed, wider than long 14
 14. *Both sexes*: Apical claw-like seta of tarsus IV (*d*) without membrane
 *Parakosa* McDaniel et Lawrence, 1962
 — *Both sexes*: Apical claw-like seta of tarsus IV (*d*) with large membrane *Trilabidocarpus* Fain, 1970
 15. *Both sexes*: Very large median postscapular shield present, wider and longer than prescapular

shield, prescapular shield with straight posterior margin *Adentocarpus* Fain, 1972
 — *Both sexes*: 2 narrow paramedian postscapular shields present, straight or L-shaped. Posterior margin of prescapular shield straight or concave ..
 *Labidocarpellus* Fain, 1976
 16. *Both sexes*: Ambulacral stalk on tarsi III–IV vestigial or completely absent 18
 — *Female*: Ambulacral stalk present on tarsi III–IV. *Male*: Ambulacral stalk present on tarsi III
 17
 17. *Both sexes*: Medio-dorsal cuticular crest absent behind prescapular shield; large postscapular shields absent *Labidocarpus* Trouessart, 1895
 — *Both sexes*: Medio-dorsal cuticular crest present behind prescapular shield; 2 large postscapular shields present *Asiolabidocarpus* Fain, 1972
 18. *Both sexes*: 1 median prescapular shield present. Ambulacral stalk of tarsi III–IV absent 19
 — *Both sexes*: 2 paramedian prescapular shields present. Ambulacral stalk of tarsi III–IV vestigial or absent *Pteropiella* Fain, 1970
 19. *Both sexes*: Legs IV equal or subequal to legs III. Prescapular shield without posterior incision ..
 20

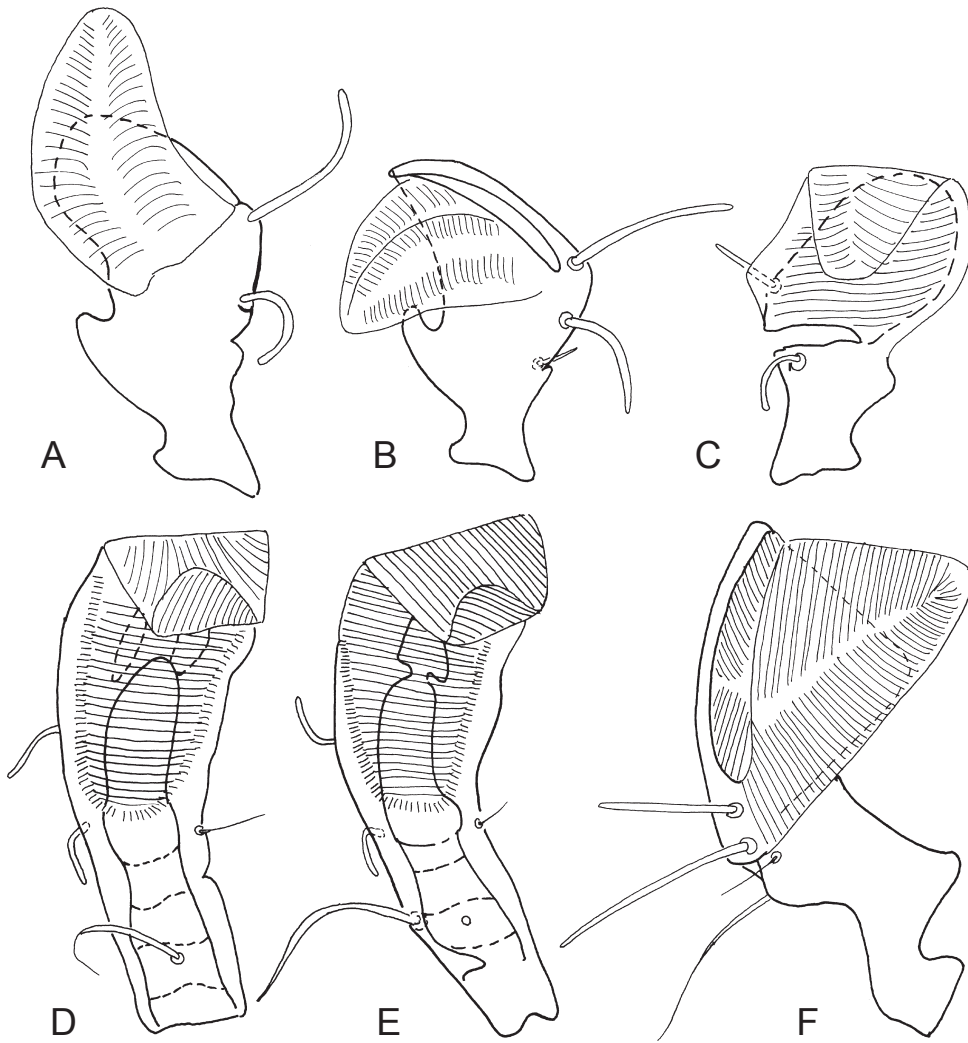


Fig. 42. Chirodiscidae, anterior legs: *Labidocarpus laviae* Fain, 1976 (A–C): A — female leg I in dorsal view, B — same, male, C — same, male leg II; *Chirodiscus amplexans* Trouessart et Neumann, 1889 (D, E): D — female leg I of in ventral view, E — same, leg II; F — male leg I of *Schizocarpus* sp. in dorsal view (after Fain (1971), with minor modifications).

— *Both sexes*: Legs IV abnormally long and with tarsi elongate, much longer than wide. Prescapular shield with posterior margin strongly incised forming 4 long lobes *Afrolabidocarpus* Fain, 1970 20. *Both sexes*: Tarsi III with 2 ventral striated thickened setae (*s* and *w*) 22
 — *Both sexes*: Tarsi III with 1 ventral striated thickened seta (*w*) 21
 21. *Both sexes*: Setae *si* and *c3* absent or alveoli *Lawrenceocarpus* Dusbabek et Cruz, 1966
 — *Both sexes*: Setae *si* and *c3* present, normally developed .. *Paralawrenceocarpus* Guerrero, 1992 22. *Both sexes*: Apical claw-like seta of tarsi III and IV not bifid. Setae *se* present, *c3* filiform 23
 — *Both sexes*: Apical claw-like seta of tarsi III and IV bifid. Setae *se* absent, *c3* alveoli *Eurolabidocarpus* Fain et Aellen, 1994 23. *Both sexes*: Chelicerae not elongated, shorter than leg I, including membranous flaps 24

— *Both sexes*: Chelicerae strongly elongated, longer than leg I, including membranous flaps (Primates) *Rynconyssus* Fain, 1967 24. *Both sexes*: Setae *si* microsetae or alveoli. Bases of setae *c2* situated at level or slightly posterior or anterior to seta *cp* bases. Coxal fields III adjoining to each other *Alabidocarpus* Ewing, 1929
 — *Both sexes*: Setae *si* long. Bases of setae *c2* situated far posterior to bases of setae *cp*. Coxal fields III separated from each other by striated cuticle *Pseudoalabidocarpus* McDaniel, 1972

**Family Listrophoridae
 Megnin et Trouessart, 1884**

Type genus: *Listrophorus* Pagenstecher, 1862
 Table 10, Figs. 43–53

Diagnosis. *Both sexes*. Cheliceral hood and ventral apophysis of movable digit present. Palps covered by palpal membranes. Body subcylindri-

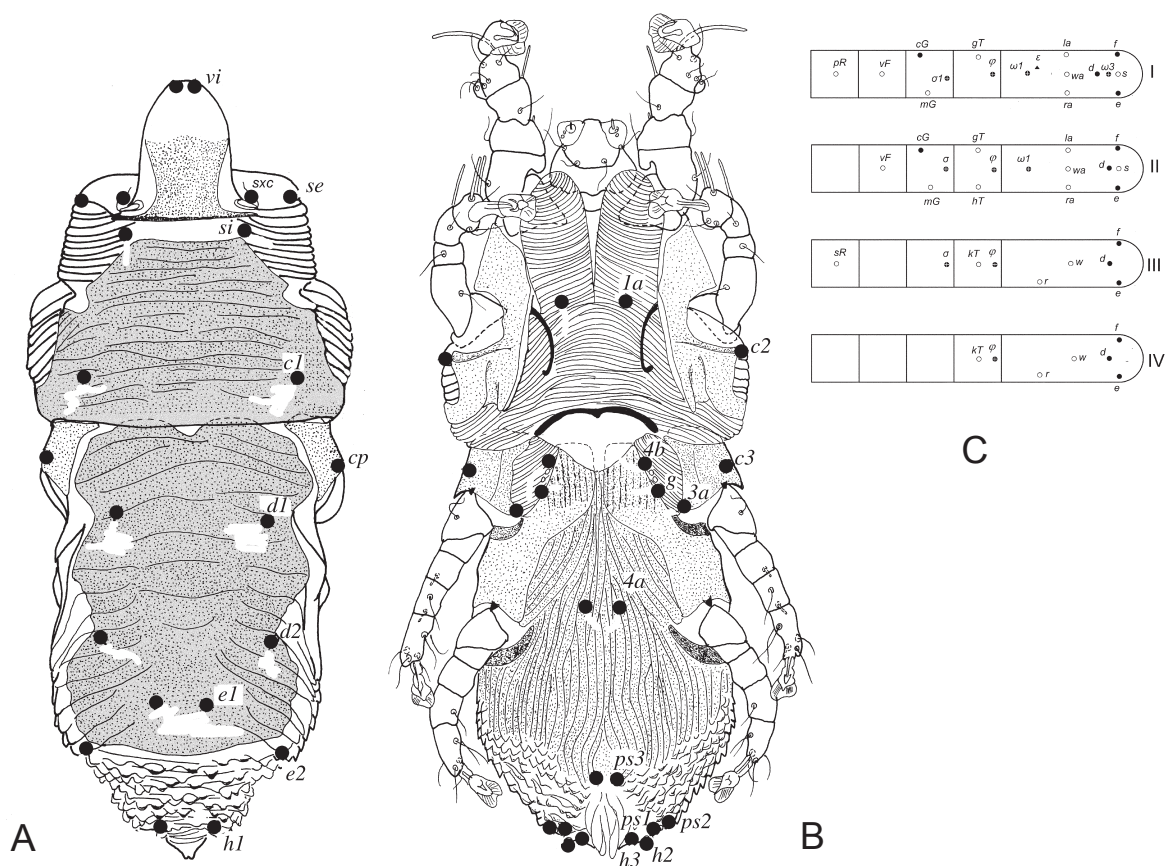


Fig. 43. Listrophoridae, scheme of setation: A — idiosoma in dorsal view, B — same in ventral view, C — legs I-IV.

cal (Listrophorinae) or flattened dorso-ventrally (Aplodontochirinae). Supracoxal sclerite and supracoxal opening distinct, setae *scx* absent (reversed in some species). Propodoonotal shield subdivided onto prescapular and postscapular shield (absent in some genera). Prescapular shield completely gnathosoma dorsally (Listrophorinae) or not covering palpal extremities (Aplodontochirinae). Genital papillae present. Membranes between coxal fields developed, modified into attaching flaps; pair of auxiliary striated membranes situated between coxal fields II. Legs without clasping organs. Setae *ba*I, II absent. Solenidia σ I-III absent (Listrophorinae) or present (Aplodontochirinae). Ambulacral sclerites distinct.

Female. Opisthogastral shields absent. Ovipore Y-shaped. Epigynum present, usually fused with apodemes IIIa. Anal opening situated terminally or ventrally. Oviparous.

Male. Opisthosomal lobes and paranal suckers usually present. Legs III and IV normally developed. Setae *d*IV and *e*IV filiform or stick-like.

Taxa included: 20 genera and 167 species in 2 subfamilies, Listrophorinae Megnin et Trouesart, 1884 — *Listrophorus* Pagenstecher, 1862 (19 species), *Aeromychirus* Fain, 1972 (3 species), *Af-*

rolistrophorus Fain, 1970, *Afrolistrophorus* Fain, 1970 (33 species), *Amlistrophorus* Fain, 1981 stat. nov. (4 species), *Asiochirus* Fain, 1970 (4 species), *Carnilistrophorus* Fain, 1980 (5 species), *Dubininetta* Fain et Lukoschus, 1978 (3 species), *Echinorella* Fain, 1980 (1 species), *Geomylichus* Fain, 1970 (28 species), *Hemigalichus* Fain, 1970 (2 species), *Leporacarus* Fain, 1970 (4 species), *Lynxacarus* Radford, 1951 (11 species), *Metalistrophorus* Fain, 1970 (4 species), *Olistrophorus* McDaniel et Whitaker, 1972 (7 species), *Prolistrophorus* Fain, 1970 (28 species), *Pteromychirus* Fain, 1980 (1 species), *Sciurochirus* Fain, 1972 (3 species), *Sclerolistrophorus* Fain, 1976 (5 species), *Quasilistrophorus* Fain, Whitaker et Lukoschus, 1978 (1 species); Aplodontochirinae — *Aplodontochirus* Fain et Hyland, 1972 (1 species).

Associations with hosts: inhabitants of fur of therian mammals, Listrophorinae — Marsupialia: Paucituberculata (Caenolestidae); Placentalia: Carnivora (Felidae, Mustelidae, Viverridae), Lagomorpha (Leporidae), Macroscelidea (Macroscelidae), Pilosa (Bradypodidae), Primates (Tarsiidae), Rodentia (Cricetidae, Ctenodactylidae, Ctenomyidae, Dipodidae, Echimyidae, Geomyiidae, Geteromyidae, Gliridae, Heteromyidae, Mu-

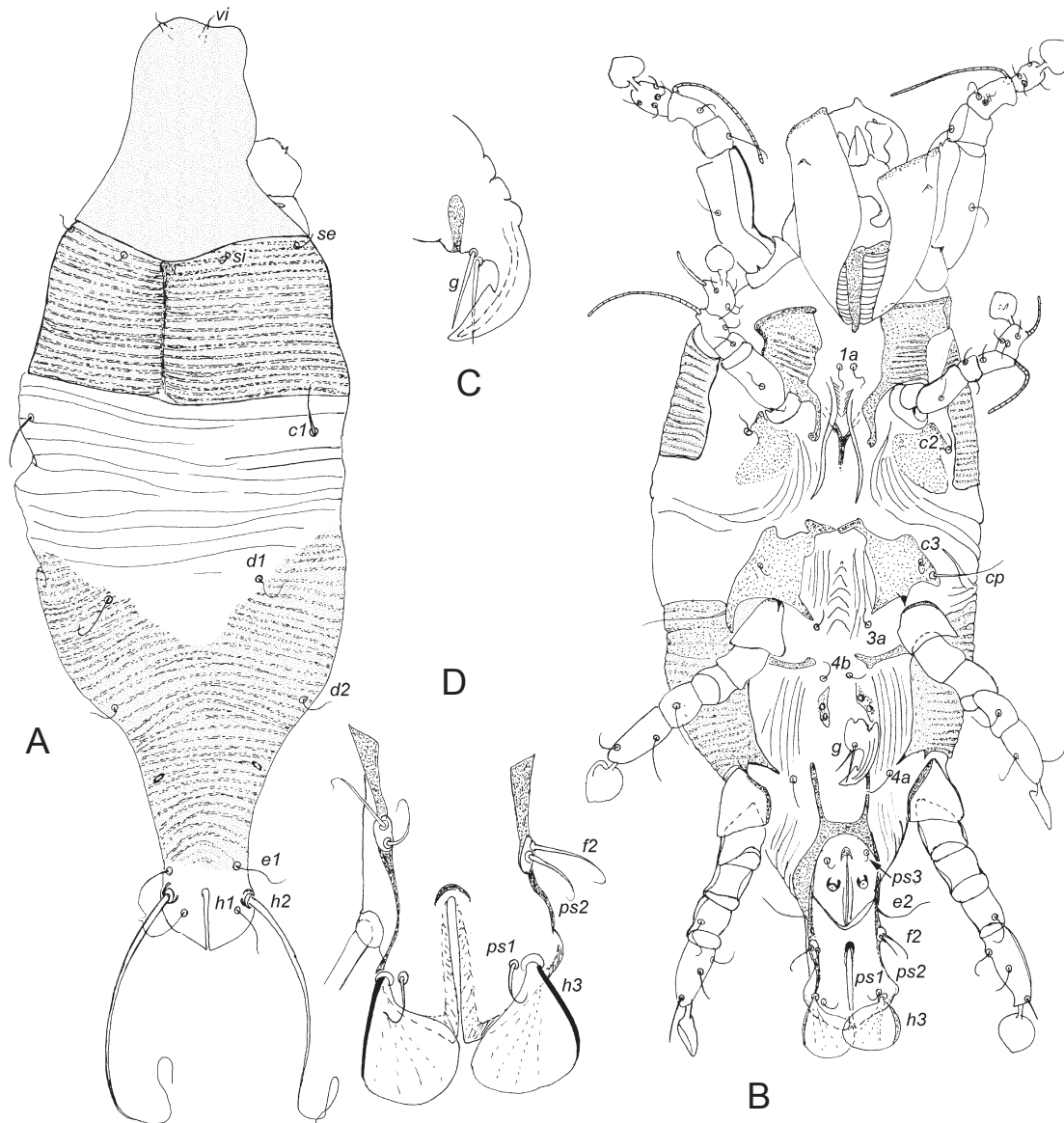


Fig. 44. *Afrolistophorus mediolineatus nesokia* (Fain et Hyland, 1980), male: A — dorsal view, B — ventral view, C — aedeagus, D — opisthosoma in ventral view.

ridae, Nesomyidae, Sciuridae, Spalacidae), Scandentia (Tupaiidae), Soricomorpha (Soricidae, Talpidae); Aplodontochirinae — Rodentia (Aplodontiidae).

I believe that set of initial host orders of listrophorins includes only four orders, Erinaceomorpha, Rodentia, Scandentia, and Soricomorpha. Ancestral carnivores probably received mites from their preys (most species of the genera *Lynxacarus* and *Carnilistophorus*).

The associations of listrophorins with marsupial hosts are secondary — mites of the genus *Prolistophorus* were probably switched to *Lestoros inca* (Caenolestidae) from South American rodents which are hosts for most species of this genus. In addition, *Amlistophorus inca* (Fain,

1976) and *Amlistophorus venezuelensis* (Fain et Lukoschus, 1983) comb. nov. are associated with marsupial hosts, *Lestoros inca* and *Monodelphis brevicaudata* (Didelphidae), respectively. Other two species of the genus *Amlistophorus* are associated with rodents. It is highly probably, therefore, that mites of this genus were also secondarily switched from rodents to these marsupials. It should be mentioned that Fain and Lukoschus (1983) assumed that *Monodelphis brevicaudatus* is an accidental host for this species because only one specimen of this species was collected from the alcohol preserved host in museum.

Only one species, *Carnilistophorus rhynchocyoni* (Fain, 1970) is recorded from Macroscelidea, however, it is possible that additional ex-

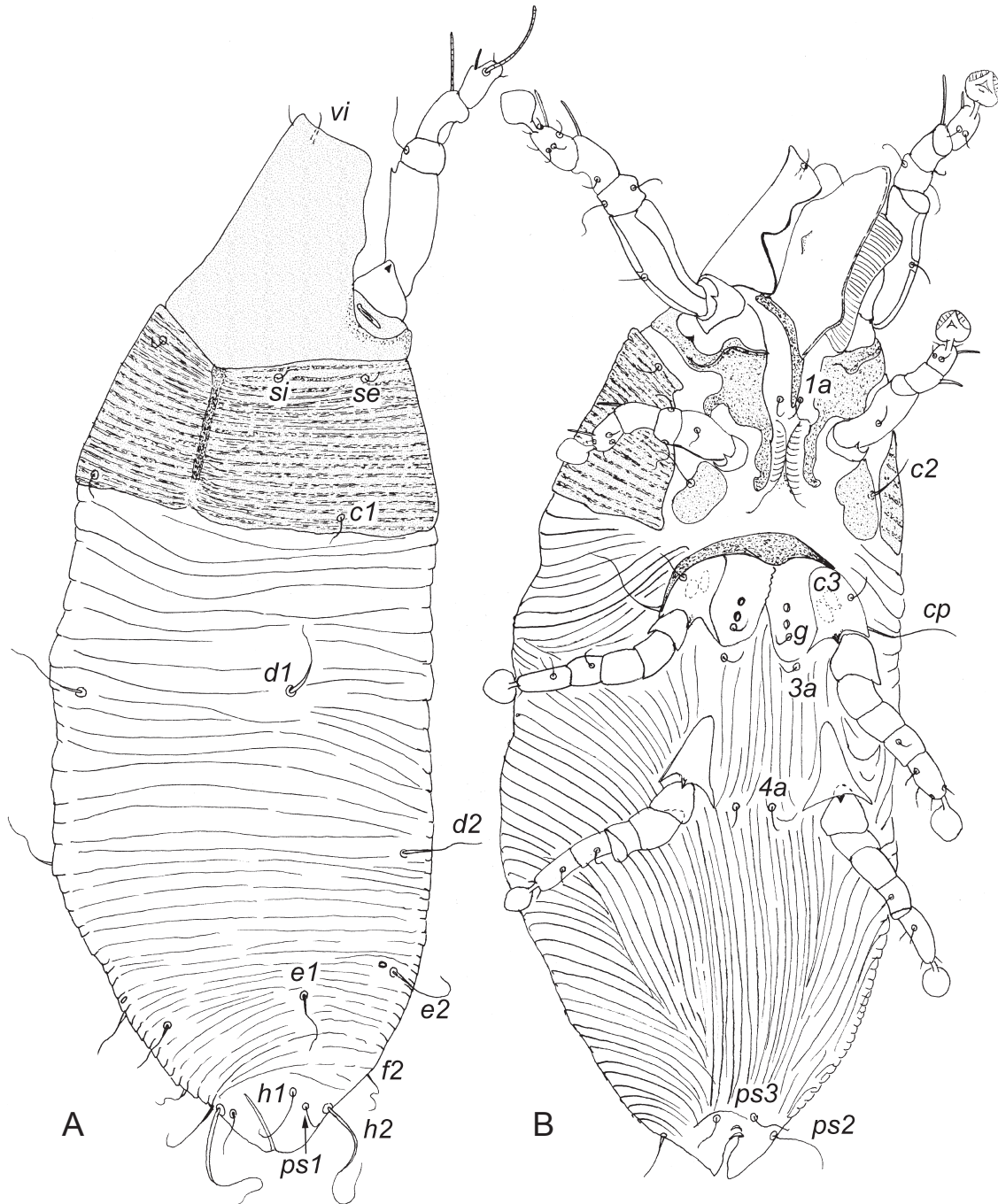


Fig. 45. *Afrolistrophorus mediolineatus nesokia* (Fain et Hyland, 1980), female. A — dorsal view, B — ventral view.

aminations of these hosts allow collecting more listrophorines. Other species of this genus are associated with Mustelidae and Viverridae from Africa (4 species) and Spalacidae from Asia (1 species).

Finally, two records, *Afrolistrophorus medius* Fain et Lukoschus, 1983 from *Tarsius pumilus* (Tarsiidae) and *Afrolistrophorus bradypus* Fain, 1981 from *Bradipus tridactylus* (Bradypodidae), are also probably the results of secondary shifts from rodents because other 31 species of this genus are associated exclusively with rodents.

Distribution. Listrophorinae — Eurasia, Africa (excluding Madagascar), North and South America; Aplodontochirinae — North America.

Remarks. (i) The genus *Amlistrophorus* Fain, 1981 stat. nov was described as a subgenus of the genus *Afrolistrophorus* (Fain 1981b). Both sexes of this subgenus differ from *Afrolistrophorus* by the presence of the lateral expansion situated in the anterior part of the prescapular shield, mushroom-shaped (vs absence of such expansions in *Afrolistrophorus*); in males, the hysteronotal shield is paired and setae *h3* are filiform (vs unpaired hystero-

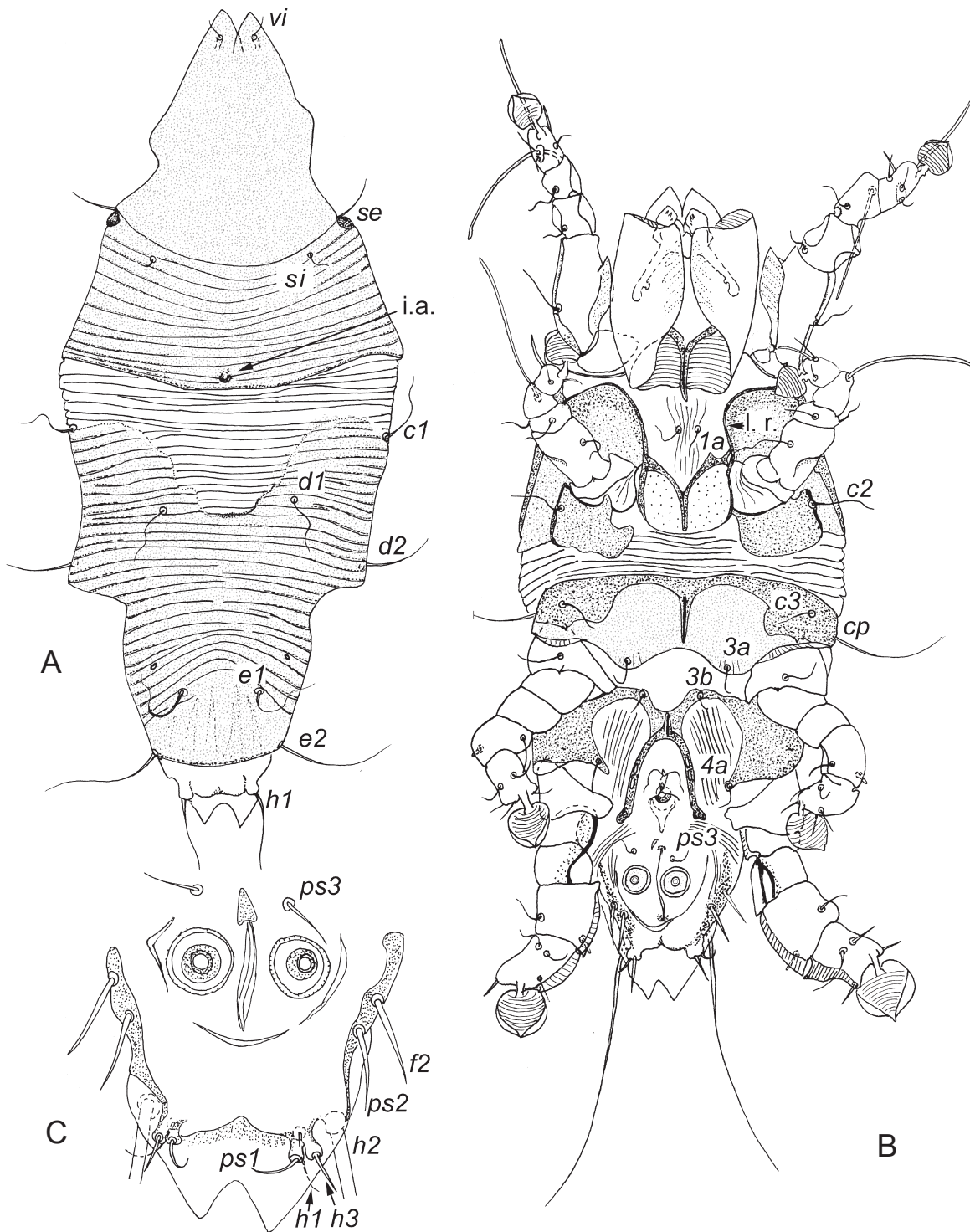


Fig. 46. *Hemigalichus chrotogale* Bochkov et OConnor, 2005, male: A — dorsal view, B — ventral view, C — opisthosoma, ventral view (after Bochkov and OConnor (2005b), with minor modifications).

onotal shield in males and foliate setae *h3* in *Afrolistrophorus*); in females, the anterior margin of the hysteronotal shield is situated at level or posterior to the level of leg IV insertion (vs anterior of legs IV insertion in *Afrolistrophorus*). I believe that this subgenus deserves the full generic status.

(ii) The genus *Spalacarus* Fain, 1980 was described from species resembles representatives of

the genus *Afrolistrophorus* but differ from them by the absence of the hysteronotal shield in females and the slightly widened male idiosoma (Fain 1980b). Later on, Fain (1981b) established subgenus *Teinolistrophorus* Fain, 1981 within the genus *Spalacarus* for two species with the elongated idiosoma and widely separated legs III and IV. In my opinion, the species of the genera *Afrol-*

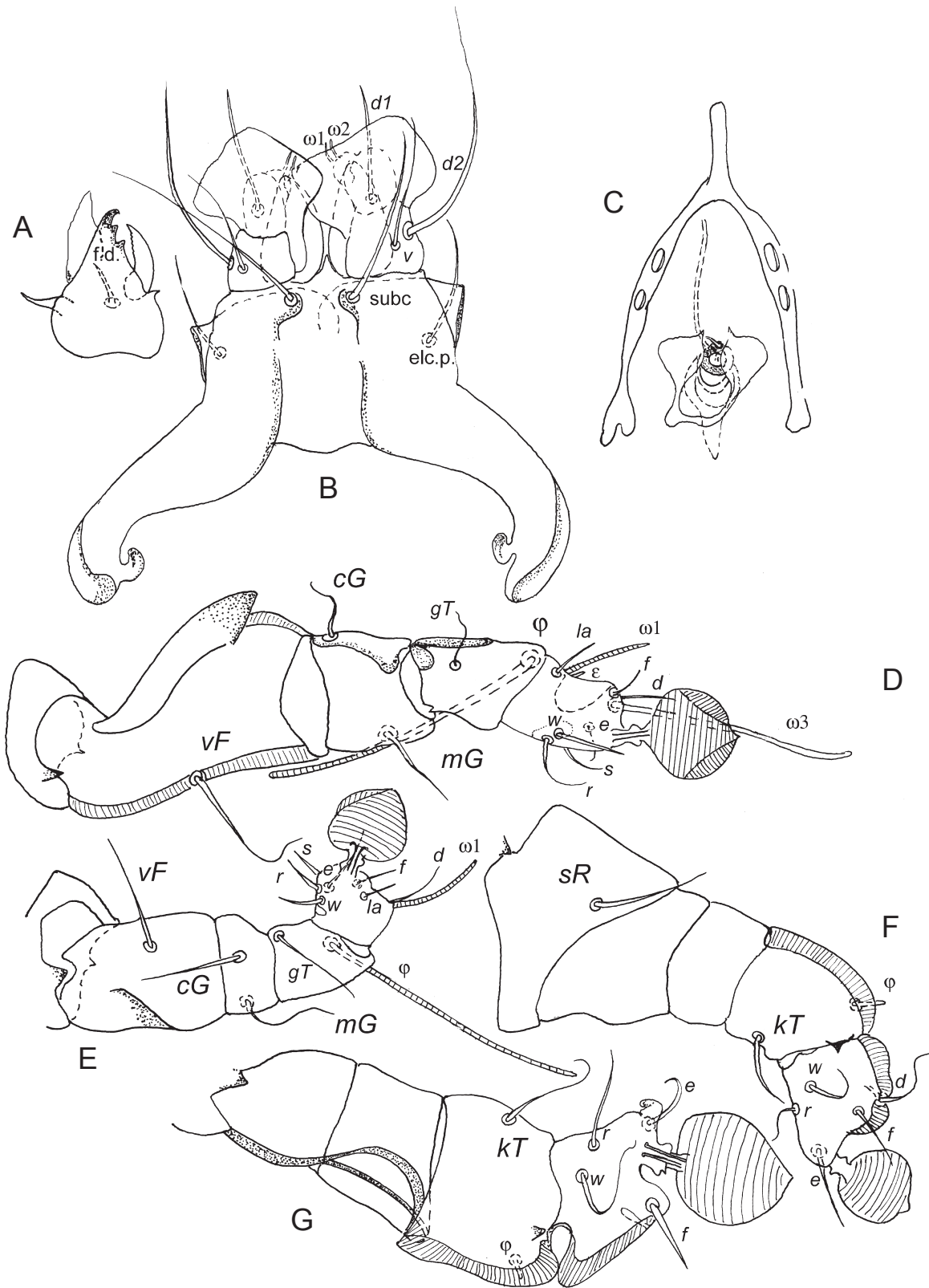


Fig. 47. *Hemigalichus chrotogale* Bochkov et OConnor, 2005, details of male. A — chelicera in lateral view, B — gnathosoma, C — genital organ, D–F — legs I–IV in ventral view, respectively (after Bochkov and OConnor (2005b), with minor modifications).

istrophorus and *Spalacarus* are not principally different and all distinct features of *Spalacarus* should be treated as the subgeneric ones, because

in some other listrophorid genera (Fain 1981b) they are treated as subgeneric. I consider *Spalacarus* stat. nov. as the subgenus of the genus *Afrol-*

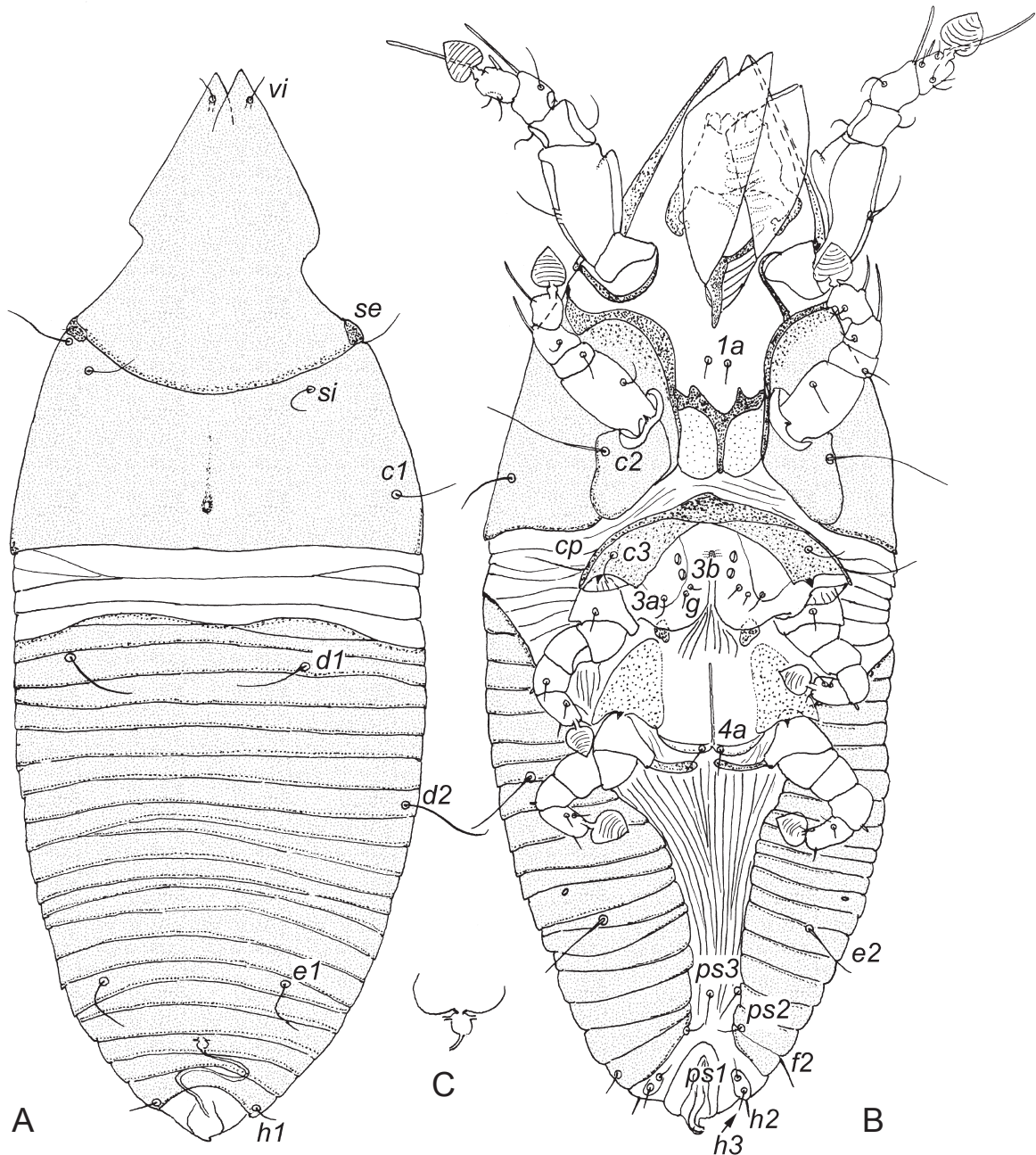


Fig. 48. *Hemigalichus chrotogale* Bochkov et OConnor, 2005, female: A — dorsal view, B — ventral view, C — spermatheca (after Bochkov and OConnor (2005b), with minor modifications).

istrophorus and synonymize the subgenus *Teinolistrophorus* syn. nov. with the subgenus *Spalacarus*.

(iii) The subgenus *Mexicochirus* Fain et Estebanes, 1996 (2 species) was described in the genus *Asiochirus* (Fain and Estebanes 1996). Fain and Bochkov (2003) compared characters of the closely related genera *Asiochirus* and *Olistrophorus*, redefined these genera and recombined some *Asiochirus* species. However, the subgenus *Mexicochirus* was not considered in this paper. Meantime according to their characteristics (mainly the mushroom-shaped anterior part of the prescapular

shield) it should be included in the genus *Olistrophorus* in sense of Fain and Bochkov (2003) or even considered as a separate genus due to some peculiar features of its species (the presence of opisthosomal lobe(s) and the postscapular shield fused with the prescapular shield). I tentatively include this subgenus in the genus *Olistrophorus*.

(iv) *Afrolistrophorus venezuelensis* Fain et Lukoschus, 1983 is known from a single male only (Fain and Lukoschus 1983) whose characters mostly correspond to male characteristics of the

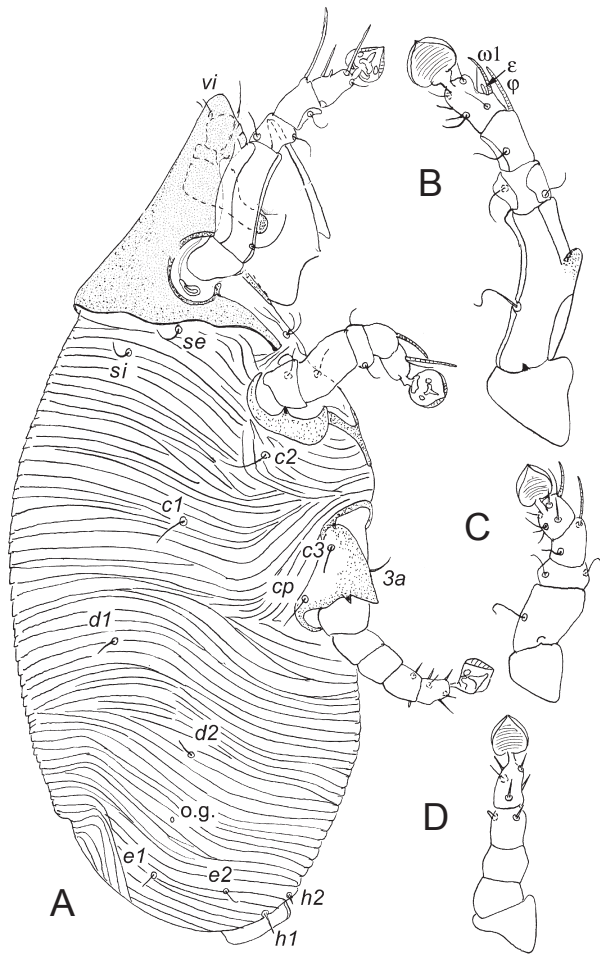


Fig. 49. *Hemigalichus chrotogale* Bochkov et OConnor, 2005, larva: A — lateral view, B–D — legs I–III in ventral view, respectively (after Bochkov and OConnor (2005b), with minor modifications).

genus *Amlistrophorus*, i.e. mushroom-shaped anterior part of the prescapular shield and filiform setae *h3*. For these reasons I include this species, *Amlistrophorus venezuelensis* (Fain et Lukoschus, 1983) comb. nov. to the genus *Amlistrophorus*.

Main references. Fain (1971) — revision of African Listrophoridae, Fain, 1973 — revision of South American Listrophoridae, Fain and Hyland (1974) — review of North American Listrophoridae, Fain (1978) — revision of the genus *Lynxacarus*, Fain (1979b) — revision of the genera *Aeromychirus* and *Sciurochirus*, Fain (1980) — notes on some little-known listrophorid genera and species, Fain (1981b) — key to listrophorin genera, Fain (1981c) — new and little-known taxa of Listrophoridae, OConnor (1982) — diagnosis of Listrophoridae and references, Fain and Lukoschus (1984) — review of the genus *Prolistrophorus*, Wurst (1993) — morphology of *Listrophorus leuckarti*, Fain and Estebanes (2000) — a review of the genus *Geomylichus*, Bochkov and OConnor

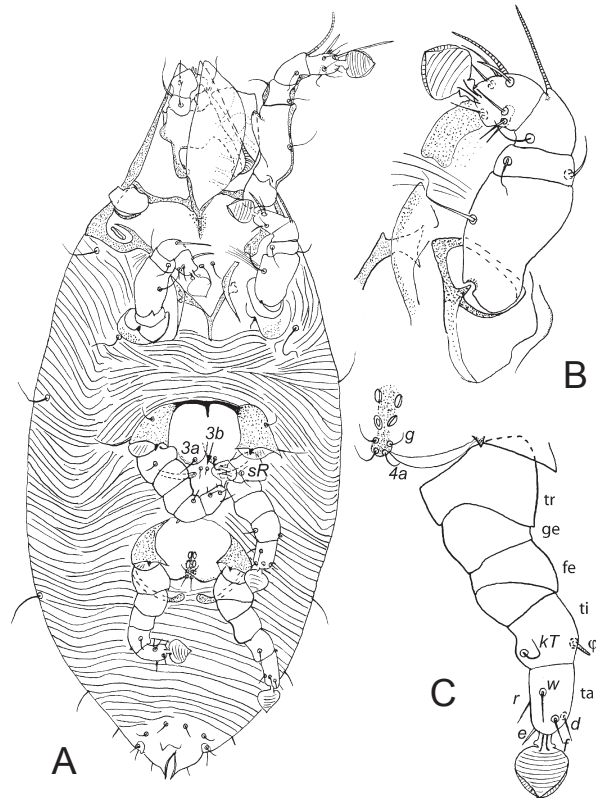


Fig. 50. *Hemigalichus chrotogale* Bochkov et OConnor, 2005, protonymph: A — dorsal view, B — ventral view, C — leg IV in ventral view (after Bochkov and OConnor (2005b), with minor modifications).

(2005b) — comparative observations on listrophorid morphology.

Key to the family Listrophoridae Megnin et Trouessart, 1884

(partly based on Fain [1981b])

1. *Both sexes*: Prescapular shield covering palpal extremities. Folds (claspings flaps) between coxal fields I curved. Solenidia σ I–III absent Listrophorinae Megnin et Trouessart, 1884 2
2. *Both sexes*: Prescapular shield not covering palpal extremities. Folds (claspings flaps) between coxal fields I not curved. Solenidia σ I–III present Aplodontochirinae Fain et Hyland, 1972, *Aplodontochirus* Fain et Hyland, 1972
2. *Both sexes*: Postscapular shield present (in *Pteromychirus* it reduced to narrow transverse band) .. 5
- *Both sexes*: Postscapular shield absent or strongly reduced to narrow longitudinal band ... 3
3. *Both sexes*: Fine striations behind prescapular shield separated by sclerotized transverse bands. *Male*: Setae *h3* foliate 4
- *Both sexes*: Fine striations behind prescapular shield not separated by sclerotized transverse

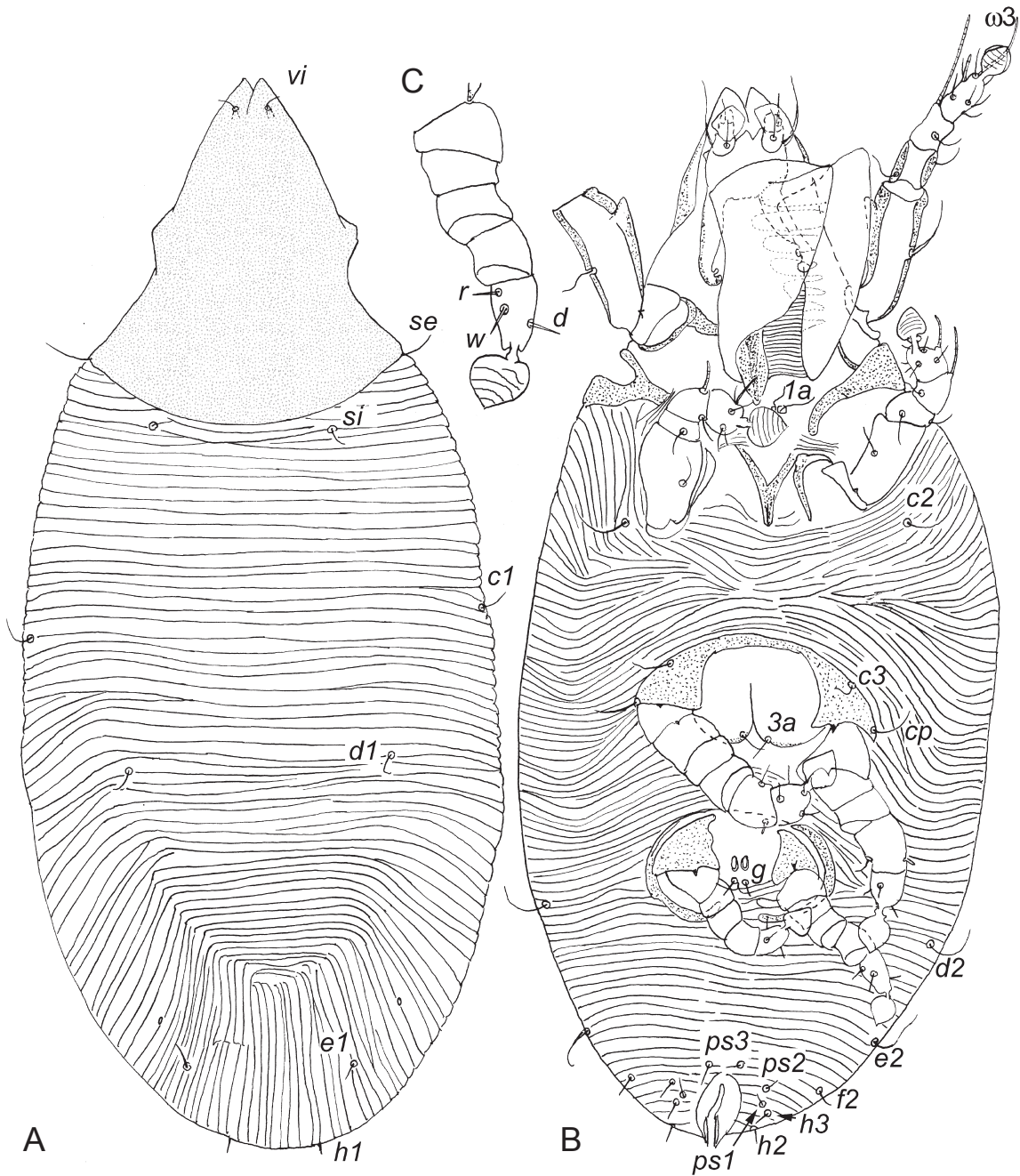


Fig. 51. *Hemigalichus chrotogale* Bochkov et OConnor, 2005: tritonymph: A — ventral view, B — coxa of leg II, C — leg IV in ventral view (after Bochkov and OConnor (2005b), with minor modifications).

bands. *Male*: Setae *h3* filiform
 *Carnilistrophorus* Fain, 1980
 4. *Both sexes*: Dorsal part of idiosoma behind pre-
 escapular shield distinctly sclerotized and punc-
 tured. Transverse striations of idiodorsum distinctly
 separated from each other. *Male*: Setae *h3*
 rounded or bilobate *Sciurochirus* Fain, 1972
 — *Both sexes*: Dorsal part of idiosoma behind pre-
 escapular shield weakly sclerotized and punctured.
 Transverse striations of idiodorsum situated very
 close to each other and numerous. *Male*: Setae *h3*
 triangular *Metalistrophorus* Fain, 1970

5. *Both sexes*: Postscapular shield distinctly devel-
 oped, subequal or 3 times shorter than prescapular
 shield 6
 — *Both sexes*: Postscapular shield strongly re-
 duced, 4 times shorter than prescapular shield
 *Pteromychirus* Fain, 1980
 6. *Both sexes*: Postscapular shield unpaired 7
 — *Both sexes*: Postscapular shield paired
 *Listrophorus* Pagenstecher, 1861
 7. *Both sexes*: Postscapular shield without median
 unsclerotized area 8
 — *Both sexes*: Postscapular shield with large median

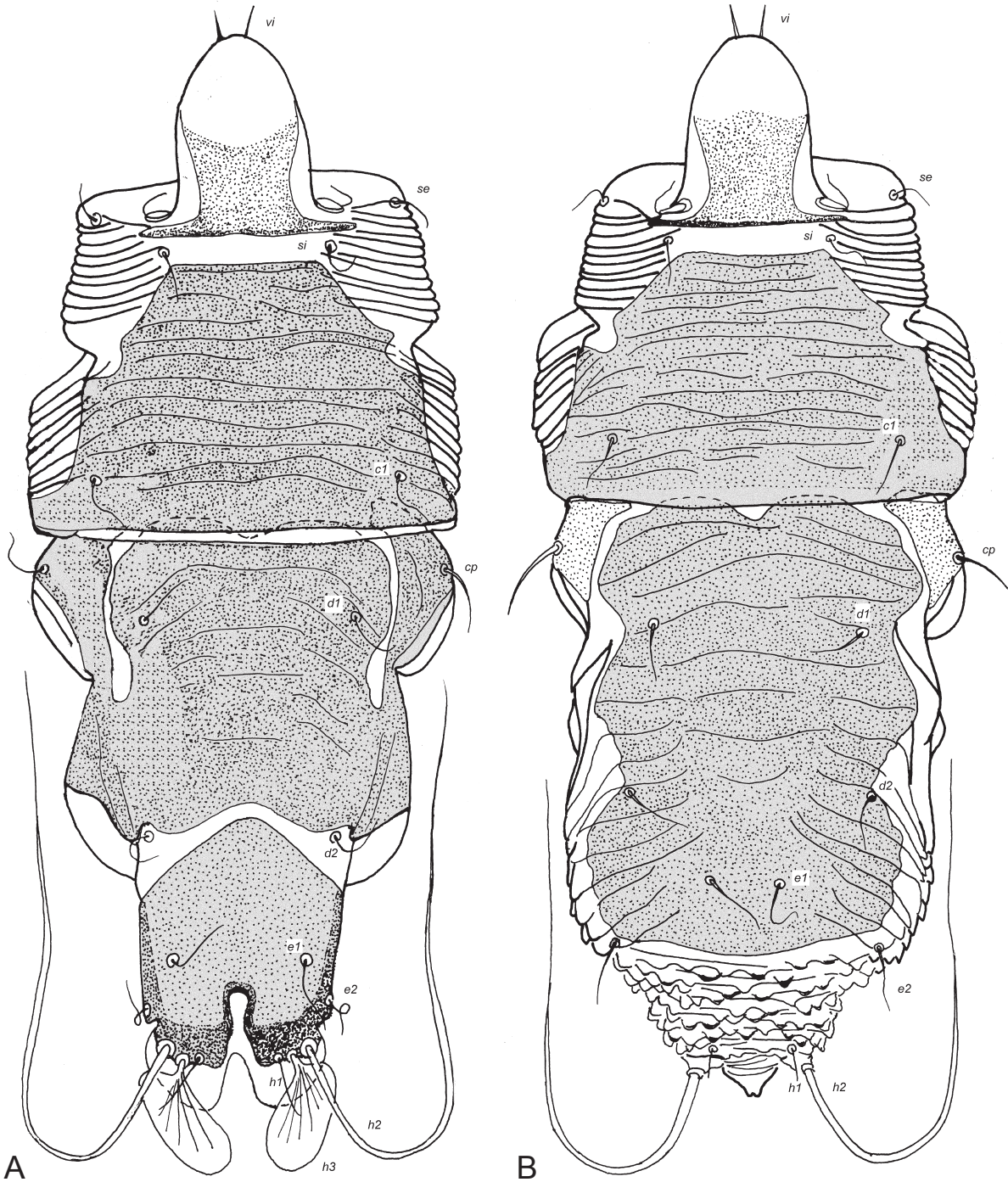


Fig. 52. *Aplodontochirus borealis* Fain et Hyland, 1972: A — male in dorsal view, B — female in dorsal view (after Fain and Hyland (1974), with minor modifications).

unsclerotized area *Prolistrophorus* Fain, 1980
 8. Both sexes: Setae *se* filiform 9
 — Both sexes: Setae *se* spur-like
 *Geomylichus* Fain, 1970
 9. Both sexes: Postscapular shield monotonously
 punctated or covered by transverse bands which not
 interrupted by punctuated longitudinal bands 10
 — Both sexes: Postscapular shield covered by
 transverse bands which interrupted by pair of
 paramedian longitudinal punctuated bands

..... *Quasilistrophorus*
 Fain, Whitaker et Lukoschus, 1978
 10. Both sexes: Propodeonotal and hysteronotal
 shields not completely covering idiosomal dorsum
 11
 — Both sexes: Propodeonotal and hysteronotal
 shields completely covering idiosomal dorsum
 *Sclerolistrophorus* Fain, 1976
 11. Both sexes: Anterior margin of prescapular
 shield without median incision 12

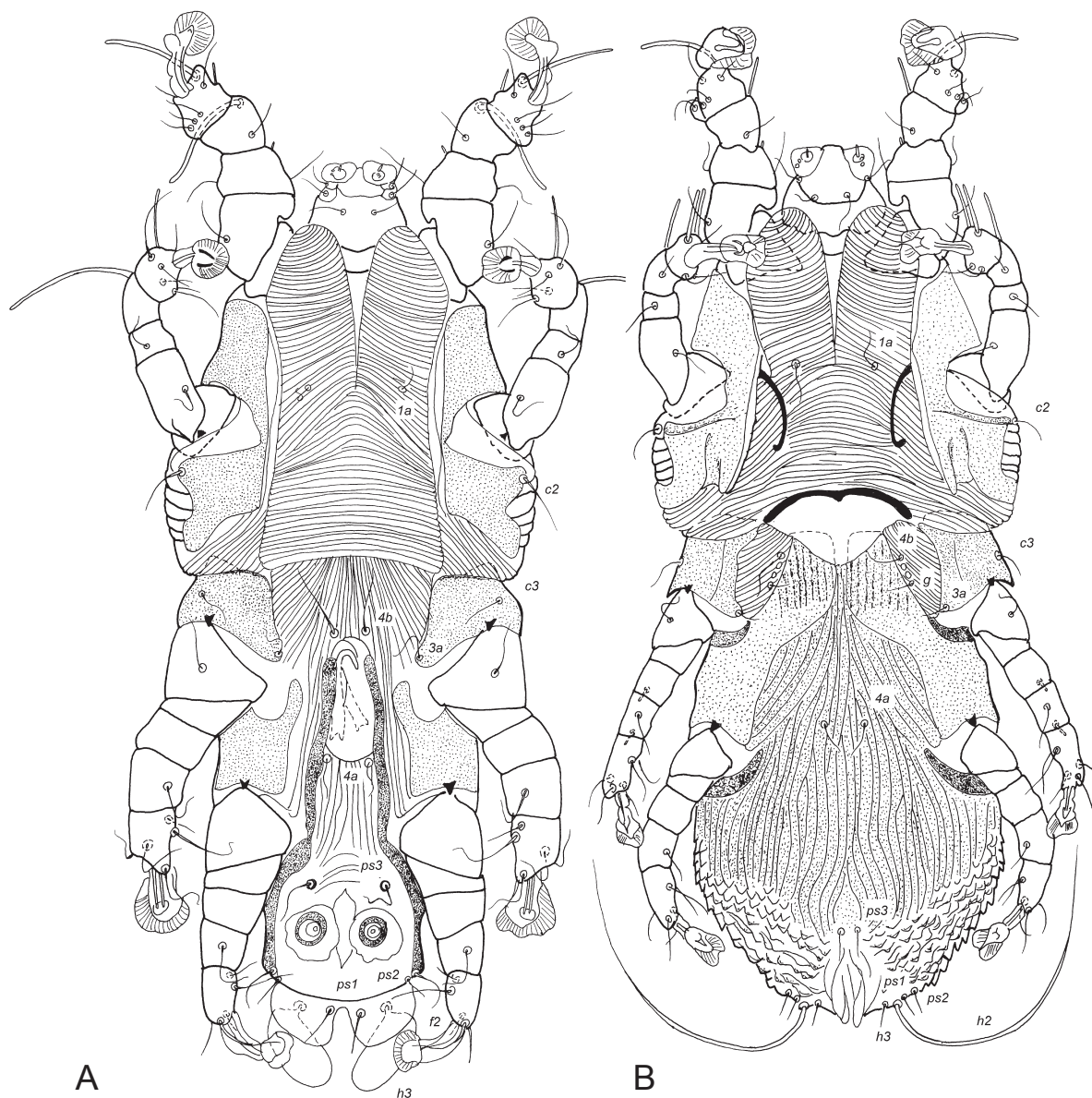


Fig. 53. *Aplodontochirus borealis* Fain et Hyland, 1972: A — male in ventral view, B — female in ventral view (after Fain and Hyland (1974), with minor modifications).

— *Both sexes*: Anterior margin of prescapular shield with deep median incision
 *Hemigalichus* Fain, 1970
 12. *Both sexes*: Prescapular shield without antero-lateral projections 14
 — *Both sexes*: Prescapular shield with pair of antero-lateral projections (mushroom-shape) 13
 13. *Both sexes*: Body distinctly elongated, 3.5–4 times longer than wide. *Female*: Hysteronotal shield present and separated from postscapular shield. *Male*: Setae *4b* present. Setae *h2* whip-like, at least 5 times longer than *h3*
 *Amlistrophorus* Fain, 1976 stat. nov.
 — *Both sexes*: Body relatively short, 2–2.5 times longer than wide. *Female*: Hysteronotal shield ab-

sent or present and fused with postscapular shield (subgenus *Mexicochirus* comb. nov.). *Male*: Setae *4b* absent. Setae *h2* short, subequal or 2 times longer than *h3*
 *Olistrophorus* McDaniel et Whitaker, 1972
 14. *Both sexes*: Femur I with dorso-apical tooth in most species. *Female*: Setal bases *h2* and *h3* situated distinctly far from each other, both these setae usually short, subequal to *ps3*. *Male*: Prescapular sclerites weakly developed, free or fused to each other and surrounding aedeagus (*Leporacarus*). Seta *dIV* present (5 setae on tarsus IV) 17
 — *Both sexes*: Femur I without dorso-apical tooth. *Female*: Setal bases *h2* and *h3* contiguous or almost contiguous, both these setae long, at least 4

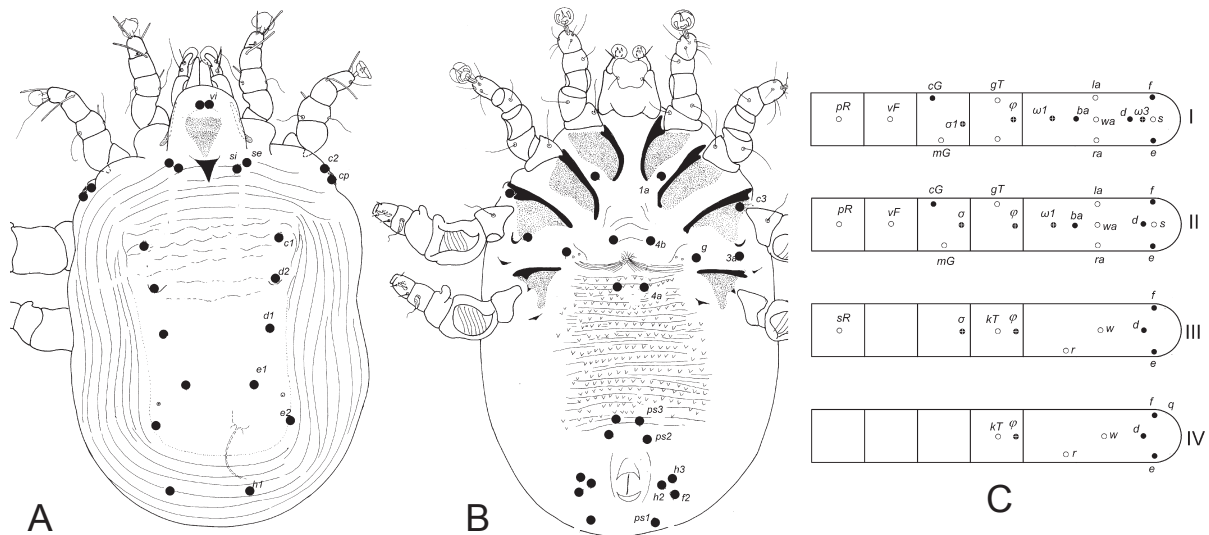


Fig. 54. Myocoptidae, scheme of setation: A — idiosoma in dorsal view, B — same in ventral view, C — legs I-IV.

times longer than *ps3*. *Male*: Prescapular sclerite distinctly developed, fused to each other forming inverted Y-shaped structure. Seta *dIV* absent (4 setae on tarsus IV) 15
 15. *Both sexes*: Setae *4b* present. *Female*: Hysteronotal shield absent 16
 — *Both sexes*: Setae *4b* absent. *Female*: Hysteronotal shield present ... *Echinosorella* Fain, 1980
 16. *Female*: Prescapular shield and bases of apodemes IIa connected by narrow sclerotized band. *Male*: Hysteronotal shield paired
 *Lynxacarus* Radford, 1951
 — *Female*: Prescapular shield and bases apodemes IIa connected by wide lateral plate. *Male*: Hysteronotal shield unpaired
 *Dubinietta* Fain et Lukoschus, 1978
 17. *Female*: Hysteronotal shield present or absent (in latter case scales on opisthosomal venter absent). Setae *ps1* and *ps2* present or absent. *Male*: Setae *h3* foliate. Setae *4b* present. Opisthosomal lobes dorsally distinct 18
 — *Female*: Hysteronotal shield absent. Scales on opisthosomal venter present. Setae *ps1* and *ps2* absent. *Male*: Setae *h3* filiform. Setae *4b* absent. Opisthosomal lobes dorsally indistinct
 *Asiochirus* Fain, 1970
 18. *Female*: Prescapular and postscapular shields separated. Hysteronotal shield present or absent. *Male*: Legs II about 1.5 times shorter than legs I. Pregenital sclerites separated, weakly developed .
 19
 — *Female*: Prescapular and postscapular shields fused. Hysteronotal shield absent. *Male*: Legs I and II subequal in length. Pregenital sclerites fused to each other, surrounding aedeagus
 *Leporacarus* Fain, 1970

19. *Both sexes*: Auxiliary clasping valves weakly developed. *Female*: Hysteronotal shield present or absent (subgenus *Spalacarus* stat. nov.). Setae *4b* absent; setae *ps1* and *ps2* present or absent
 *Afrolistrophorus* Fain, 1970
 — *Both sexes*: Auxiliary clasping valves distinctly developed. *Female*: Hysteronotal shield present. Setae *4b* present; setae *ps1* and *ps2* present
 *Aeromychirus* Fain, 1972

Family Myocoptidae Gunther, 1942

Type genus: *Myocoptes* Claparede, 1869
 Table 11, Figs. 54–63

Diagnosis. *Both sexes*. Cheliceral hood and ventral apophysis of movable digit present. Palpal segments fused dorsally. Idiosoma slightly elongate, dorso-ventrally flattened or subcylindrical (*Trichoecius*). Supracoxal sclerite distinct, supracoxal opening indistinct, setae *scx* absent. Genital papillae present. Intercoxal attaching organs absent. Legs III and IV in females and immature stages and legs III in males modified into clasping organs. Femora of these legs enlarged and ventrally concave, genua distinctly developed; tibia and tarsi devoid of the pretarsi and shortened. Clasping apparatus formed by all these podomeres, excluding trochanters. Tarsi-genital part curving inward and fixings host hair between femoral concavity and these 3 podomeres. Setae *sIII* and famulus ϵ absent. Ambulacral sclerites small but recognizable.

Female. Opisthogastral shields absent. Ovipore Y-shaped. Epigynum present (absent in *Myocoptes*). Anal opening situated terminally or ventrally. Oviparous.

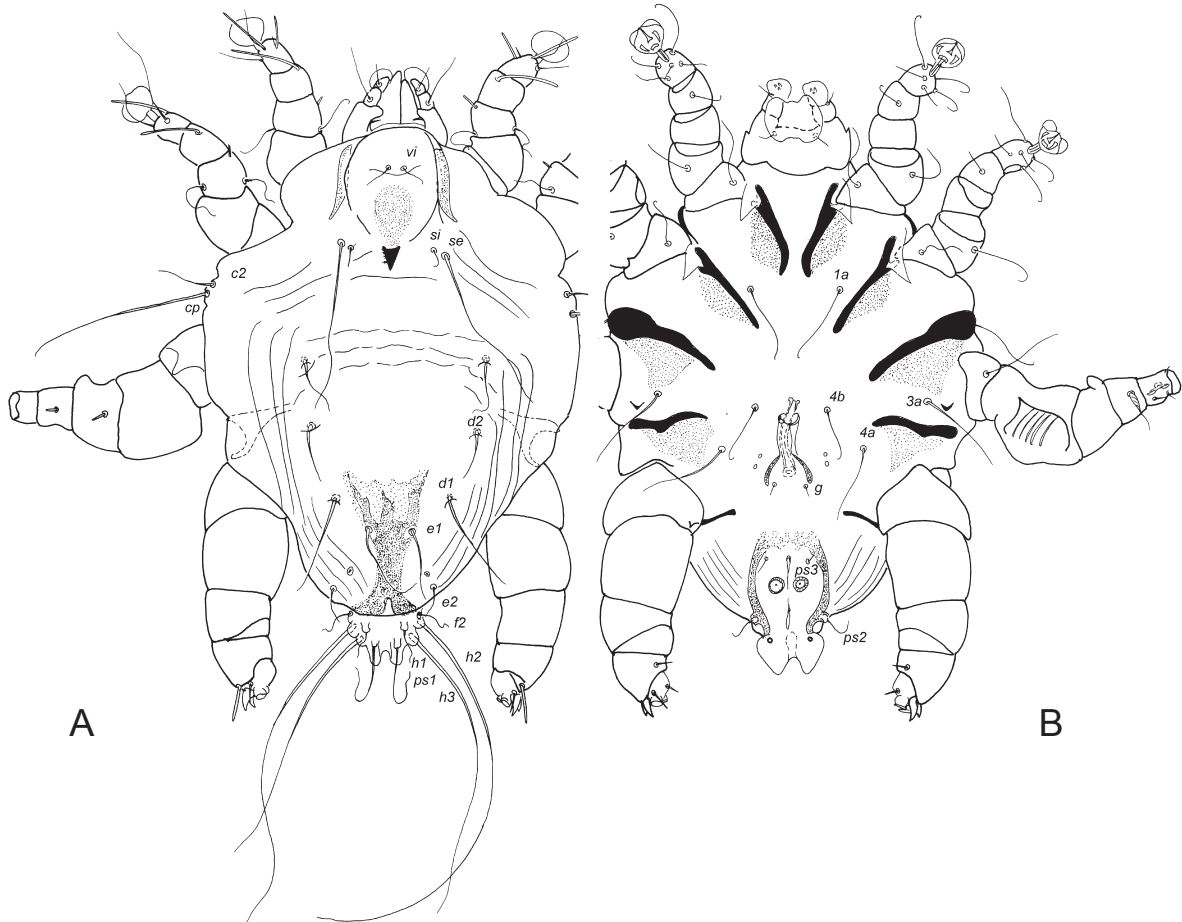


Fig. 55. *Myocoptes ondatrae* Lukoschus et Rouwet, 1968, male: A — dorsal view, B — ventral view.

Male. Opisthosomal lobes and paranal suckers present or absent. Legs III and IV normally developed. Legs IV of male without ambulacra. Setae *d*IV and *e*IV sucker-like.

Taxa included: 6 genera and 60 species, *Apocalypsis* Bochkov gen. nov. (1 species), *Crinicastor* Poppe, 1889 (4 species), *Gliricoptes* Lawrence, 1956 (9 species), *Myocoptes* Claparede, 1869 (22 species), *Sciurocoptes* Fain, Munting et Lukoschus, 1970 (2 species), *Trichoecius* Canestrini, 1899 (22 species).

Associations with hosts: inhabitants of skin and secondarily fur (*Trichoecius*) of rodents and carnivores, Carnivora (Mustelidae) and Rodentia (Anomaluridae, Calomyscidae, Cricetidae, Dipodidae, Gliridae, Muridae, Nesomyidae, Platanthomyidae, Sciuridae).

Myocoptes ictonyx Fain, 1970 is the only myocoptid species described from non rodent host, *Ictonyx striatus* (Mustelidae) from Africa. Ancestor of this species probably shifted on this host from its prey or even this host is accidental for this parasite because only two mite females were collected (Fain 1970a).

Distribution. Cosmopolite (not recorded from Madagascar).

Remarks. *Gliricoptes allactaga* Fain et Lukoschus, 1979 was described by Fain and Lukoschus (1979) from *Allactaga sibirica* (Dipodidae) from Mongolia. *G. allactaga* clearly differs by some characteristics from other myocoptids and I establish for this species a new genus whose diagnosis is provided below.

Genus *Apocalypsis* Bochkov gen. nov.

Type species: *Gliricoptes allactaga* Fain et Lukoschus, 1979

Both sexes: Body strongly flattened dorso-terminally. Posterior margin of propodonotal shield with pair of triangular median sclerites. Scales or tubercles absent. Setae *e*1 absent. Apodemes Ia not fused to each other. Coxal fields II and III situated close to each other. Condylaphores without apical hooks. Tarsus III with inflated (finger-like) seta. Genua I and II without solenidion σ .

Female: Hysteronotal shield absent. Epigynum present. Setae *f*2 absent. Anal opening situ-

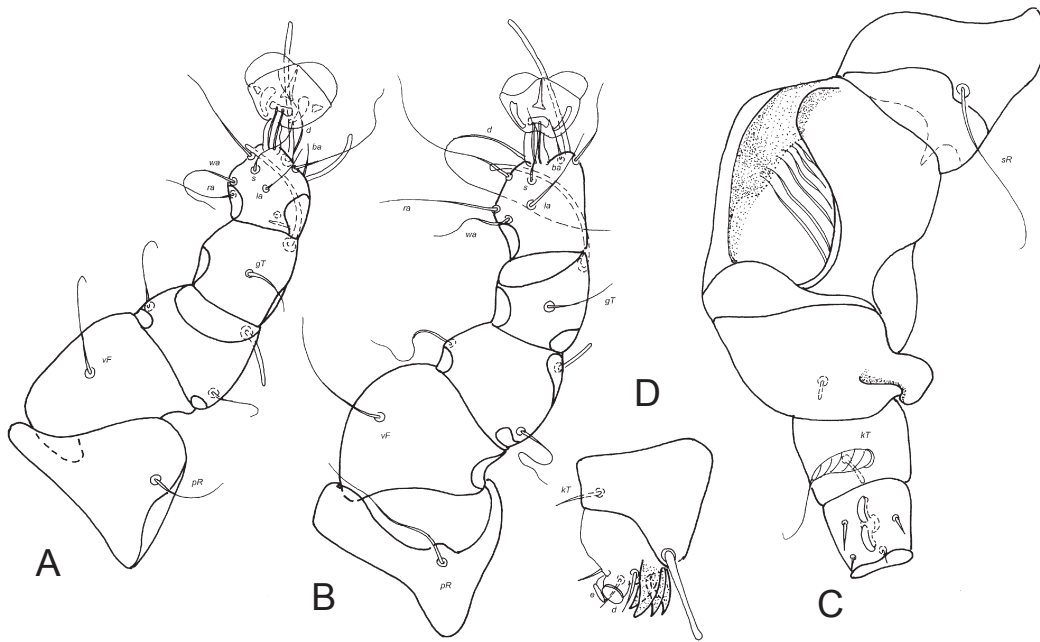


Fig. 56. *Myocoptes ondatrae* Lukoschus et Rouwet, 1968, legs of male: A — leg I in ventral view, B — leg II in ventral view, C — leg III in ventral view, D — tarsus IV in dorsal view.

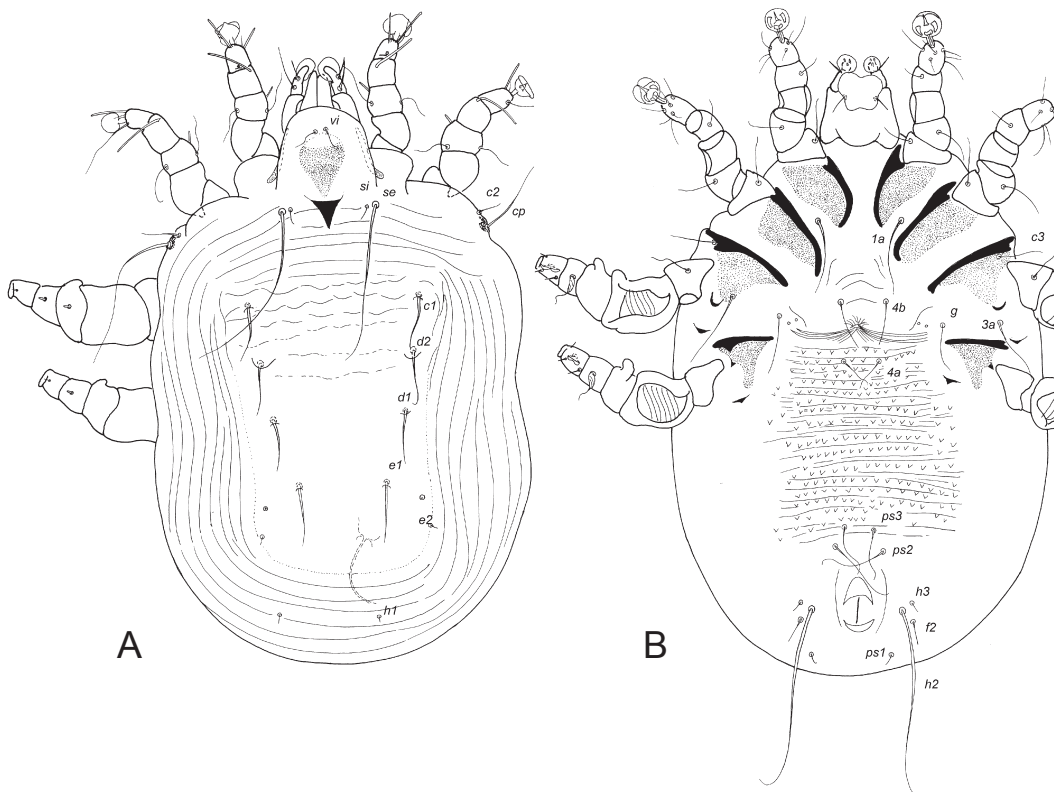


Fig. 57. *Myocoptes ondatrae* Lukoschus et Rouwet, 1968, female: A — dorsal view, B — ventral view.

ated ventrally. Bursa copulatrix opened ventroterminally.

Male: Hysteronotal shield present. Coxal field II without median cuticular fold. Paranal suckers present. Posterior end of opisthosoma widely rounded, lobar membrane strongly reduced and situated ventrally. Tarsi IV without projections (spines).

Other species included: type species only.

Differential diagnosis. The new genus is close to *Gliricoptes*, in both genera, setae *e1*, *f2* (in most species of *Gliricoptes*) and solenidia σ I and II absent, the posterior margin of the propodonotal shield has a pair of triangular paramedian sclerites. Differential characters of these genera are given in the key below.

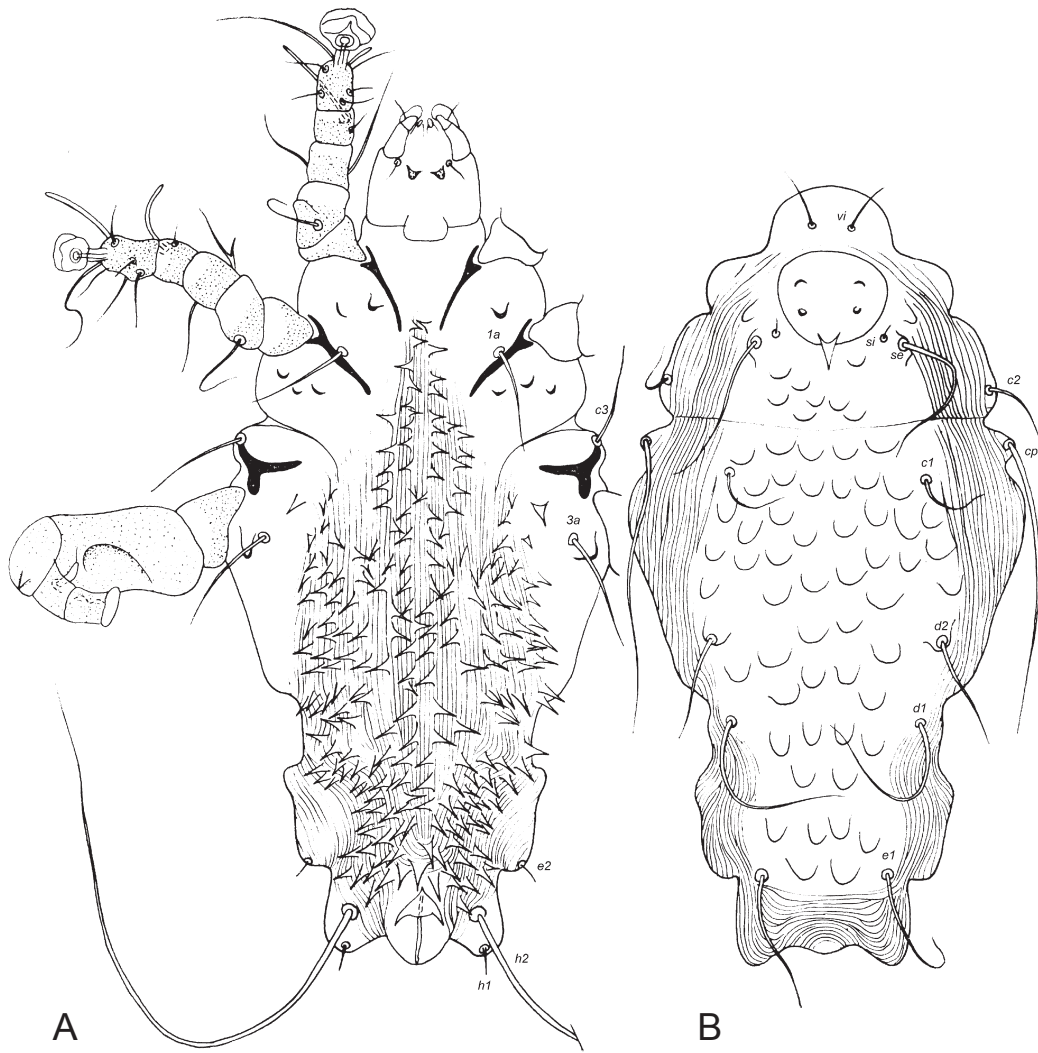


Fig. 58. *Myocoptes musculus* (Koch, 1844), larva: A — dorsal view, B — ventral view (after Fain et al. (1970), with minor modifications).

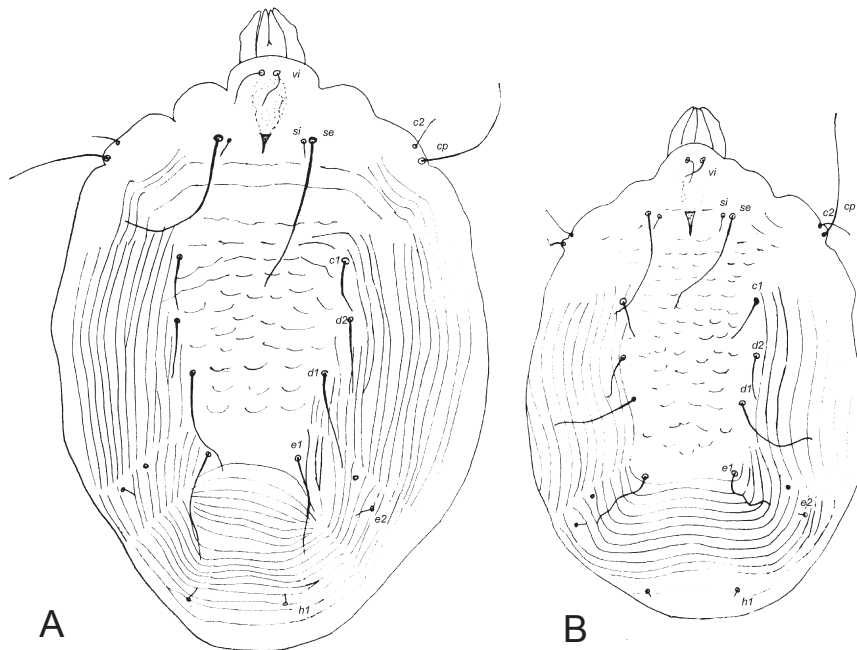


Fig. 59. *Myocoptes musculus* (Koch, 1844), nymphs in dorsal view: A — protonymph, B — tritonymph (after Fain et al. (1970), with minor modifications).

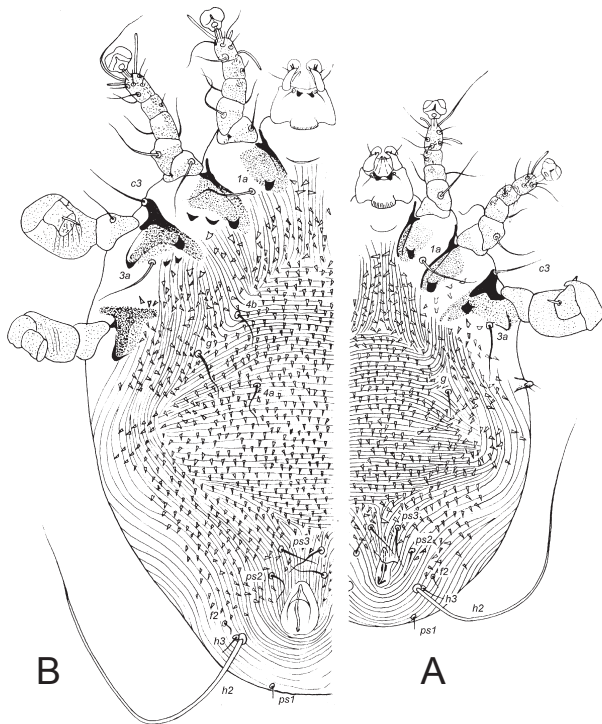


Fig. 60. *Myocoptes musculus* (Koch, 1844), nymphs in ventral view: A — protonymph, B — tritonymph (after Fain et al. (1970), with minor modifications).

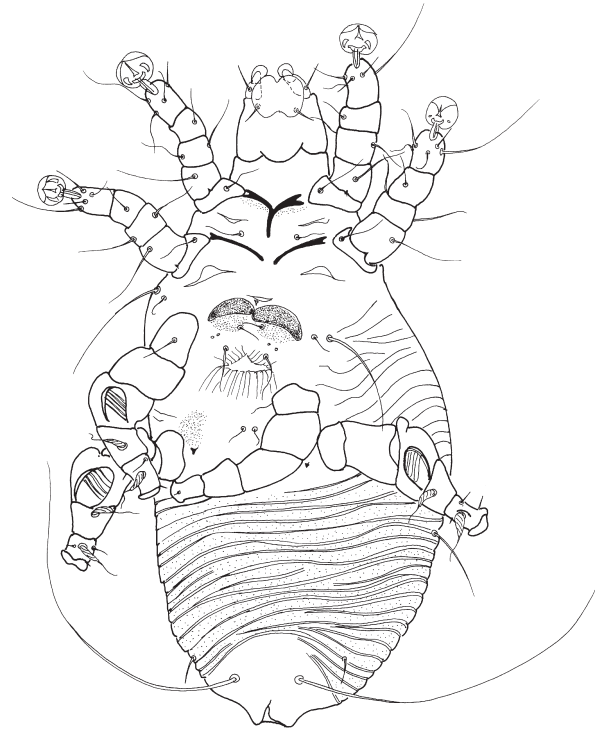


Fig. 61. *Trichoecius* sp., female in ventral view.

Etymology. I guess this is the last genus described in the family Myocoptidae because the problem of 2012. Therefore I named this genus *Apocalypsis*.

Main references. Fain (1970a) — revision of African Myocoptidae, Fain et al. (1970) — revision of European Myocoptidae, OConnor (1982) — diagnosis of Myocoptidae and references, Bockov and OConnor (2008) — comparative observations on myocoptid morphology.

Key to the family Myocoptidae Gunther, 1942

1. *Both sexes*: Body strongly flattened dorso-ventrally. Apodemes Ia not fused to each other. *Female*: Anal opening situated ventrally. Bursa copulatrix opened ventro-terminally or terminally. *Male*: Coxal field II without median cuticular fold. Paranal suckers present 2
 — *Both sexes*: Body slightly subcylindrical. Apodemes Ia fused to each other. *Female*: Anal opening situated dorsal or dorso-terminally. Bursa copulatrix opened dorsally. *Male*: Coxal field II with median cuticular fold. Paranal suckers absent *Trichoecius* Canestrini, 1899
2. *Both sexes*: Posterior margin of propodonotal shield with pair of triangular paramedian sclerites or without them. Genua I and II without solenidion. Setae *e1* absent 4

3. *Both sexes*: Tarsus III bearing basally inflated seta with whip-like extension. *Female*: Epigynum distinct. Hysteronotal shield present, without scales. *Male*: Opisthosomal lobes widely separated from each other. Tarsi IV without projections *Sciurocoptes* Fain et al., 1969
- *Both sexes*: Tarsus III bearing inflated bilobe seta without whip-like extension. *Female*: Epigynum indistinct. Hysteronotal shield absent but hysteronotum with non striated area covered by scale-like or tuberculous pattern (in *M. striatus*, this area striated). *Male*: Opisthosomal lobes closely situated to each other. Tarsi IV with 4 spine-like projections *Myocoptes* Claparede, 1869
4. *Both sexes*: Posterior margin of propodonotal shield with pair of triangular paramedian sclerites. Setal series *h+ps* complete (6 setae) — setae *h1* (or *ps1*) present. Coxal fields II and III situated close to each other. *Male*: Hysteronotal shield present 5
 — *Both sexes*: Posterior margin of propodonotal shield without median sclerites. Setal series *h+ps* not complete (5 setae) — setae *h1* (or *ps1*) absent. Coxal fields II and III situated far from each other.

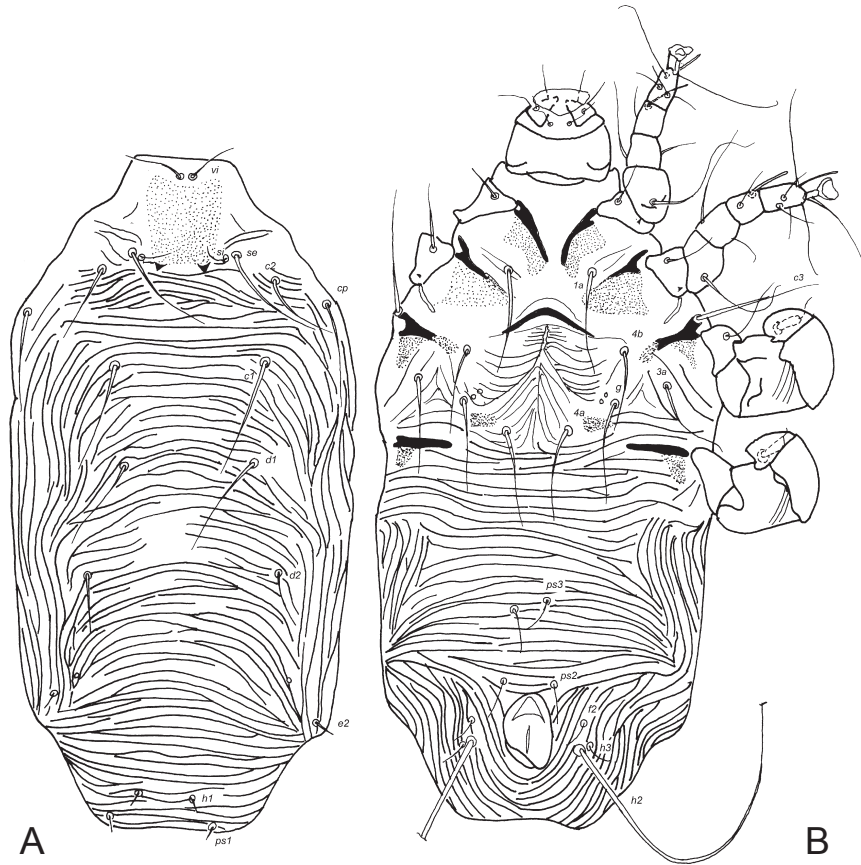


Fig. 62. *Apocalypsis allactaga* (Fain et Lukoschus, 1979), male: A — ventral view, B — dorsal view (after Fain and Lukoschus (1979), with minor modifications).

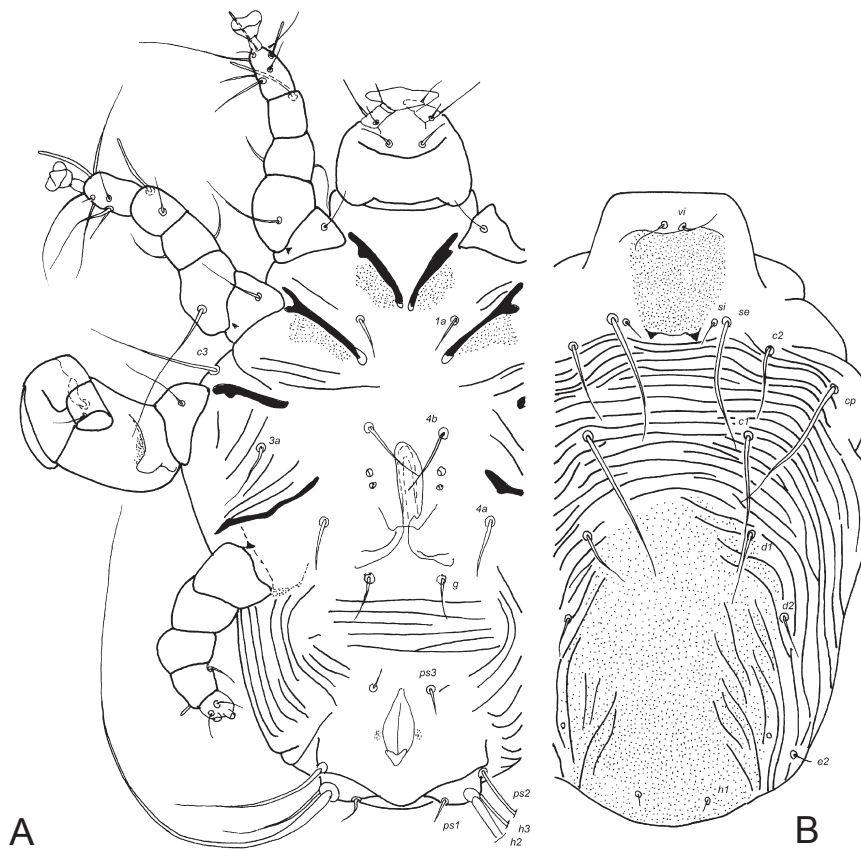


Fig. 63. *Apocalypsis allactaga* (Fain et Lukoschus, 1979), female: A — dorsal view, B — ventral view (after Fain and Lukoschus (1979), with minor modifications).

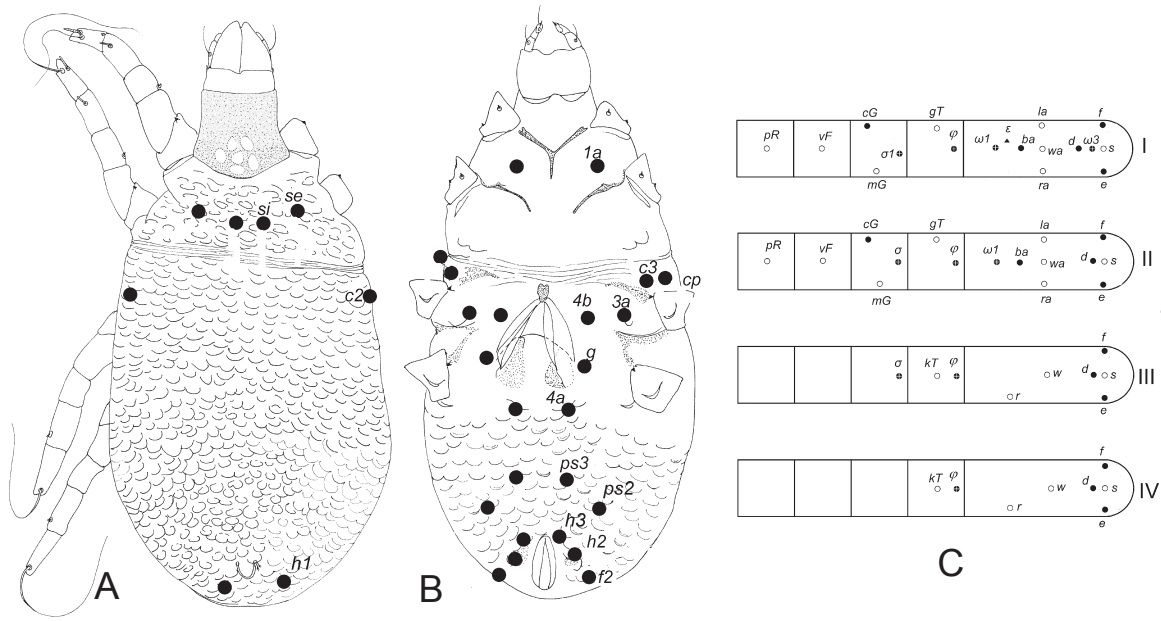


Fig. 64. Gastronyssidae, scheme of setation: A — idiosoma in dorsal view, B — same in ventral view, C — legs I-IV.

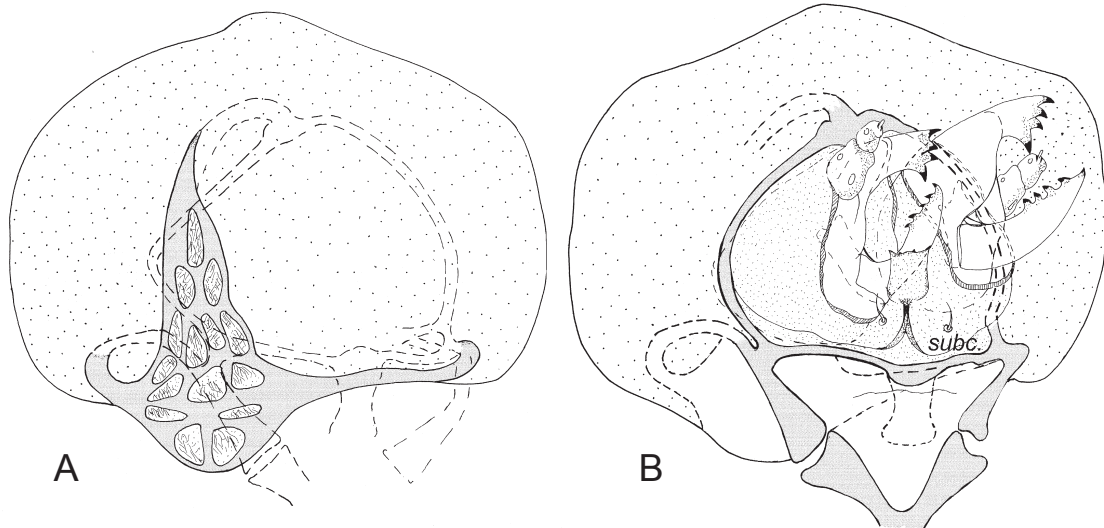


Fig. 65. *Gastronyssus bakeri* Fain, 1955, female propodonotal shield and gnathosoma: A — dorsal view, B — ventral view (after Bochkov et al. (2008b), with minor modifications).

Male: Hysteronotal shield absent
 *Crinicastor* Poppe, 1889
 5. *Both sexes*: Condylphores with apical hooks.
Female: Hysteronotal shield present. *Male*: Posterior end of opisthosoma bilobate. Paranal suckers present
Gliricoptes Lawrence, 1956 — *Both sexes*: Condylphores without apical hooks. *Female*: Hysteronotal shield absent. *Male*: Posterior end of opisthosoma widely rounded. Paranal suckers absent
 *Apocalypsis* Bochkov gen. n.

Family Gastronyssidae Fain, 1956

Type genus: *Gastronyssus* Fain, 1955.

Table 12, Figs. 64–77

Diagnosis. *Both sexes*. Cheliceral hood and ventral apophysis of movable digit absent. Palpal

eupathidia absent. Posterior margin of subcapitulum with pair of ventral projections (absent in *Gastronyssus*). Idiosoma slightly elongated. Supracoxal sclerite and supracoxal opening indistinct, setae *scx* absent. Propodonotal shield present, other dorsal shields absent. Genital papillae absent. Intercoxal attaching organs absent. Many idiosomal setae lost. Legs without clasping organs. In many genera, setae *sIV* present, solenidia $\sigma I, II$ and III absent, solenidion $\omega 3$ present only in males of some genera. Pretarsi normally developed, strongly reduced or absent. Condylphore guide lost. Ambulacral sclerites strongly reduced, almost indiscernible.

Female. Opisthogastral shields absent. Ovipore Y-shaped. Epigynum strongly reduced. Anal

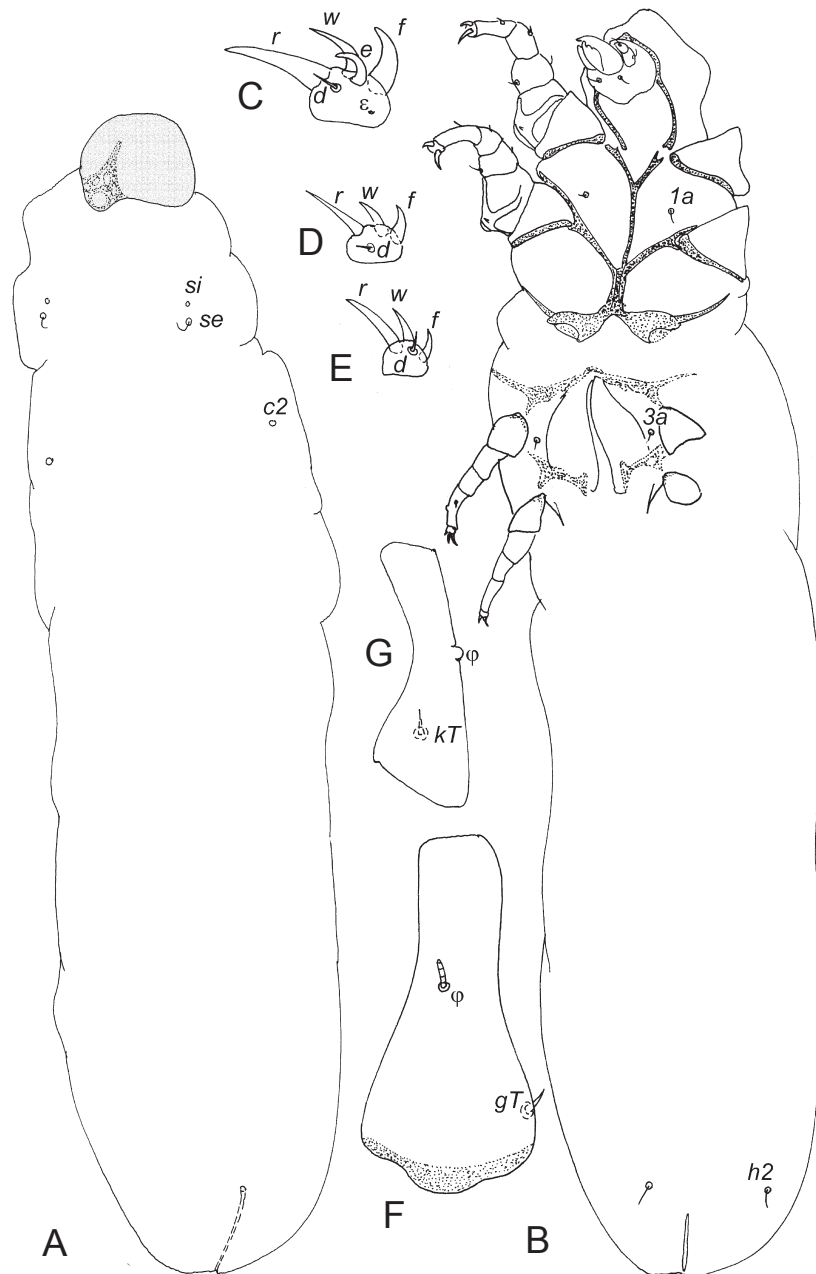


Fig. 66. *Gastronyssus bakeri* Fain, 1955, female: A — dorsal view, B — ventral view, C — tarsus I in dorsal view, D — tarsus III in dorsal view, E — tarsus IV in dorsal view, F — tibia I in dorsal view, G — tibia III in dorsal view (after Bochkov et al. (2008b), with minor modifications).

opening situated ventrally or terminally. Oviparous.

Male. Opisthosomal lobes and paranal suckers absent. Legs III and IV normally developed. Setae *d*IV filiform, *e*IV alveoli or microspines.

Taxa included. 9 genera and 42 species in 2 subfamilies, Gastronyssinae Fain, 1959, Gastronyssini Fain, 1959 — *Gastronyssus* Fain, 1955 (2 species), Phyllostomyssini Bochkov et OConnor, 2008 — *Phyllostomyssus* Fain, 1970 (1 species), Rodhainyssini Fain, 1964 — *Eidolonysus* Fain, 1967 (2 species), *Mycteronyssus* Fain, 1959 (2 species), *Opsonysus* Fain, 1959 (7 spe-

cies), *Pseudoopsonysus* Bochkov et OConnor, 2008 (3 species), *Rodhainyssus* Fain, 1956 (12 species), Yunkeracarinae Fain, 1964 — *Sciuracarus* Fain, 1964 (1 species), *Yunkeracarus* Fain, 1957 (12 species).

Associations with hosts: Gastronyssinae — inhabitants of the nasal passages, eye orbits, stomach and duodenum of Chiroptera (Emballonuridae, Hipposideridae, Megadermatidae, Molossidae, Nycteridae, Phyllostomidae, Pteropodidae, Rhinolophidae, Vespertilionidae); Yunkeracarinae — inhabitants of the nasal passages of Rodentia (Cricetidae, Muridae, Sciuridae).

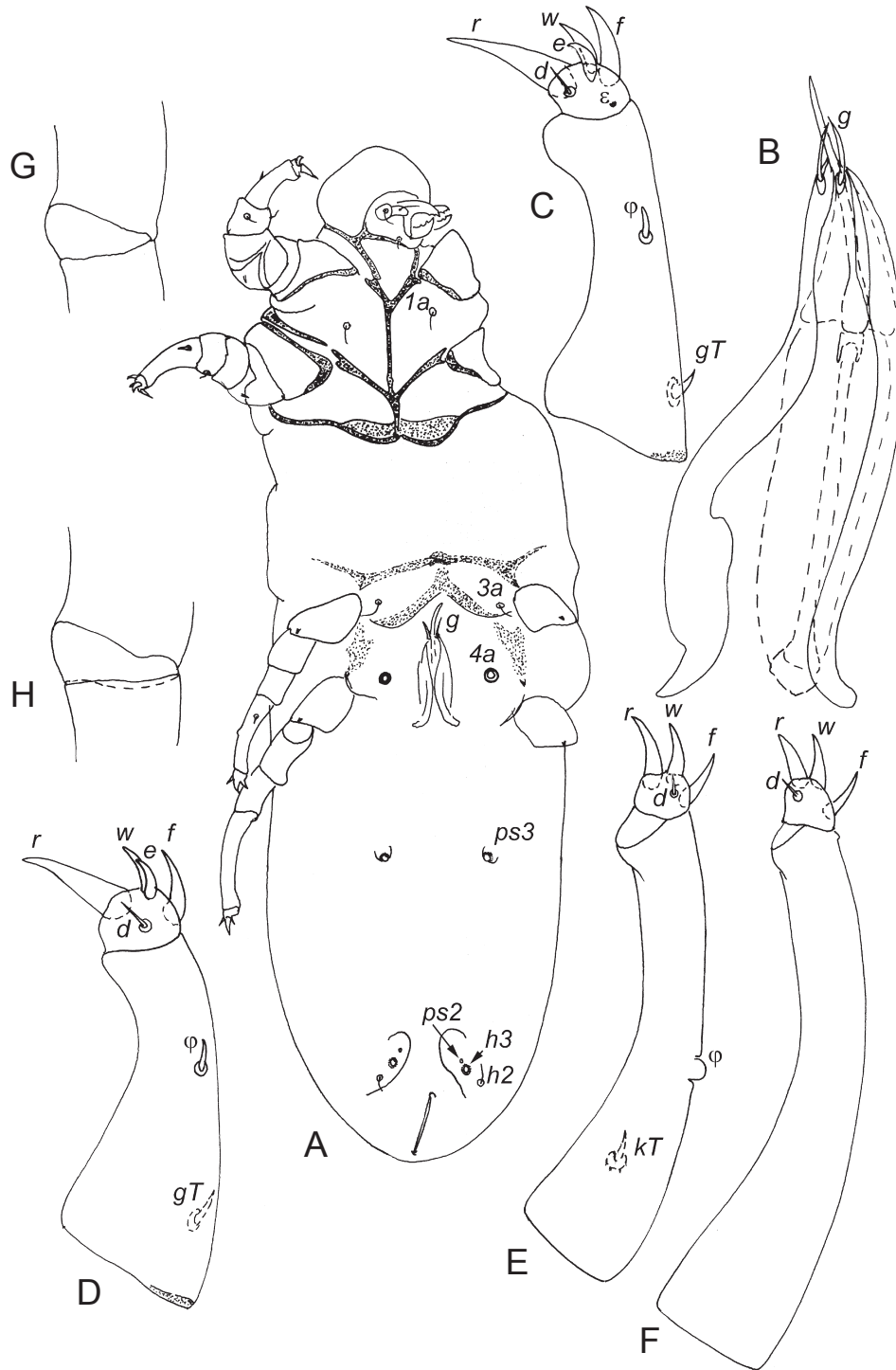


Fig. 67. *Gastronyssus bakeri* Fain, 1955, male: A — ventral view, B — aedeagus, C — tibia and tarsus I in dorsal view, D — tibia and tarsus II in dorsal view, E — tibia and tarsus III in dorsal view, F — tibia and tarsus IV in dorsal view, G — posterior part of genu III in ventral view, H — posterior part of genu IV in ventral view (after Bochkov et al. (2008b), with minor modifications).

Distribution. Gastronyssinae — cosmopolite; Yunkeracarinae — Eurasia, Africa (including Madagascar), North and South America.

Main references. Fain (1956) — diagnosis of Gastronyssidae, Fain (1964a) — review of Gastronyssidae, Fain (1967) — revision of the genus *Rodhainyssus*, Bochkov et al. (2008b) — revision of Gastronyssidae.

Key to genera of the family Gastronyssidae Fain, 1959 (both sexes)

1. Setae *elc.p* cone-like. Idiosoma covered with scale-like striations. Setae *cp*, *baI–II*, and *pRI–II* present (*pR* absent in *Y. limnomys*). Setae *eIII–IV* situated antiaxially, represented by microsetae. Tarsi of legs I–II subequal in length to respective tibiae. Pretarsi of all legs distinctly developed

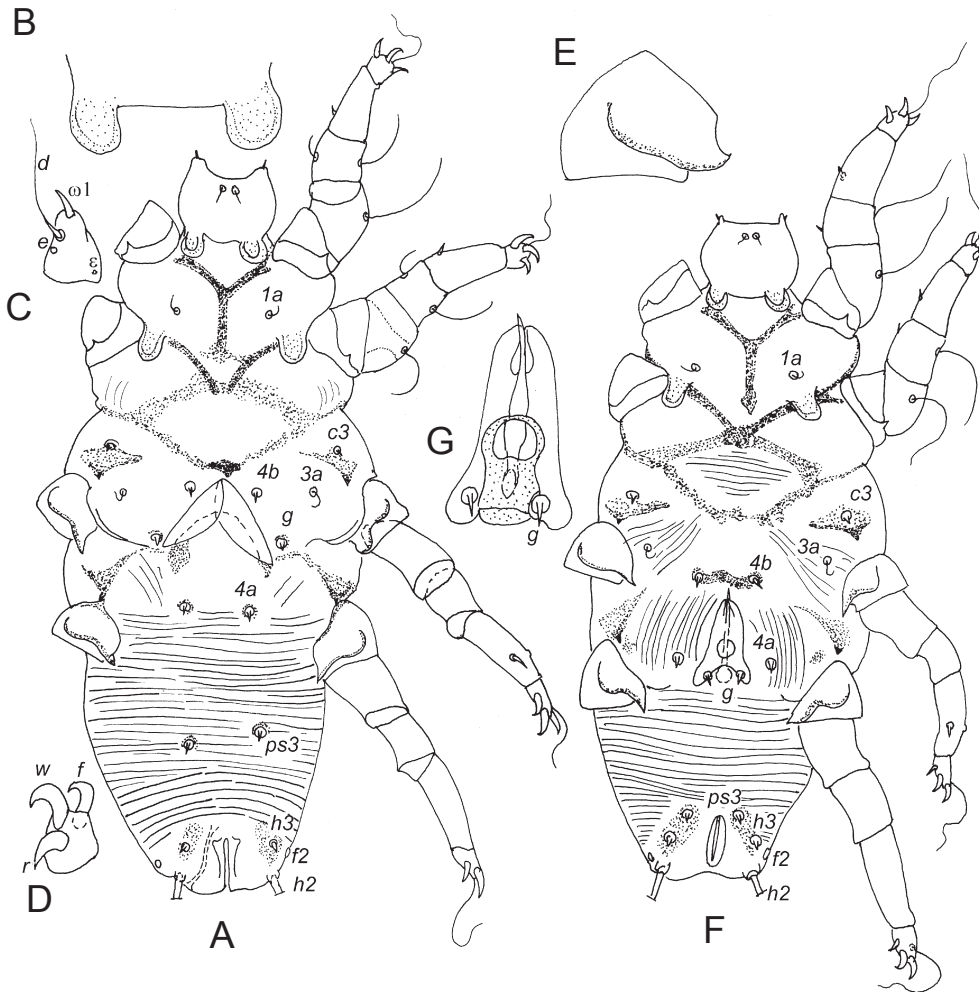


Fig. 68. *Rodhainyssus myotis* Fain, 1967, female: A — ventral view, B — postero-ventral projections of gnathosoma, C — tarsus I in dorsal view, D — same in ventral view, E — trochanter III in ventral view; male: F — ventral view, G — aedeagus (after Bochkov et al. (2008b), with minor modifications).

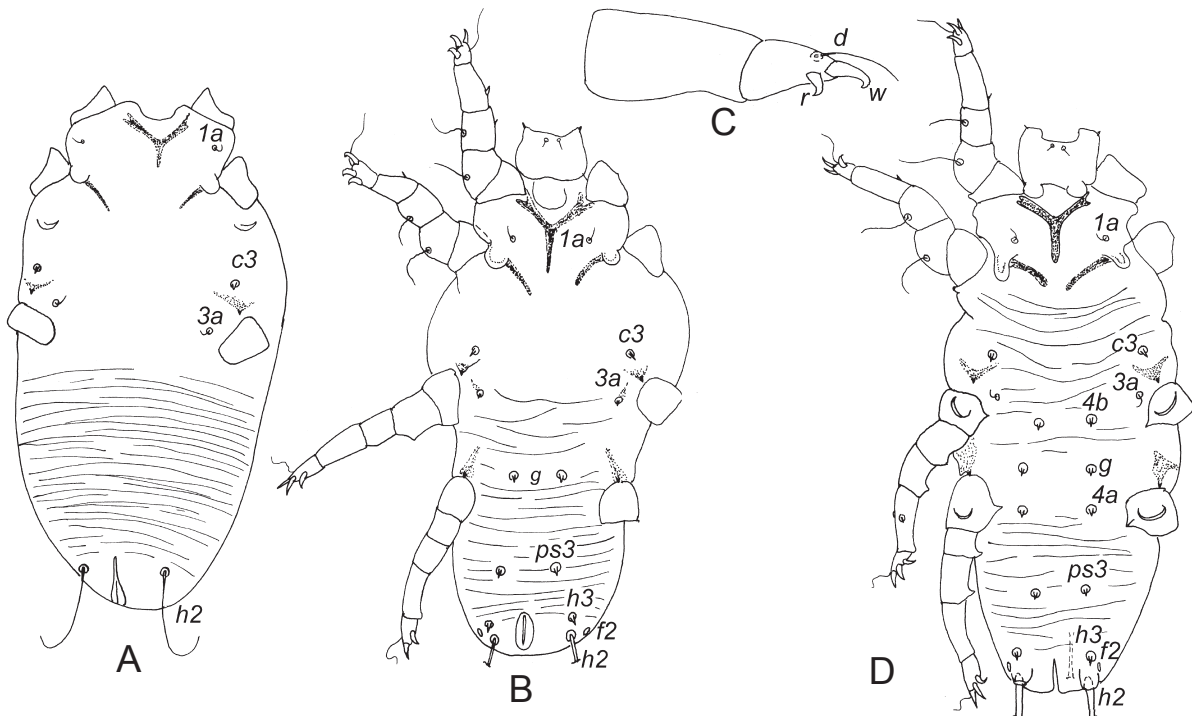


Fig. 69. *Rodhainyssus myotis* Fain, 1967, larva: A — ventral view; protonymph: B — ventral view, C — tibia and tarsus III in ventral view; tritonymph: D — ventral view (after Bochkov et al. (2008b), with minor modifications).

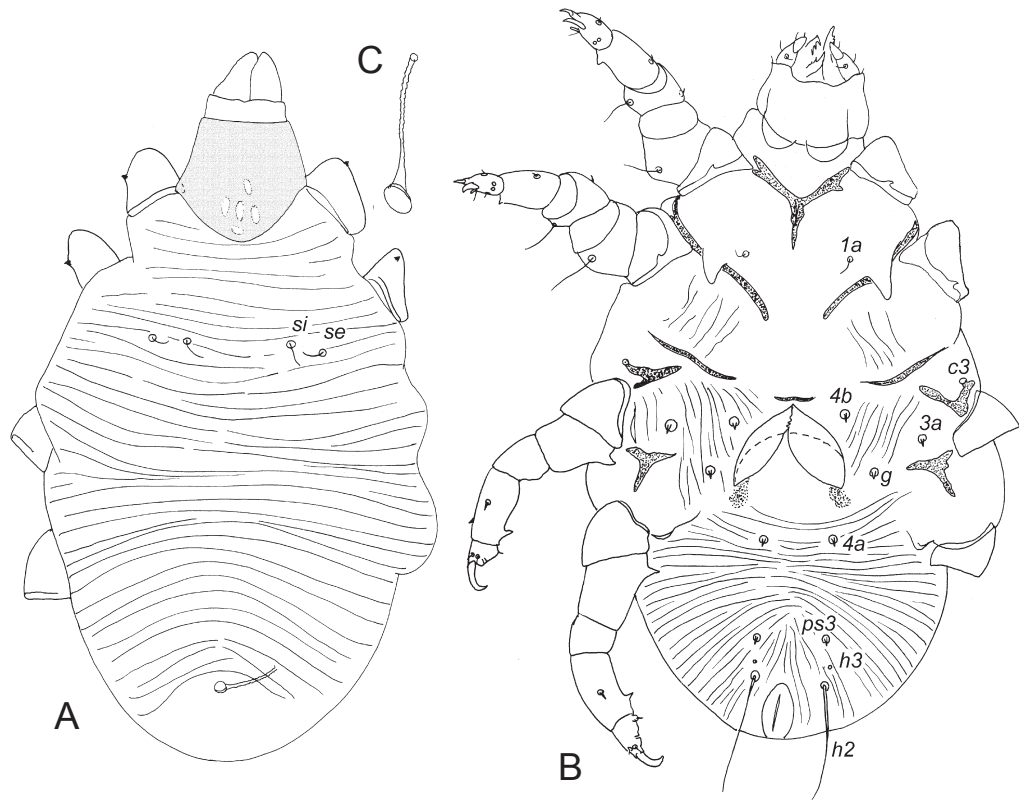


Fig. 70. *Phyllostomyssus conradyunkeris* Fain, 1970, female. A — dorsal view, B — ventral view, C — bursa copulatrix (after Bochkov et al. (2008b), with minor modifications).

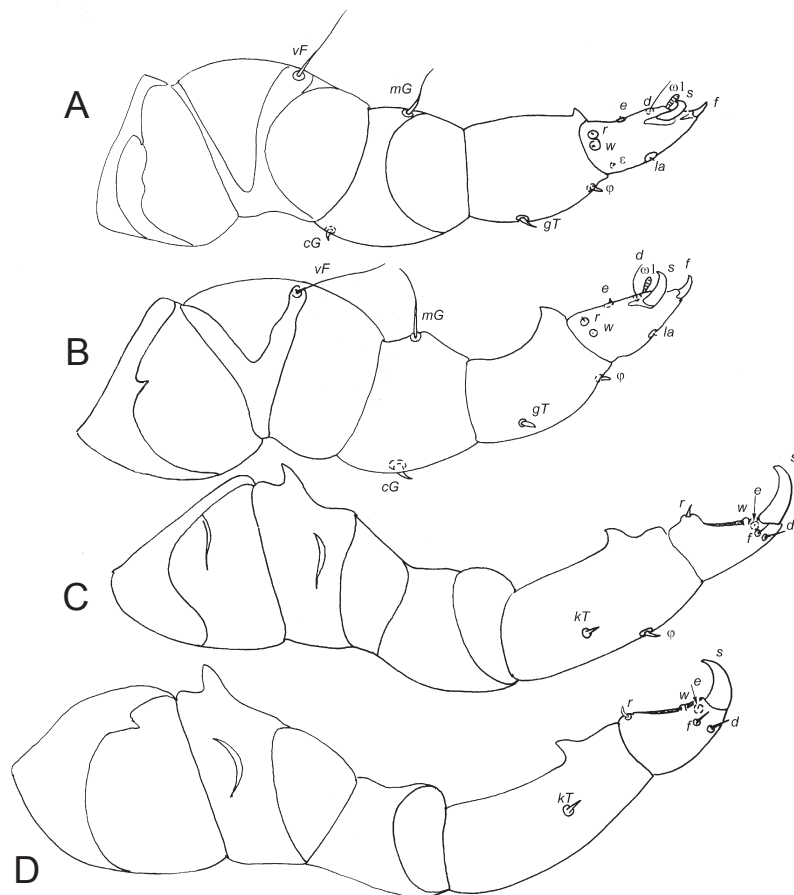


Fig. 71. *Phyllostomyssus conradyunkeris* Fain, 1970, female legs in ventral view: A — leg I, B — leg II, C — leg III, D — leg IV (after Bochkov et al. (2008b), with minor modifications).

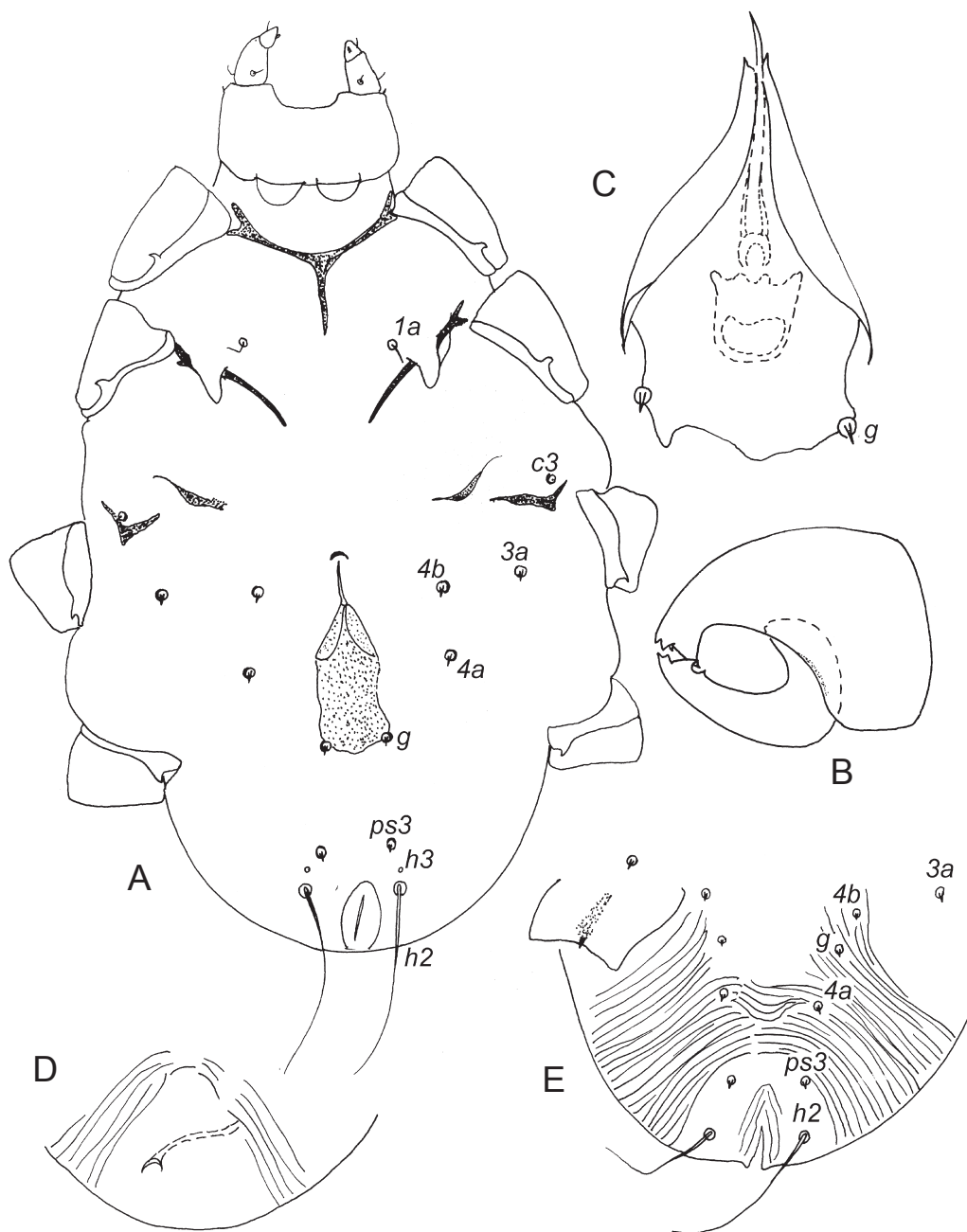


Fig. 72. *Phyllostomyssus conradyunkeri* Fain, 1970, male: A — ventral view, B — chelicera in lateral view, C — aedeagus; female tritonymph: D — posterior end of opisthosoma in dorsal view, E — opisthosoma in ventral view (after Bochkov et al. (2008b), with minor modifications).

consisting of ambulacral stalk and disc
 Yunkeracarinae Fain, 1964 8
 — Setae *elc.p* stick-like or absent. Idiosoma covered with transverse striations or without striations. Setae *cp*, *baI–II* and *pRI–II* absent. Setae *eIII–IV* absent, or situated dorsally and represented by alveoli. Tarsi of legs I–II distinctly shorter than respective tibiae. Pretarsi of legs I–II absent; pretarsi of legs III–IV absent or strongly reduced with only ambulacral stalk present
 Gastronyssinae Fain, 1959 2
 2. Gnathosoma situated terminally, not fused with propodonotal shield. Pair of ventral projections

present on subcapitular margin. Idiosoma 2–2.5 times longer than wide 3
 — Gnathosoma displaced ventrally, dorsal surface of subcapitulum fused with propodonotal shield. Ventral projections of subcapitular margin absent. Idiosoma vermiform, 5–6 times longer than wide ..
Gastronyssini Fain, 1959
 (*Gastronyssus* Fain, 1955)
 3. Setae *si* represented by alveoli. Ventral sclerotization in median part of idiosoma between coxal fields I and II present. Postgenital shield in males absent. Setae *eI–II* in dorsal position, represented by alveoli. Setae *laI–II* absent

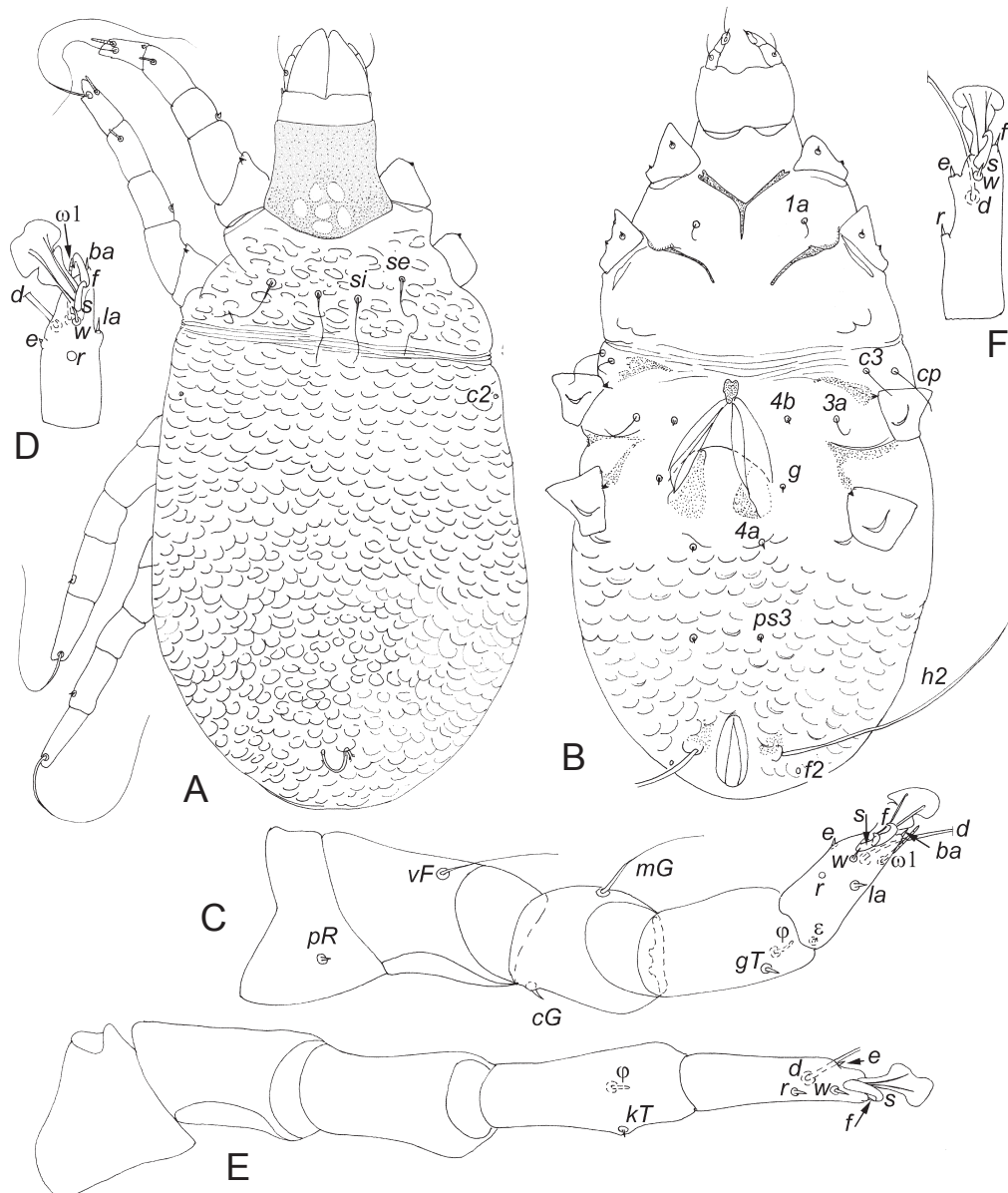


Fig. 73. *Yunkeracarus lophuromys* Bochkov et OConnor, 2008, female: A — dorsal view, B — ventral view, C — leg I in ventral view, D — tarsus II in ventral view, E — leg III in ventral view, F — tarsus IV in ventral view (after Bochkov et al. (2008b), with minor modifications).

Rodhainyssini Fain, 1964 4
 — Setae *si* filiform. Ventral sclerotization in median part of idiosoma between coxal fields I and II absent. Postgenital shield in males absent. Setae *eI–II* in antiaxial position, represented by microsetae. Setae *laI–II* present *Phyllostomyssus* Fain, 1970)
 4. Tarsi III–IV distinctly shorter than respective tibiae. Setae *sIII–IV* absent 5
 — Tarsi III–IV subequal to respective tibiae. Setae *sIII–IV* present *Eidolonyssus* Fain, 1967
 5. Anus situated ventrally. Pretarsi III–IV absent 6
 — Anus situated terminally. Rudiments of ambu-

lacrual stalk III–IV present
 *Mycteronyssus* Fain, 1970
 6. Setae *subc.* absent. Idiosoma elongated, ellipsoid in outline, with butterfly-like sclerotization between coxal fields II. Propodeonotal shield fused with apodemes Ia. Setae *dI–IV* subequal or shorter than respective tarsi 7
 — Setae *subc.* present. Idiosoma elongated, slightly rhomboid in outline, with straight or/and oblique sclerotized bands between coxal fields II. Propodeonotal shield separated from apodemes Ia. Setae *dI–IV* distinctly longer than respective tarsi *Rodhainyssus* Fain, 1956
 7. Posterior margin of propodeonotal shield concave. Coxal fields III closed. Setae *f2* present. Se-

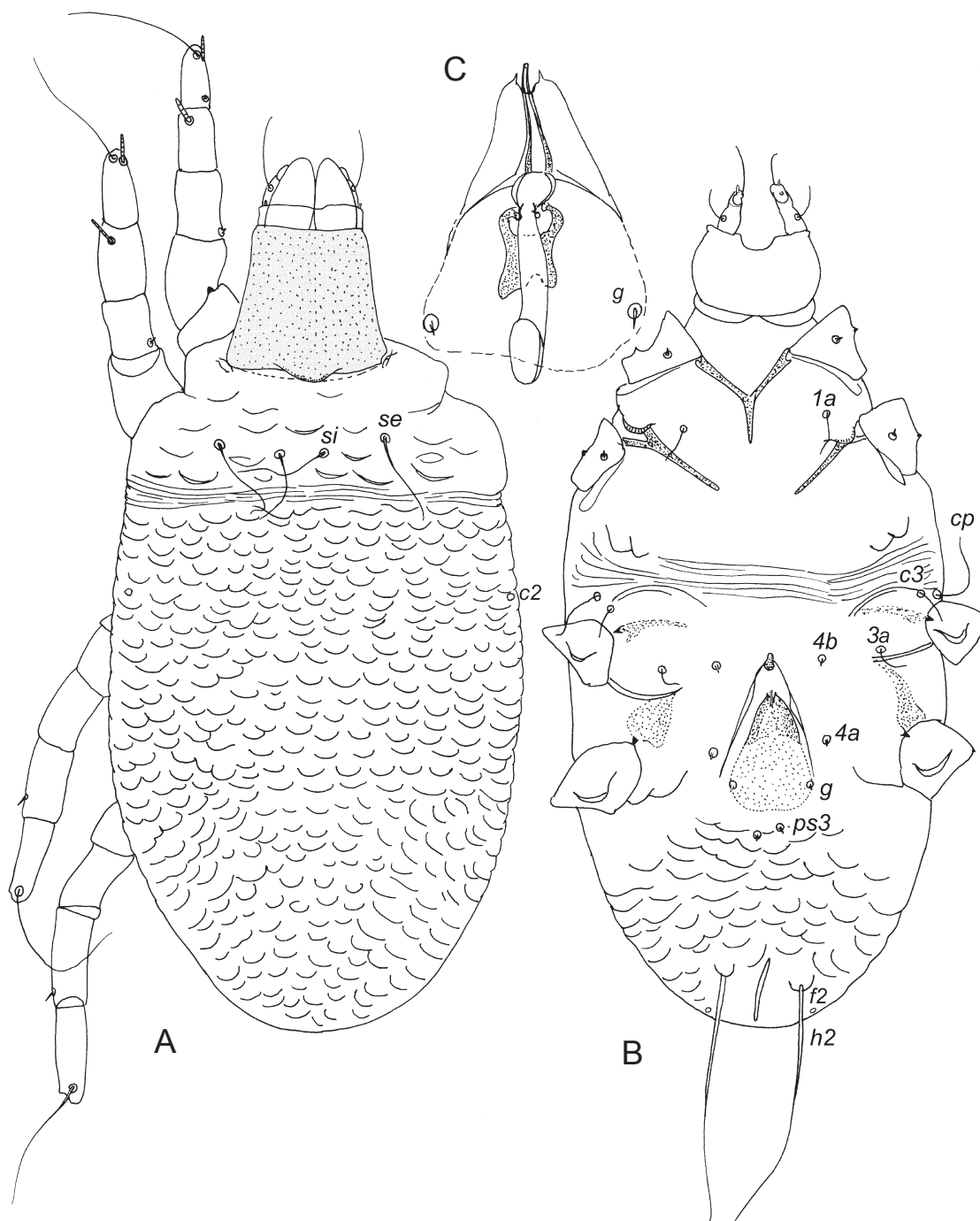


Fig. 74. *Yunkeracarus lophuromys* Bochkov et OConnor, 2008, male: A — dorsal view, B — ventral view, C — aedeagus (after Bochkov et al. (2008b), with minor modifications).

tae *ps3* and *h3* spine-like. *Male*: Setae *ps2* absent ... *Pseudoopsonyssus* Bochkov et OConnor, 2008 — Posterior margin of propodonotal shield widely rounded or bearing median projection. Coxal fields III open. Setae *f2* absent. Setae *ps3* represented by alveoli or absent; setae *h3* represented by alveoli or microspines. *Male*: Setae *ps2* present *Opsonyssus* Fain, 1959 8. Ventral posterior projection of subcapitular margin widely rounded. Setae *c2* alveoli. Solenidion ϕ IV present *Yunkeracarus* Fain, 1957

— Ventral posterior projection of subcapitular margin triangular. Setae *c2* microsetae. Solenidion ϕ IV absent *Sciuracarus* Fain, 1957

Family Lemurnyssidae Fain, 1957

Type genus: *Lemurnyssus* Fain, 1957.

Table 13, Figs. 78, 79

Diagnosis. *Both sexes.* Cheliceral hood and ventral apophysis of movable digit absent. Chelicerae modified; fixed digit strongly reduced; movable digit elongated, needle-like, bearing small

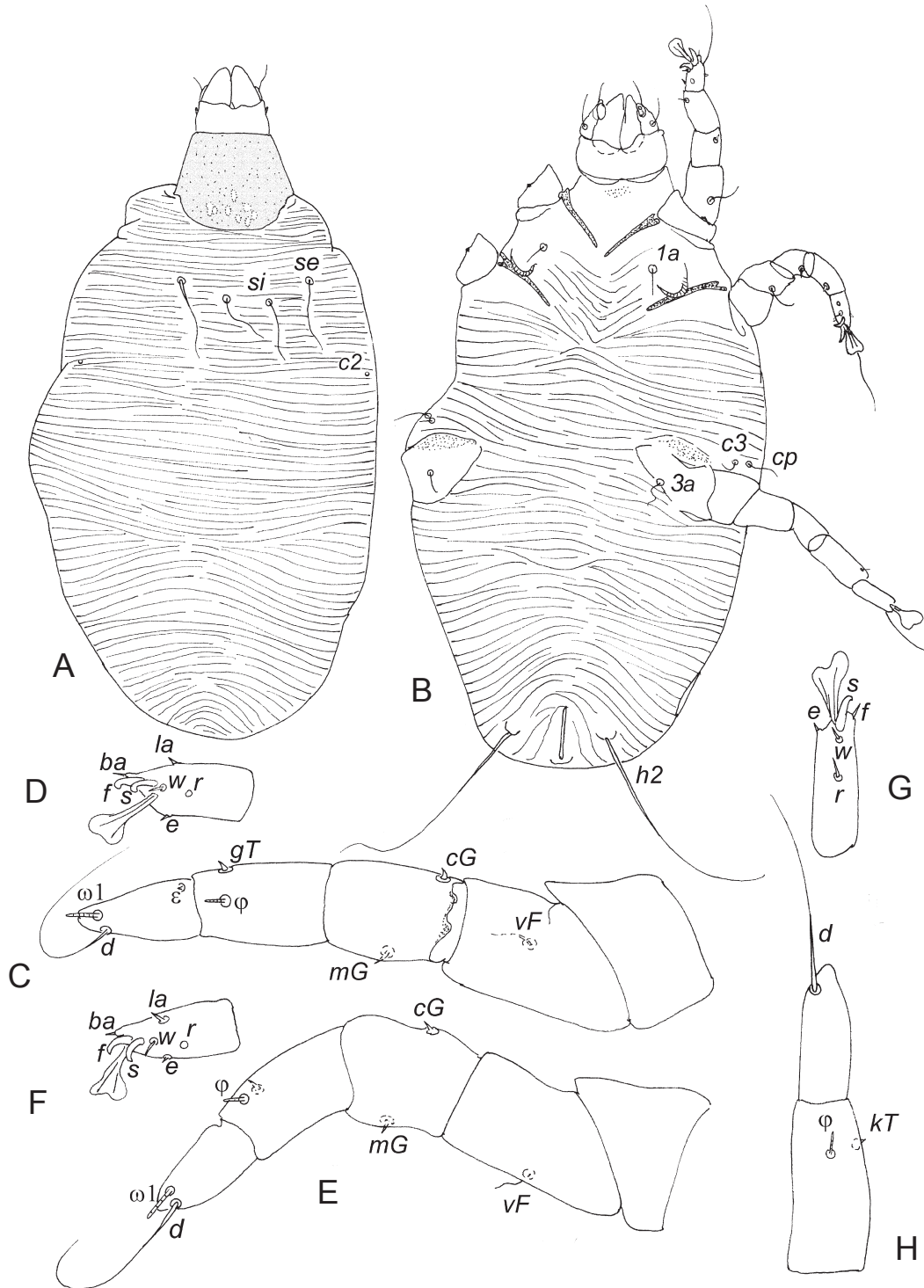


Fig. 75. *Yunkeracarus lophuromys* Bochkov et OConnor, 2008, larva: A — dorsal view, B — ventral view, C — leg I in dorsal view, D — tarsus I in ventral view, E — leg III in dorsal view, F — tarsus IV in ventral view, H — tarsus and tibia III in dorsal view, G — tarsus III in ventral view (after Bochkov et al. (2008b), with minor modifications).

teeth; other cheliceral structures absent. Palpal segments fused to each other. Setae *dT* of palpal tarsi and eupathidia absent. Subcapitulum without setae *subc*. Idiosoma distinctly elongated. Supracoxal sclerite and supracoxal opening indistinct, setae *scx* absent. Propodonal shield divided onto prescapular and postscapular shields; paired meta-

podosomal, and hysteronotal shields present. Genital papillae absent. Anal opening situated terminally. Intercoxal attaching organs absent. Many idiosomal setae, including *4b*, lost. Legs without clasping organs. Setae *sIV* present, solenidia σ I, II and III absent. Pretarsi normally developed or strongly reduced. Condylphore guide lost. Am-

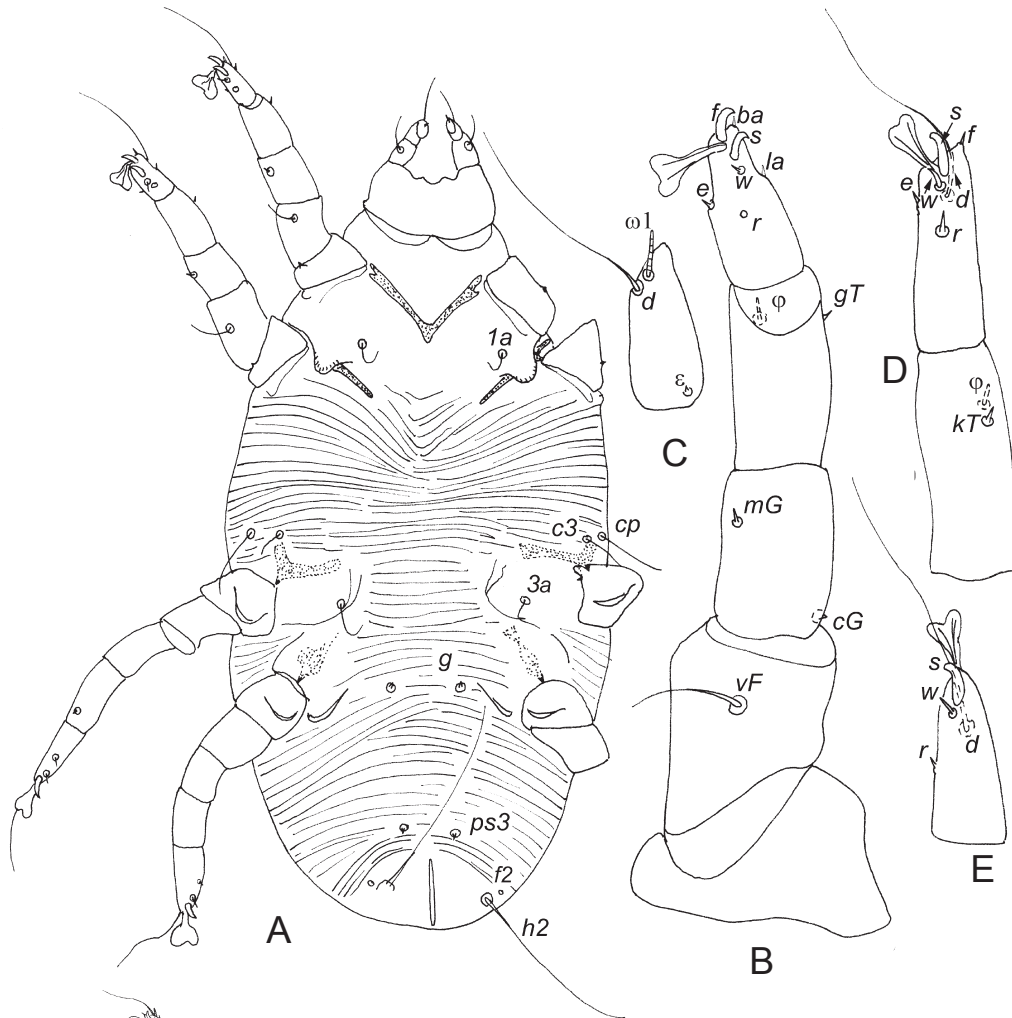


Fig. 76. *Yunkeracarus lophuromys* Bochkov et OConnor, 2008, protonymph: A — ventral view, B — leg I in ventral view, C — tarsus I in dorsal view, D — tarsus and tibia III in ventral view, E — tarsus IV in ventral view (after Bochkov et al. (2008b), with minor modifications).

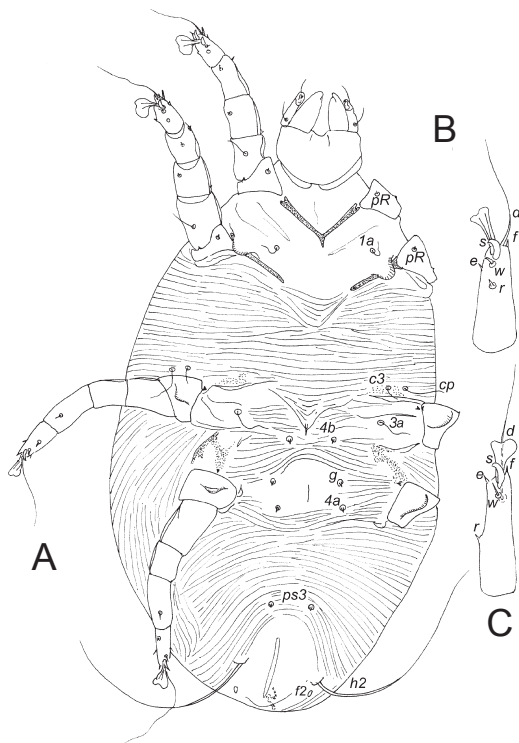


Fig. 77. *Yunkeracarus lophuromys* Bochkov et OConnor, 2008, tritonymph: A — ventral view, B — tarsus III in ventral view; C — tarsus IV in ventral view (after Bochkov et al. (2008b), with minor modifications).

bulacral sclerites strongly reduced, almost indiscernible.

Female. Opisthogastral shield present. Ovipore Y-shaped. Epigynum strongly reduced. Ovoviviparous.

Male. Opisthosomal lobes and paranal suckers absent. Legs III and IV normally developed. Setae *d*IV and *e*IV filiform.

Taxa included: 2 genera and 4 species — *Lemurnyssus* Fain, 1957 (1 species), *Mortelmansia* Fain, 1959 (3 species).

Associations with hosts: inhabitants of the nasal passages of Primates (Callitrichidae, Cebidae, Galagidae).

Distribution. Africa (not recorded from Madagascar), South America.

Main references. Fain (1957) — diagnosis of Lemurnyssidae, Fain (1964b) — review Le-

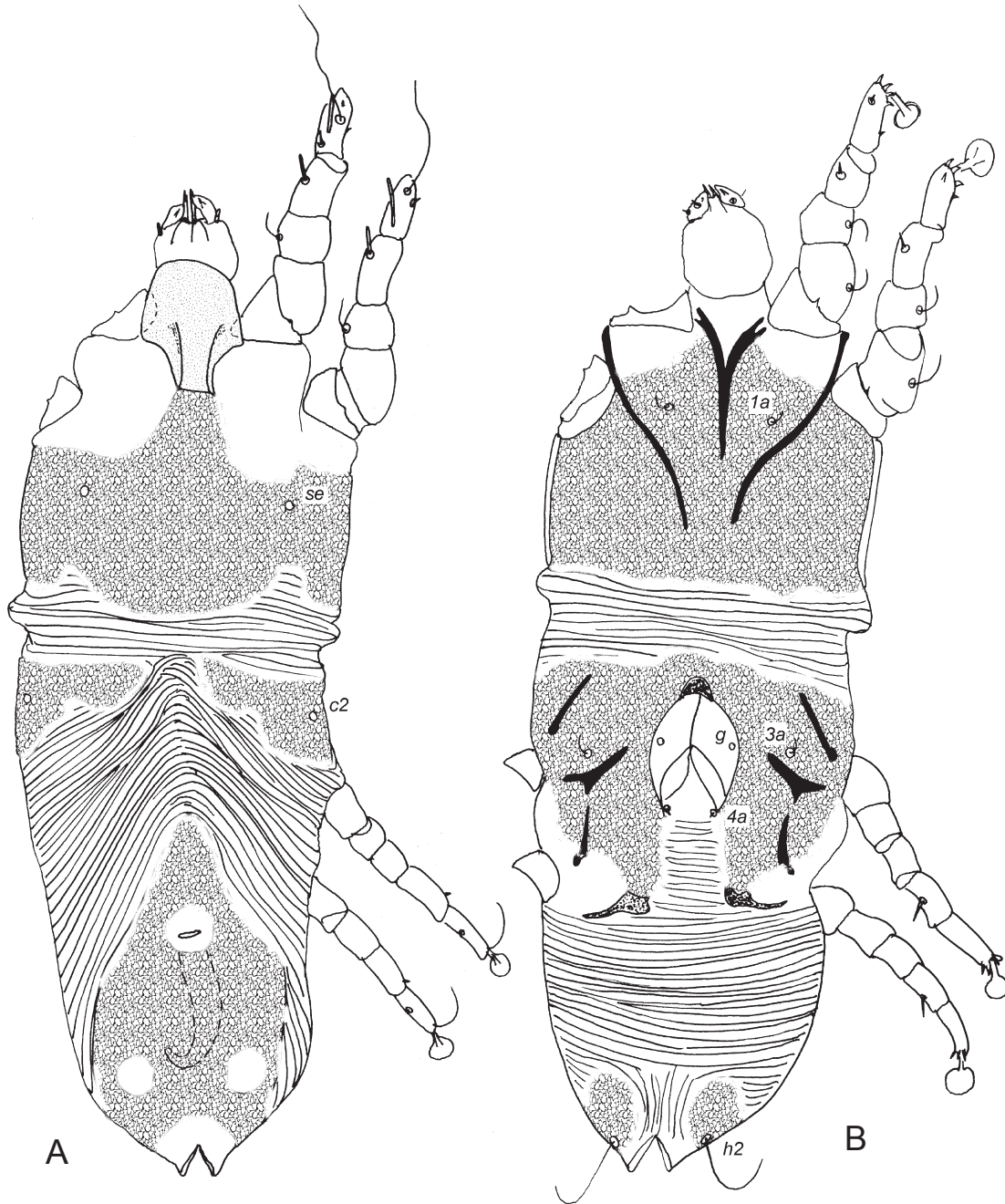


Fig. 78. *Lemurnyssus galagoensis* Fain, 1957, female: A — dorsal view, B — ventral view (after Bochkov et al. (2008b), with minor modifications).

murnyssidae, Bochkov et al. (2008b) — revision of Lemurnyssidae.

Key to genera of the family Lemurnyssidae Fain, 1957 (both sexes)

1. Idiosomal setae *c3* and *cp* absent. In females opisthogastric shields paired, strongly reduced. Leg setae *rI–II* spur-like, *rIII–IV* and *kTIV* present, *cGII* filiform. In males pretarsi normally developed; setae *ps3* present; aedeagus distinctly thickened *Lemurnyssus* Fain, 1957 — Idiosomal setae *c3* and *cp* present. Leg setae *rI–II* alveoli, *rIII–IV* and *kTIV* absent, *cGII* spur-

like. In females opisthogastric shield entire, distinctly developed. In males pretarsi III–IV strongly reduced; setae *ps3* absent; aedeagus in shape of narrow tube *Mortelmansia* Fain, 1959

Family Pneumocoptidae Fain, 1957

Type genus: *Pneumocoptes* Baker, 1951

Table 14, Figs. 80–82

Diagnosis. *Both sexes.* Cheliceral hood and ventral apophysis of movable digit absent. Setae *dT* of palpal tarsi and eupathidia absent. Subcapitulum without setae *subc*. Idiosoma ovoid in outlines. Supracoxal sclerite and supracoxal open-

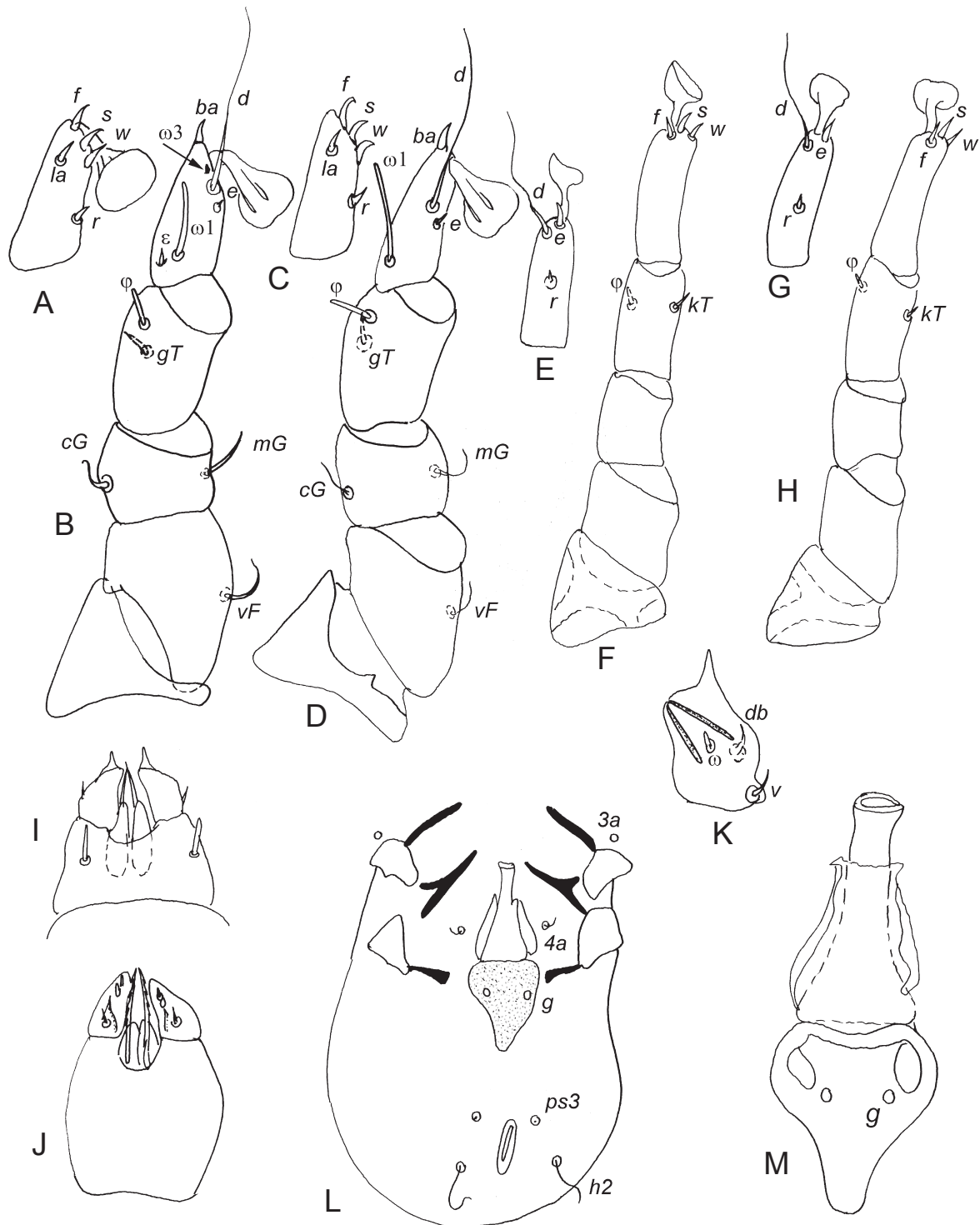


Fig. 79. *Lemurnyssus galagoensis* Fain, 1957, female (A–H): A — tarsus I in ventral view, B — leg I dorsal view, C — tarsus II in ventral view, D — leg II in dorsal view, E — tarsus III in dorsal view, F — leg III in ventral view, G — tarsus IV in dorsal view, H — leg IV in ventral view; male (I–M): I — gnathosoma in dorsal view, J — same in ventral view, K — palp in ventral view, M — aedeagus (after Bochkov et al. (2008b), with minor modifications).

ing indistinct, setae *scx* absent. Propodonal and hysteronotal shields present, distinctly developed. Propodonal shield with short longitudinal apodeme between bases of setae *si*. Small postanal sclerite present terminally. Opisthosoma weakly developed and legs IV inserted very close to posterior

body margin. Genital papillae absent. Intercostal attaching organs absent. Many idiosomal setae lost. Anal opening situated ventrally, close to posterior margin of opisthosoma. Legs without clasping organs. Solenidia $\omega 3$, $\sigma 1$, II and III, and famulus ϵ absent. Trochanters and femora III–IV joining at

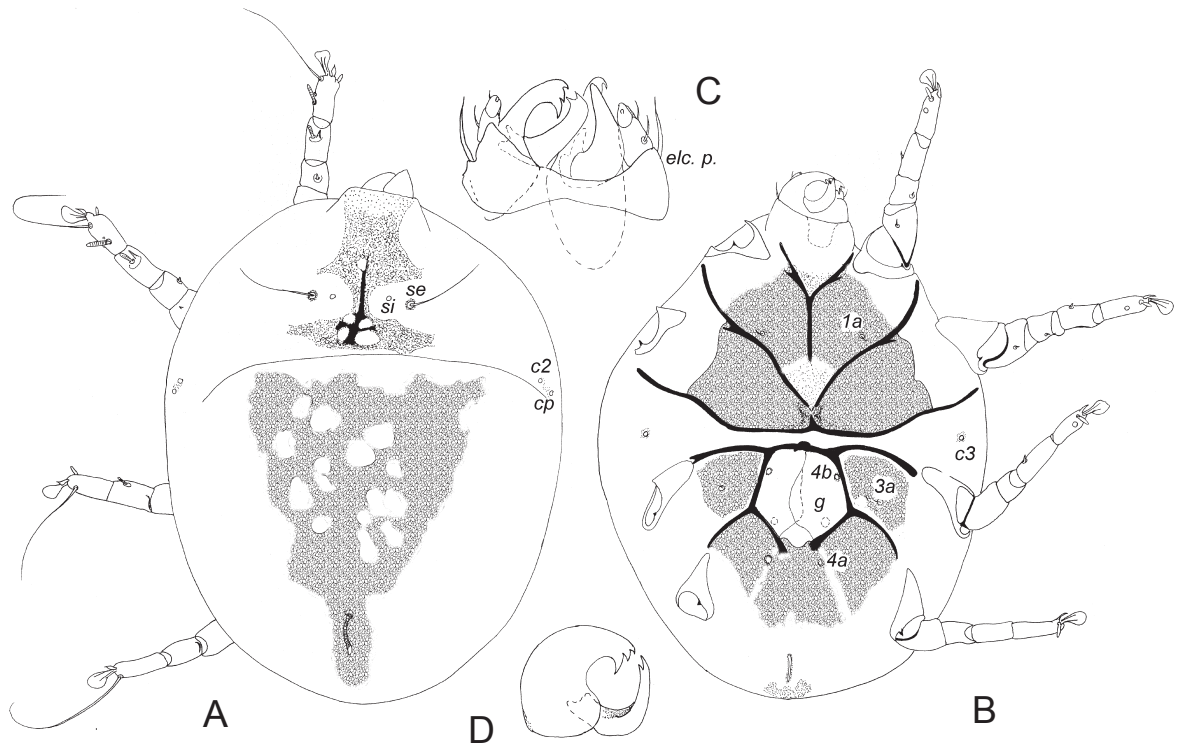


Fig. 80. *Pneumocoptes penrosei* Baker, 1951, female: A — dorsal view, B — ventral view, C — gnathosoma in ventral view, D — chelicera in lateral view (after Bochkov et al. (2008b), with minor modifications).

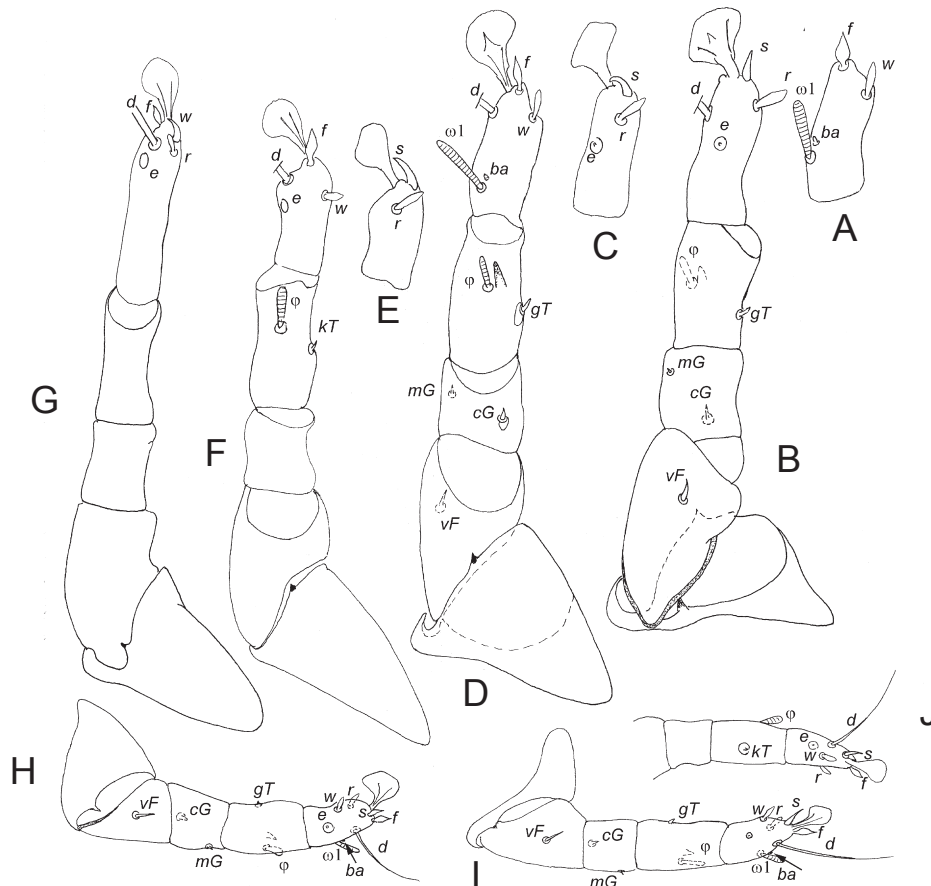


Fig. 81. *Pneumocoptes penrosei* Baker, 1951, female legs (A–G): A — tarsus I in dorsal view, B — leg I in ventral view, C — tarsus II in ventral view, D — leg II in dorsal view, E — tarsus III in ventral view, F — leg III in dorsal view, G — leg IV in dorsal view; larval legs in ventral view (I–H): I — leg I, J — leg II, H — leg III (after Bochkov et al. (2008b), with minor modifications).

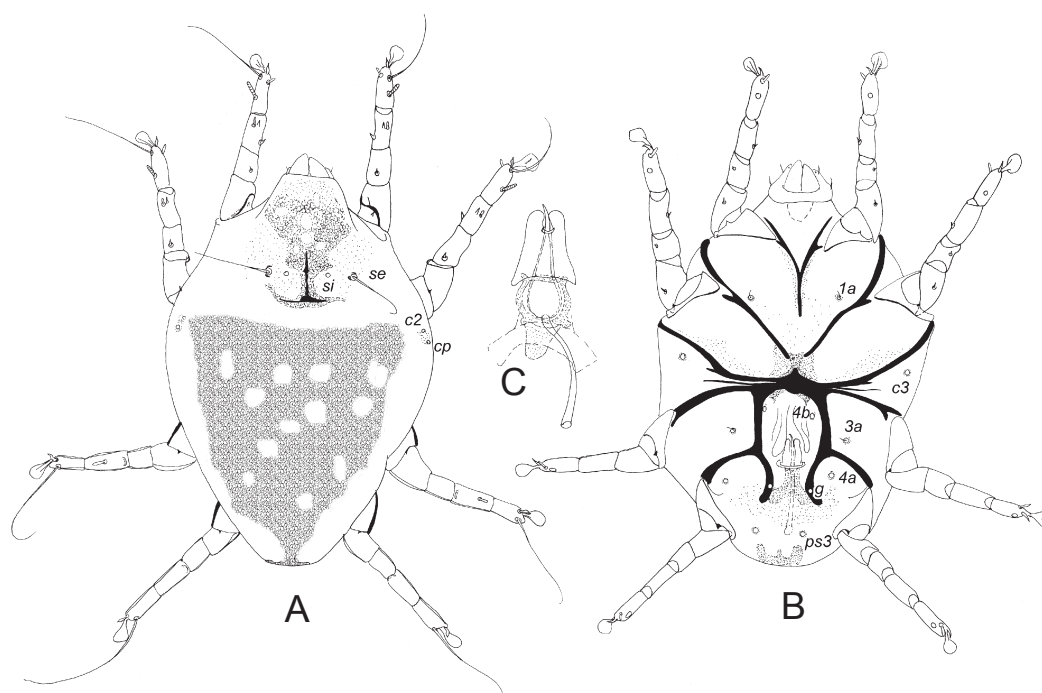


Fig. 82. *Pneumocoptes penrosei* Baker, 1951, male: A — dorsal view, B — ventral view, C — aedeagus (after Bochkov et al. (2008b), with minor modifications).

right angles. Tibiae I–II each with small dorsal spur “guarding” solenidion ϕ . Pretarsi normally developed. Condyliphore guide lost. Ambulacral sclerites strongly reduced, almost indiscernible.

Female. Opisthogastral shield present. Ovipore Y-shaped. Epigynum reduced, completely fused with distal ends of apodemes II. Ovoviviparous.

Male. Opisthosomal lobes and paranal suckers absent. Legs III and IV normally developed. Setae *dIV* and *eIV* not modified into suckers.

Taxa included. Only type genus and 4 species.

Associations with hosts: inhabitants of the lungs of Rodentia (Cricetidae, Sciuridae).

Distribution. Europe and North America.

Main references. Baker (1951) — description of *Pneumocoptes*, Kouchakij and Loos-Frank (1984) — observations on external morphology of *Pneumocoptes* spp., Bochkov et al. (2008b) — revision of Pneumocoptidae.

**Subfamilies incertae sedis
within sarcoptid complex**

Subfamily Listropsoralginae Fain, 1965

Type genus: *Listropsoralges* Fain, 1965.

Table 15, Figs. 83, 84

Diagnosis. *Both sexes.* Cheliceral hood and ventral apophysis of movable digit present. Idiosoma dorso-ventrally flattened. Supracoxal sclerite and supracoxal opening indistinct, setae *scx*

absent. Propodonotal and hysteronotal shields present, distinctly developed. Genital papillae present. Intercoxal attaching organs absent. Anal opening situated ventrally, close to posterior margin of opisthosoma. Legs I–II with retrorse projections. Pretarsi normally developed on legs I and II, weakly developed or absent on legs III and IV. Ambulacral sclerites discernible. Setae *ba*, *la*, and *waI*, II, and famulus ϵ absent.

Female. Opisthogastral shield present. Ovipore Y-shaped. Epigynum distinctly developed. Oviparous.

Male. Opisthosomal lobes present but their membranes weakly developed, paranal suckers present. Genital papillae absent or present. Legs III without ambulacra. Legs IV weakly developed, much shorter than legs III. Setae *dIV* and *eIV* filiform or *dIV* sucker-like (*Petauralges*).

Taxa included. 3 genera and 7 species, *Listropsoralges* Fain, 1965 (4 species), *Listropsoralgoides* Fain et Lukoschus, 1970 (1 species), *Petauralges* Fain et Lukoschus, 1979 (2 species).

Associations with hosts: inhabitants of the skin of therian mammals, Marsupialia: Dasyuromorphia (Dasyuridae), Didelphimorphia (Didelphidae), Diprotodontia (Petauridae); Placentalia: Rodentia (Echimyidae).

The common ancestor of this subfamily probably parasitized marsupials. The parasitism of listropsoralgins on rodents of the family Echimyidae

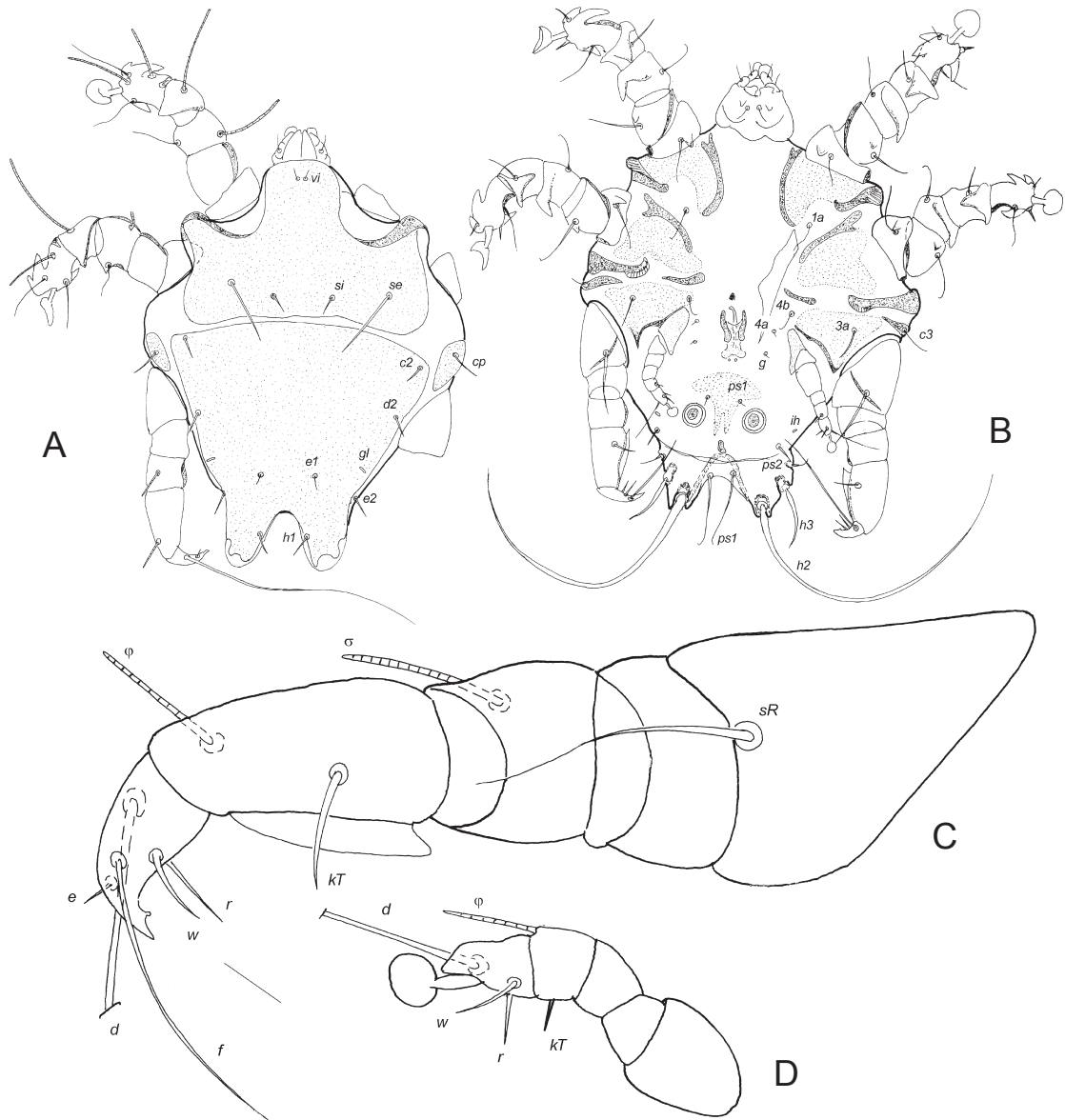


Fig. 83. *Listropsoralges caluromys* Bochkov et Wauthy, 2009, male: A — dorsal view, B — ventral view, C — leg III in ventral view, D — leg IV in ventral view (after Bochkov and Wauthy (2009), with minor modifications).

is probably a consequence of their switching from the New World marsupials (Bochkov and Wauthy 2009).

Distribution. Australia and South America.

Main references. Fain (1965b) — review of Listropsoralginae, Fain and Lukoschus (1970) — description of *Listropsoralgoides*, Domrow (1988) — morphology of *Petauralges*, Bochkov and Wauthy (2009) — revision of *Listropsoralges*.

Key to genera of the subfamily Listropsoralginae Fain, 1965

1. *Both sexes:* Setae *dI* absent. *Female:* Setae *g* located posterior to level of coxal fields IV. *Male:* Adanal shield present. Genital papillae absent. Setae *dIV* filiform 2

— *Both sexes:* Setae *dI* present. *Female:* Setae *g* located at level of coxal fields IV. *Male:* Adanal shield absent. Genital papillae present. Setae *dIV* sucker-like
 *Petauralges* Fain et Lukoschus, 1979
 2. *Both sexes:* Strongly sclerotized spur-like projections of coxal fields II absent. Setae *h2* whip-like, longer than legs IV; setae *sIII* absent. *Male:* Setae *psI* present
 *Listropsoralges* Fain, 1965
 — *Both sexes:* Strongly sclerotized spur-like projections of coxal fields II present. Setae *h2* short, much shorter than legs IV; setae *sIII* present. *Male:* Setae *psI* absent
 *Listropsoralgoides* Fain et Lukoschus, 1970

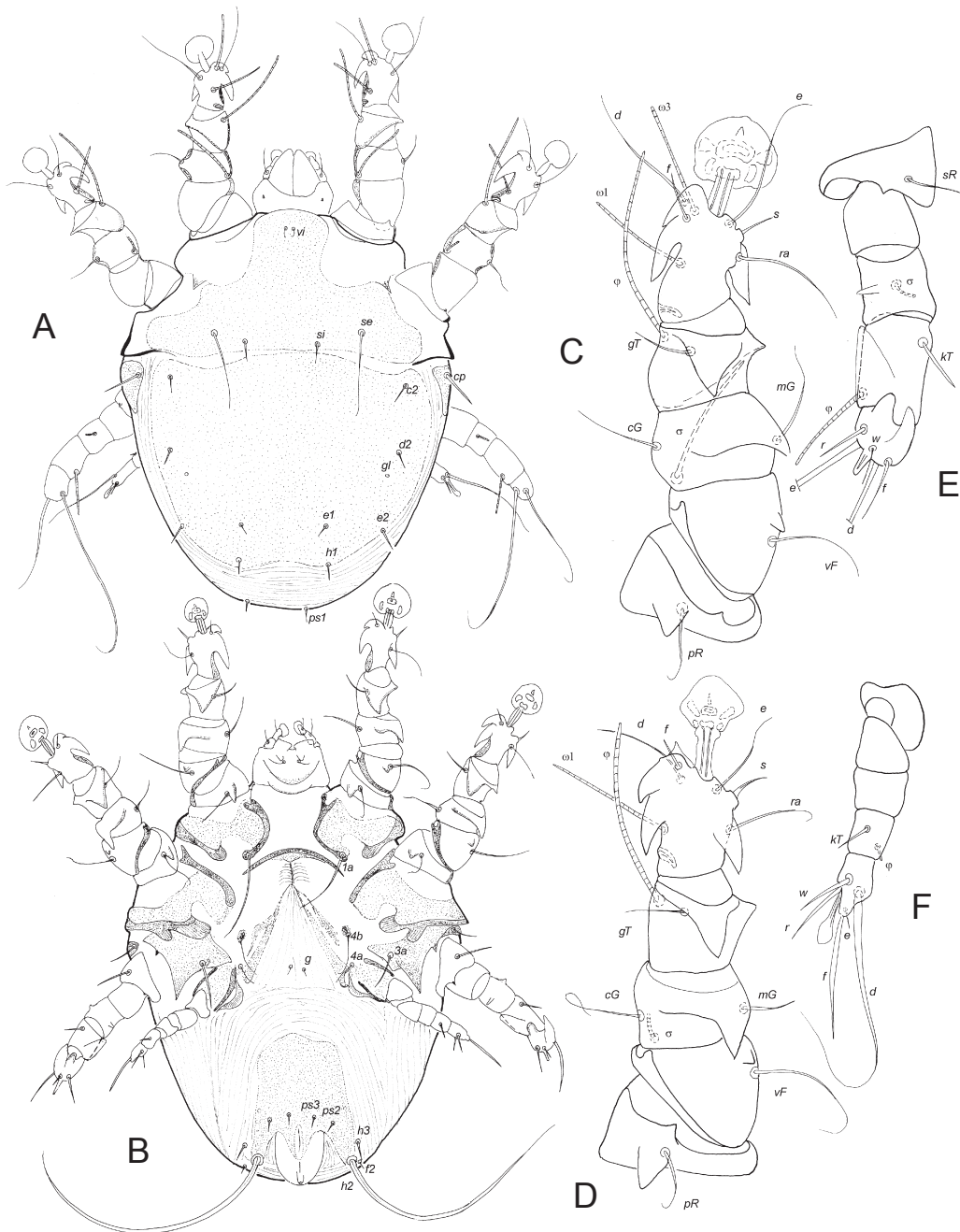


Fig. 84. *Listropsoralges caluromys* Bochkov et Wauthy, 2009, female: A — dorsal view; B — ventral view; C–F — legs I–IV in ventral view respectively (after Bochkov and Wauthy (2009) with minor modifications).

Subfamily Dromiciocoptinae Fain, 1970

Type genus: *Dromiciocoptes* Fain, 1970.

Table 16, Figs. 85–87

Diagnosis. *Both sexes.* Cheliceral hood and ventral apophysis of movable digit present. Idiosoma dorso-ventrally flattened. Supracoxal sclerite and supracoxal opening indistinct, setae *scx* present. Propodonal and hysteronotal shields present, distinctly developed. Genital papillae present. Intercoxal attaching organs absent. Anal opening situated ventrally, close to posterior margin of opisthosoma. Legs I and II without clasp-

organs with normally developed pretarsi. Ambulacral sclerites discernible. Female legs III and IV and legs III of males strongly modified as hair-clasping organs; femora of these legs strongly reduced and deeply inserted into respective trochanters; genua not modified; tibiae flattened; tarsi fold back over ventral surface of respective tibiae and also flattened. Tarsi bearing distinct pretarsi, which strongly curved ventrally, flattened and armed with sclerotized crests.

Female. Opisthogastral shield present. Ovipore Y-shaped. Epigynum distinctly developed. Oviparous.

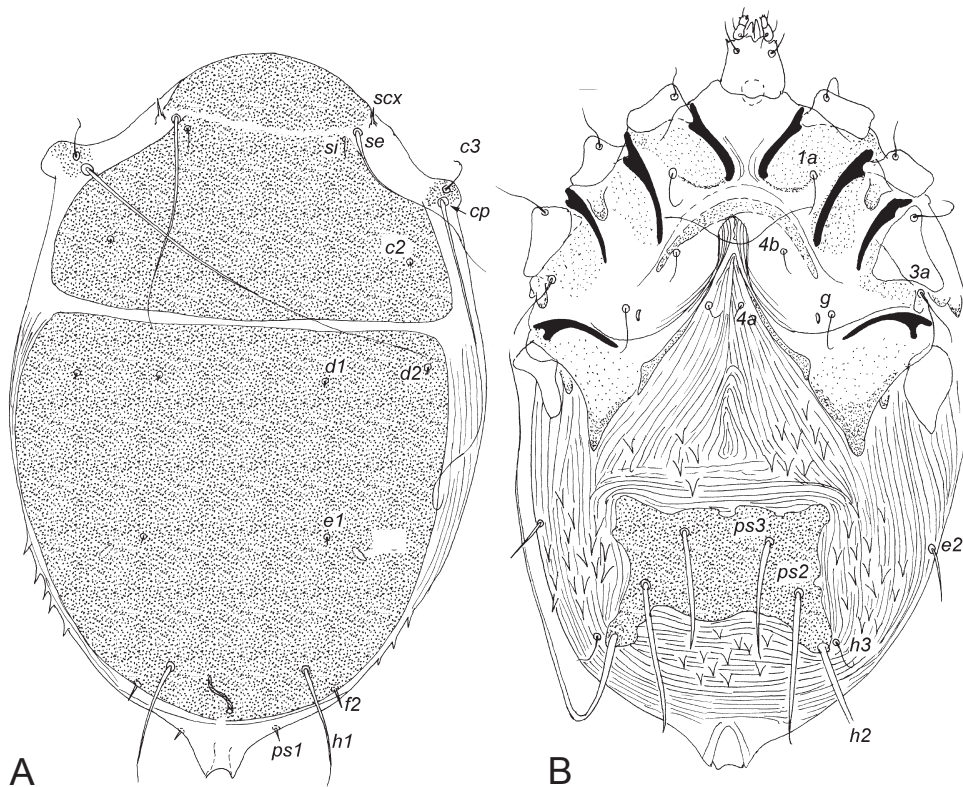


Fig. 85. *Dromicioptes marmosops* Bochkov et OConnor, 2008, female: A — dorsal view, B — ventral view (after Bochkov and OConnor (2008), with minor modifications).

Male. Opisthosomal lobes present, paranal suckers present. Legs IV without ambulacra. Legs III and IV normally developed. Setae *d*IV filiform and *e*IV alveoli.

Taxa included. Only type genus with 3 species.

Associations with hosts: inhabitants of the skin of marsupial mammals, Didelphimorphia (Didelphidae) and Paucituberculata (Caenolestidae).

Distribution. South America.

Main references. Fain (1970b) — diagnosis of Dromicioptinae, Bochkov and OConnor (2008) — comparative morphology of Dromicioptinae.

Families of Psoroptid complex

Diagnosis. *Both sexes:* Cheliceral hood and ventral apophysis of movable digit present. Dorso-apical spur on tarsi I and II distinctly developed, reduced or absent (in this case setae *s*I and II angle-like). Intercoxal striate membranes between coxal fields I and II absent. Setae *h*1 absent. Genital papillae present. Anal opening situated ventrally. Central sclerite of ambulacral discs of legs I and II with widely rounded or almost straight anterior margin. Females oviparous. Males with

opisthosomal lobes and paranal suckers (in most Cebalginae paranal suckers absent).

Family Psoroptidae Canestrini, 1892

Type genus: *Psoroptes* Cervais, 1841.

Table 17, Figs. 88–104

Diagnosis. *Both sexes.* Idiosoma dorso-ventrally flattened. Supracoxal sclerite and supracoxal opening distinct, setae *scx* present or absent. Setae *si* situated close to seta *se* bases. Apodemes Ia not fused into sternum (fused in some Makialginae). Legs I–II without clasping organs or with distinct retrorse projections. Ambulacral sclerites discernible. Dorso-apical projection distinctly developed. Setae *s*I and II filiform, rarely hook-like in some Cebalginae and Psoralginae.

Female. Hysteronotal shield present or absent. Opisthosomal lobes absent but in some genera posterior margin of opisthosoma with narrow projections. Ovipore Y-shaped or in form of transverse slit (most Psoroptinae). Epigynum distinctly developed or absent (most Psoroptinae). Oviparous.

Male. Hysteronotal shield distinctly developed. Legs IV shorter than legs III or both leg pairs normally developed. Setae *d*IV and *e*IV sucker-like or filiform.

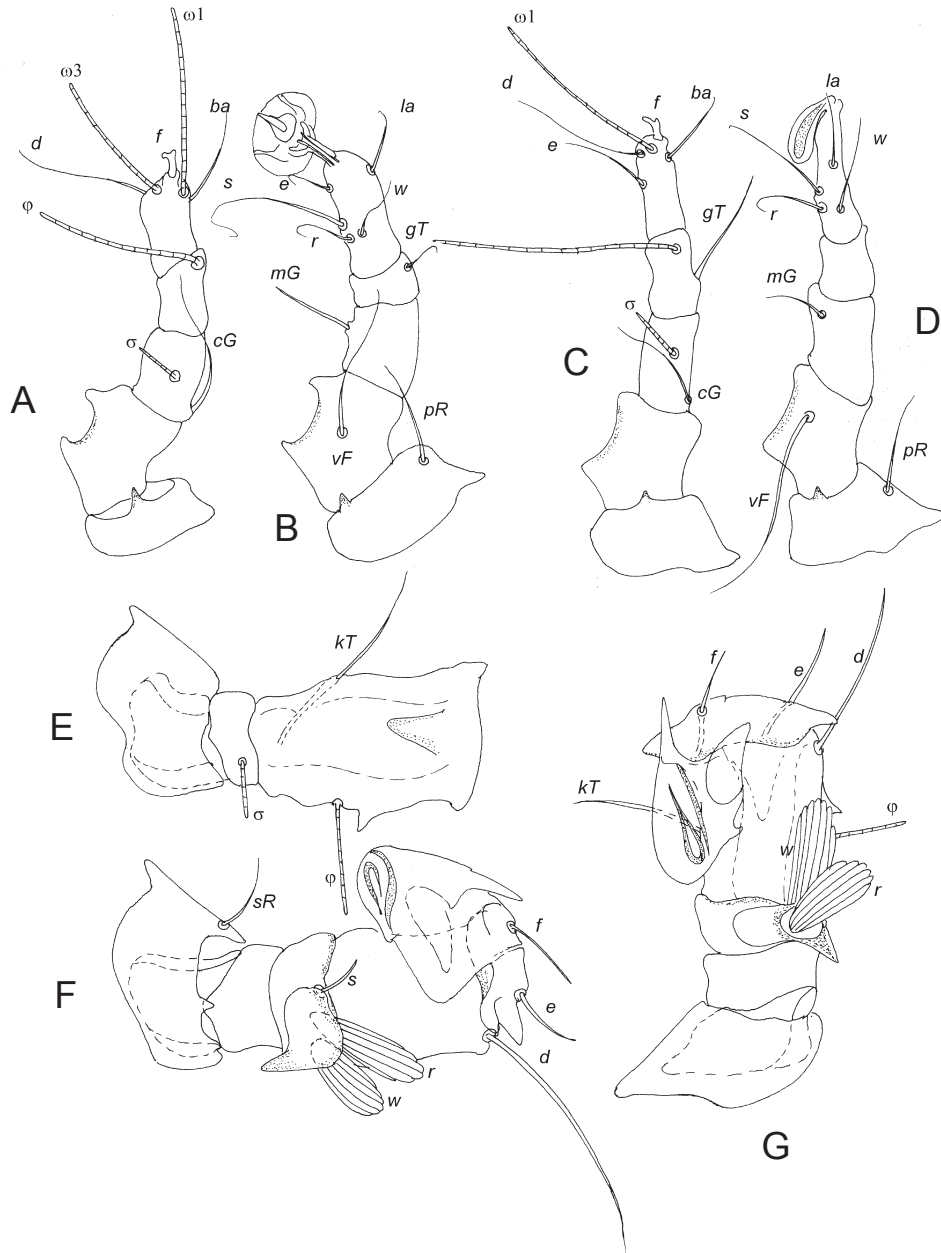


Fig. 86. *Dromicicoptes marmosops* Bochkov et OConnor, 2008, legs of female: A — leg I in dorsal view, B — same in ventral view, C — leg II in dorsal view, D — same in ventral view, E — leg III in dorsal view, F — same in ventral view, G — leg IV in ventral view (after Bochkov and OConnor (2008), with minor modifications).

Taxa included: 29 genera and 50 species, Cebalginae Fain, 1962 — *Alouattalges* Fain, 1963 (1 species), *Cebalges* Fain, 1962 (1 species), *Cebalgoides* Fain, 1963 (1 species), *Fonsecalges* Fain, 1962 (2 species), *Procebalges* Fain, 1963 (1 species), *Schizopodalges* Fain, 1963 (1 species); Makialginae Gaud et Mouchet, 1959 — *Cheiragalges* Fain, 1963 (1 species), *Daubentonialges* Fain, 1972 (1 species), *Galagalges* Fain, 1963 (1 species), *Gaudalges* Fain, 1963 (4 species), *Lemuralges* Fain, 1963 (1 species), *Makialges* Gaud et Till, 1957 (3 species); Marsupialginae Fain, 1963 — *Marsupialges* Fain, 1963 (1 species); Nasalial-

ginae Fain et Nadchatram, 1979 — *Nasalialges* Fain et Nadchatram, 1979 (1 species); Psoralginae Oudemans, 1908 — *Acaroptes* Womersley, 1953 (2 species), *Coendalges* Fain et Mendez, 1979 (1 species), *Edentalges* Fonseca, 1954 (3 species), *Myoproctalges* Fain et Lukoschus, 1974 (1 species), *Psoralges* Trouessart, 1896 (2 species), Psoroptinae Canestrini, 1892 — *Caparinia* Canestrini, 1894 (5 species), *Chorioptes* Gervais et van Beneden, 1859 (5 species), *Choriopterosoptes* Sweatman, Walker et Bindernagel, 1964 (2 species), *Choriotodectes* Fain, 1975 (1 species), *Echimyralges* Fain, 1967 (1 species), *Hyraoptes*

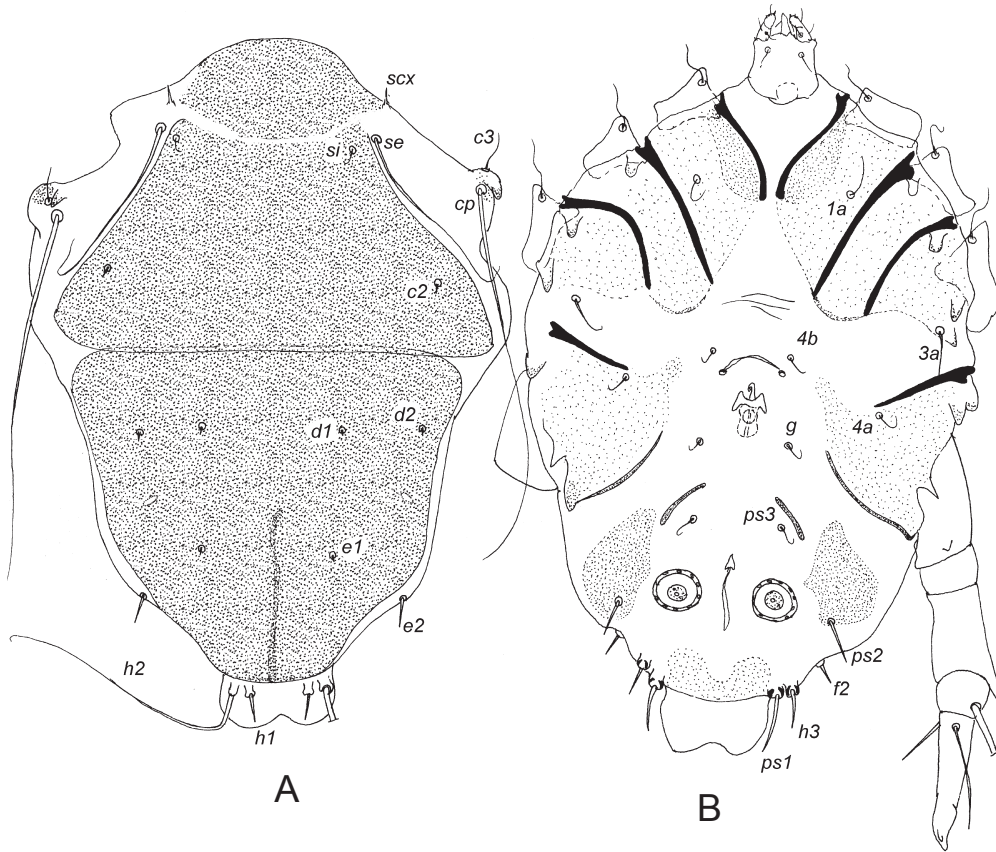


Fig. 87. *Dromicicoptes marmosops* Bochkov et OConnor, 2008, male: A — dorsal view, B — ventral view (after Bochkov and OConnor (2008), with minor modifications).

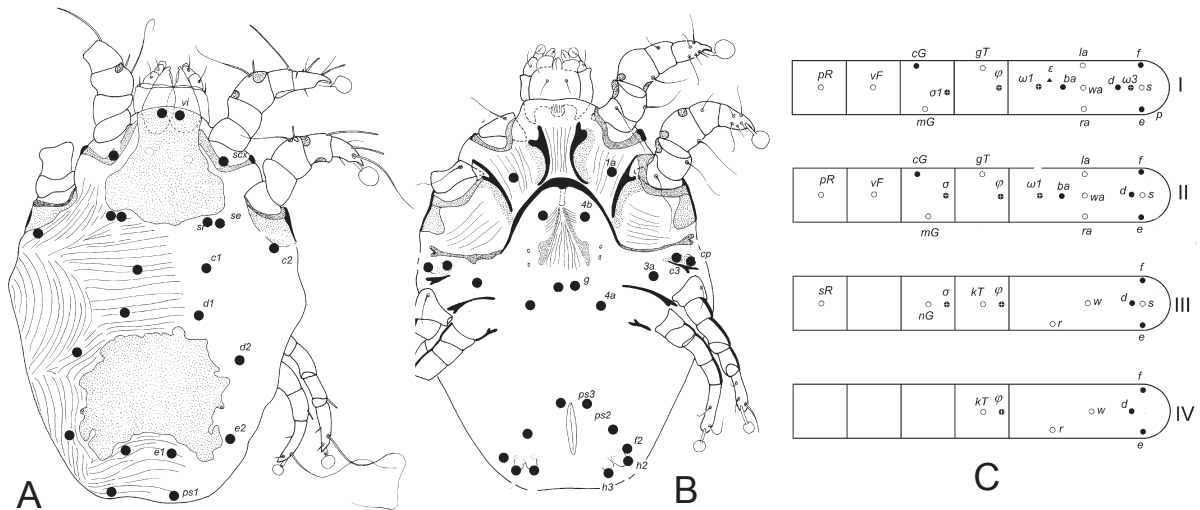


Fig. 88. Psoroptidae, scheme of setae: A — idiosoma in dorsal view, B — same in dorsal view, C — legs I-IV.

Fain et Lukoschus, 1981(1 species), *Otodectes* Canestrini, 1894 (1 species), *Psorochorioptes* Fain, 1963 (1 species), *Psoroptes* Gervais, 1841 (3 species), *Trouessalges* Fonseca, 1954 (1 species).

Associations with hosts: inhabitants of the skin of therian mammals, Cebalginae — Placentalia: Primates (Atelidae, Cebidae, Pitheciidae); Maki-alginae — Placentalia: Primates (Cheirogaleidae, Daubentoniidae, Galagidae, Indriidae, Lemuridae,

Lepilemuridae), Marsupialginae — Marsupialia: Didelphimorphia (Didelphidae); Nasialginae — Placentalia: Primates (Cercopithecidae); Psoralginae — Placentalia: Pilosa (Bradypodidae, Megalonychidae, Myrmecophagidae), Rodentia (Dasyproctidae), Marsupialia: Diprotodontia (Vombatidae); Psoroptinae — Placentalia: Artiodactyla (different families), Carnivora (different families), Erinaceomorpha (Erinaceidae), Hyra-

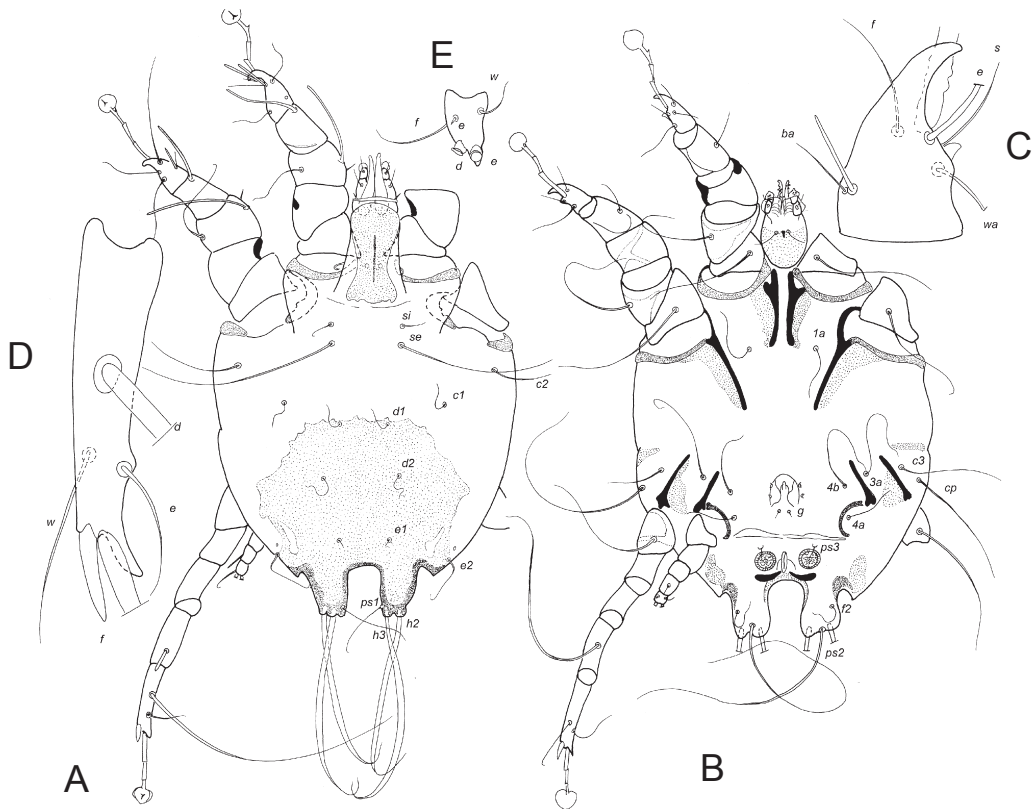


Fig. 89. *Psoroptes pienaari* Fain, 1970, male: A — dorsal view, B — ventral view, C — tarsus I in dorsal view, D — tarsus III in dorsal view, E — tarsus IV in ventral view.

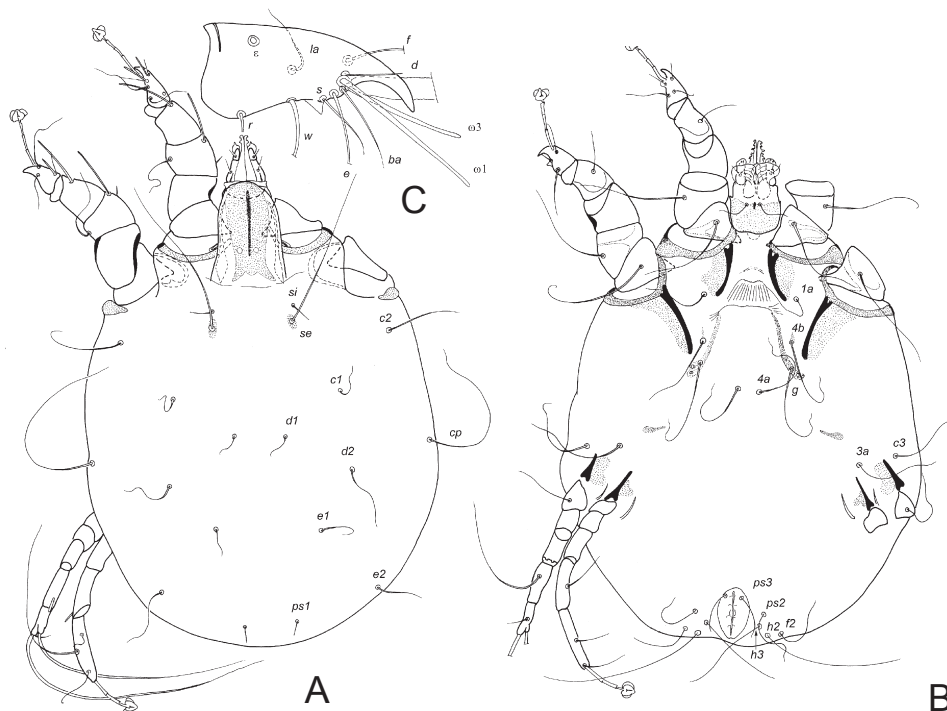


Fig. 90. *Psoroptes pienaari* Fain, 1970, female: A — dorsal view, B — ventral view, C — tarsus I in dorsal view.

coidea (Procaviidae), Lagomorpha (Leporidae), Perissodactyla (different families), Rodentia (Echimyidae).

Distribution. Cebalginae — South America; Makialginae — Africa (including Madagascar);

Marsupialginae — South America; Nasalialginae — Asia; Psoralginae — South America and Australia; Psoroptinae — cosmopolites.

Remarks. (i) The validity of most species of the genus *Psoroptes* is a subject for lively discus-

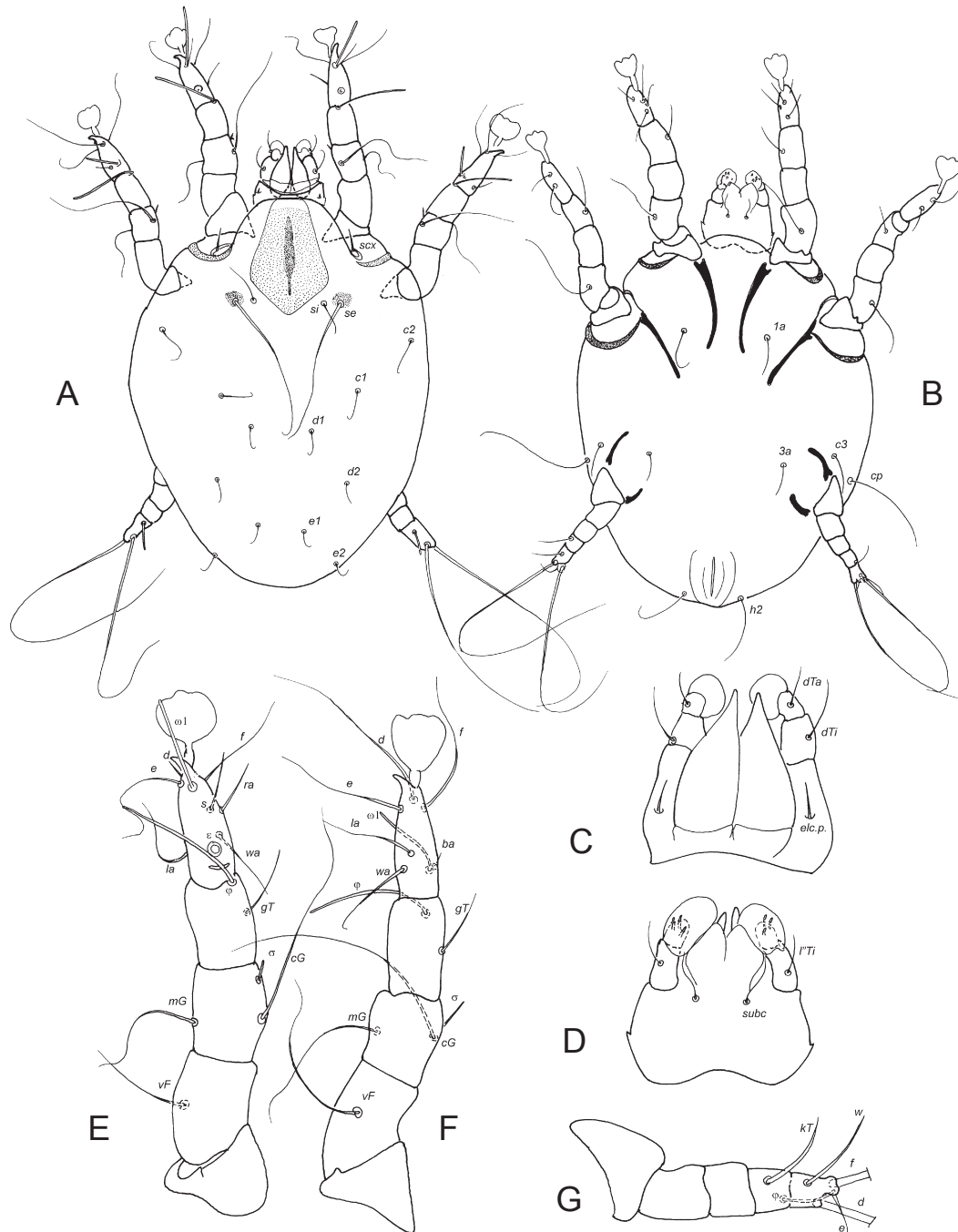


Fig. 91. *Otodectes cynotis* (von Hering, 1838), larva: A — dorsal view, B — ventral view, C — gnathosoma in dorsal view, D — same in ventral view, E — leg I in ventral view, F — leg II in ventral view, G — leg III in ventral view.

sion. The morphological and molecular studies provide solid evidences for conspecificity of four *Psoroptes* species: *Psoroptes equi* (von Hering, 1838), *Psoroptes ovis* (von Hering, 1838), *Psoroptes cuniculi* (Delafond, 1859), and *Psoroptes cervinus* (Ward, 1915) (Zahler et al. 2000; Ramey et al. 2000; Pegler et al. 2005). Three latter species and *Psoroptes natalensis* (Hirst, 1919) were synonymized by Zahler et al. (2000) with *Psoroptes equi* (von Hering, 1838). However, Wall and Kolbe (2006) clearly showed that name of *Psoroptes*

ovis has priority. At the same time, *Psoroptes natalensis* was synonymized by Zahler et al (2000) based entirely on very rough morphological comparison concerning just a few structures, whereas molecular sequences for this species were not obtained (Pegler et al. 2005). To present, I prefer consider it as a separate species. Thus, I include in the genus *Psoroptes* three species: *P. ovis*, *P. natalensis*, and *P. pienaarri* (Fain, 1970), but underline that status of the two last species need in additional examinations.

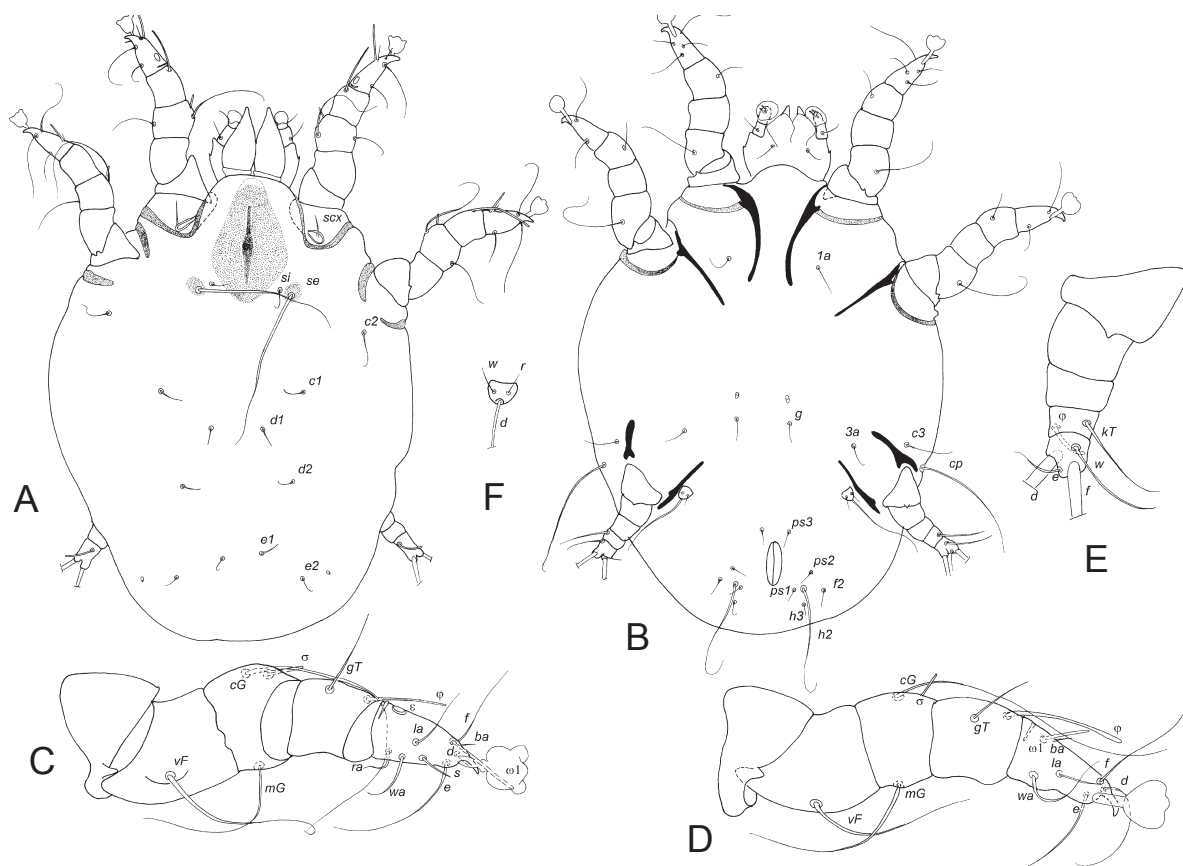


Fig. 92. *Otodectes cynotis* (von Hering, 1838), protonymph: A — dorsal view, B — ventral view, C — leg I in ventral view, D — leg II in ventral view, E — leg III in ventral view, D — leg IV in ventral view.

(ii) The distinctions between *Chorioptes bovis* (von Hering, 1845) and *Chorioptes japonensis* Takahashi et Nogami, 2001 from *Capricornis crispus* (Bovidae) from Japan (Takahashi et al. 2001) are absolutely insignificant and keep within limits of the intraspecific variability of such morphologically variable species as *Chorioptes bovis*. Therefore I consider this species as a junior synonym of *C. bovis*. It should be also mentioned that conclusions by Zahler et al. (2001) about uncertain status of *Chorioptes* spp. other than *Chorioptes bovis* and *C. texanus* Hirst, 1924 were made without deep morphological analysis and without investigation of type series and, therefore, are not taken here into consideration.

Main references. Fain (1963) — review of Psoroptidae associated with Primates, Fain (1965b) — review of Psoroptidae associated with marsupials and edentates, Fain (1975) — key to Psoroptinae, OConnor (1984) — phylogeny of Cebalginae, Bochkov et al. (2010, in press). — revision of Malkialginae.

**Key to genera of the family Psoroptidae
Canestrini, 1892**

- 1. *Both sexes*: Setae *vi* present 13
- *Both sexes*: Setae *vi* absent 2

- 2. *Both sexes*: Setae *la*II and solenidion σ III absent 12
- *Both sexes*: Setae *la*II and solenidion σ III present Psoroptinae Canestrini, 1892
- 3. *Both sexes*: Tarsi III with 4 setae (setae *s* and *r* absent). *Female*: Epigynum absent or almost reduced. Ovipore in shape of transversal slit. *Male*: setae *f*III stout, strongly thickened 6
- *Both sexes*: Tarsi III with 6 setae (setae *s* and *r* present). *Female*: Epigynum present, distinctly developed, arch-like. Ovipore in shape of an inverted Y. *Male*: setae *f*III filiform 4
- 4. *Females*: Hysteronotal shield present. Tarsi III with 1 ultralong seta. Tarsi III and IV with pretarsi. *Male*: Opisthosomal lobes adjoining. Pretarsi IV present 5
- *Females*: Hysteronotal shield absent. Tarsi III with 3 ultralong setae. Tarsi III and IV without pretarsi. *Male*: Opisthosomal lobes distinctly separated from each other. Pretarsi IV absent *Trouessalges* Fonseca, 1954
- 5. *Both sexes*: Setae *c*1 present. Genu I with 2 solenidia (solenidion σ 2 present). *Female*: Hysteronotal shield weakly developed, its anterior margin situated posterior to level of opisthosomal glands. *Male*: Tarsi III with 2 setae those length

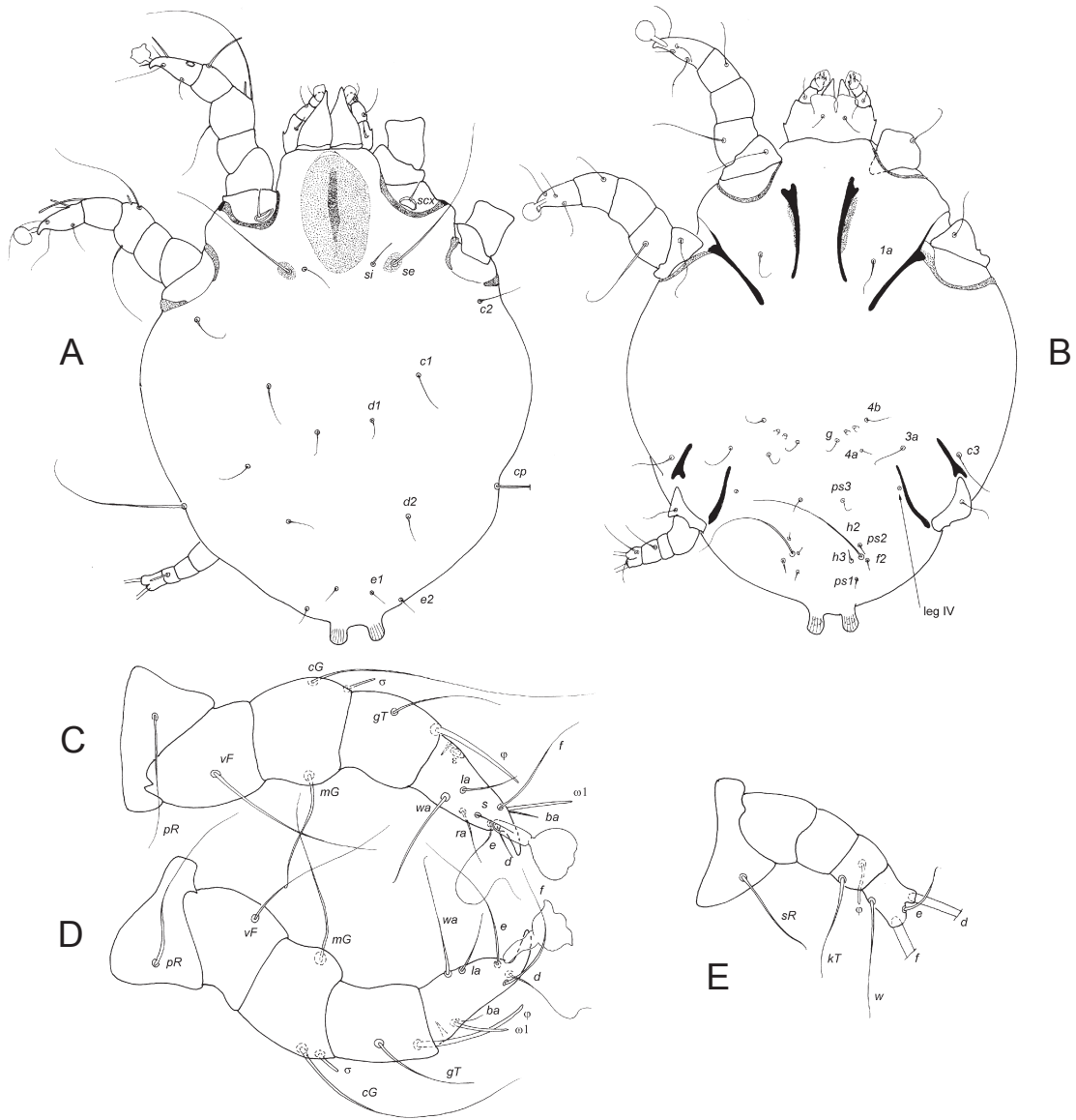


Fig. 93. *Otodectes cynotis* (von Hering, 1838), tritonymph: A — dorsal view, B — ventral view, C — leg I in ventral view, D — leg II in ventral view, E — leg III in ventral view.

more than ½ length of legs IV
 *Hyracoptes* Fain et Lukoschus, 1981
 — *Both sexes*: Setae *c1* absent. Genu I with 1 solenidion (solenidion $\sigma 2$ absent). *Female*: Hysteronotal shield distinctly developed, its anterior margin situated far anterior to level of opisthosomal glands. *Male*: Tarsi III with 1 seta, over half long as legs IV *Echimyalgus* Fain, 1967
 6. *Both sexes*: Tarsi I and II with not segmented pretarsal stalks 7
 — *Both sexes*: Tarsi I and II with segmented pretarsal stalks *Psoroptes* Gervais, 1841
 7. *Both sexes*: Tarsi III with 1 ultralong seta which length more than ½ length of legs IV 9
 — *Both sexes*: Tarsi III with 2 or 3 ultralong setae which length much longer than ½ length of legs IV 8

8. *Both sexes*: Tarsi III with 3 ultralong setae which length much more than ½ length of legs IV. *Males*: Opisthosomal lobes moderately developed *Caparinia* Canestrini, 1894
 — *Both sexes*: Tarsi III with 2 ultralong setae which length much more than ½ length of legs IV. *Males*: Opisthosomal lobes almost reduced
 *Otodectes* Canestrini, 1894
 9. *Both sexes*: Chelicerae moderately developed, cheliceral and palpal apices situated approximately at same level 10
 — *Both sexes*: Chelicerae strongly elongated, their apices situated far anterior to palpal apices
 *Choriopsoroptes* Sweatman, Walker et Bindernagel, 1964
 10. *Female*: Setae *ps3* present. *Male*: Pretarsi IV present 11

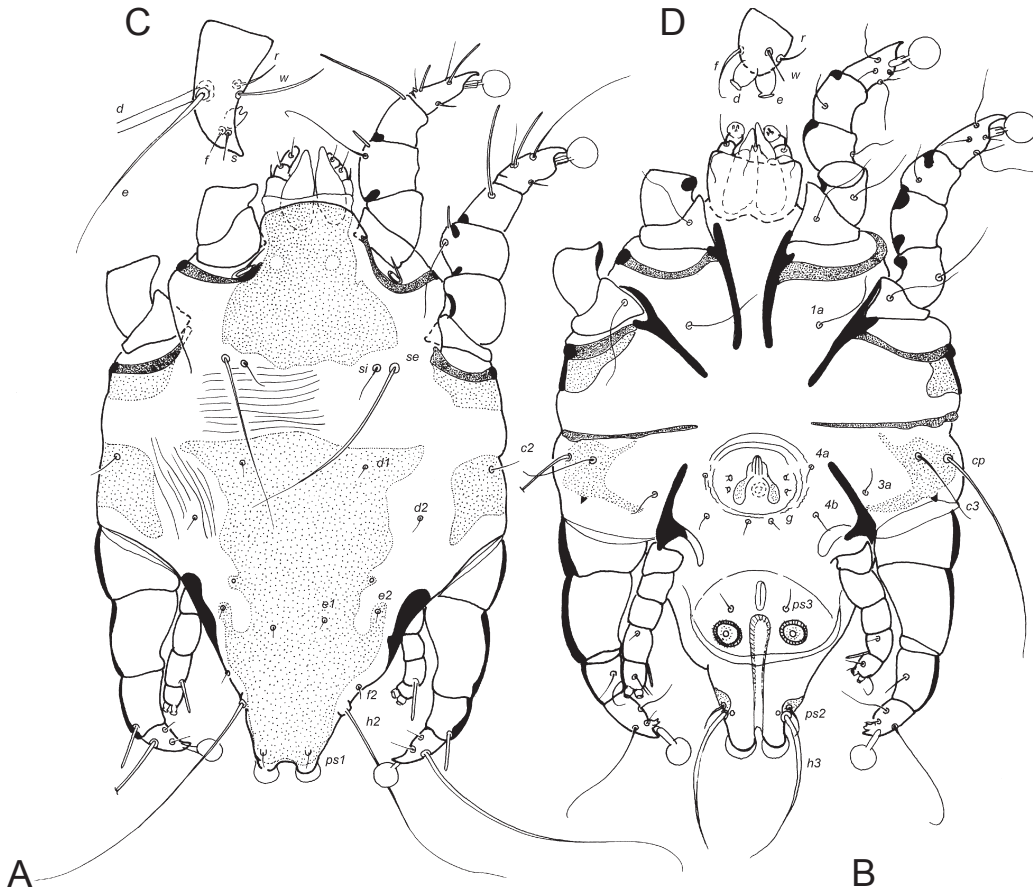


Fig. 94. *Echimyalgus belluominii* Fain, 1967, male: A — dorsal view, B — ventral view, C — tarsus III in ventral view, D — tarsus IV in ventral view.

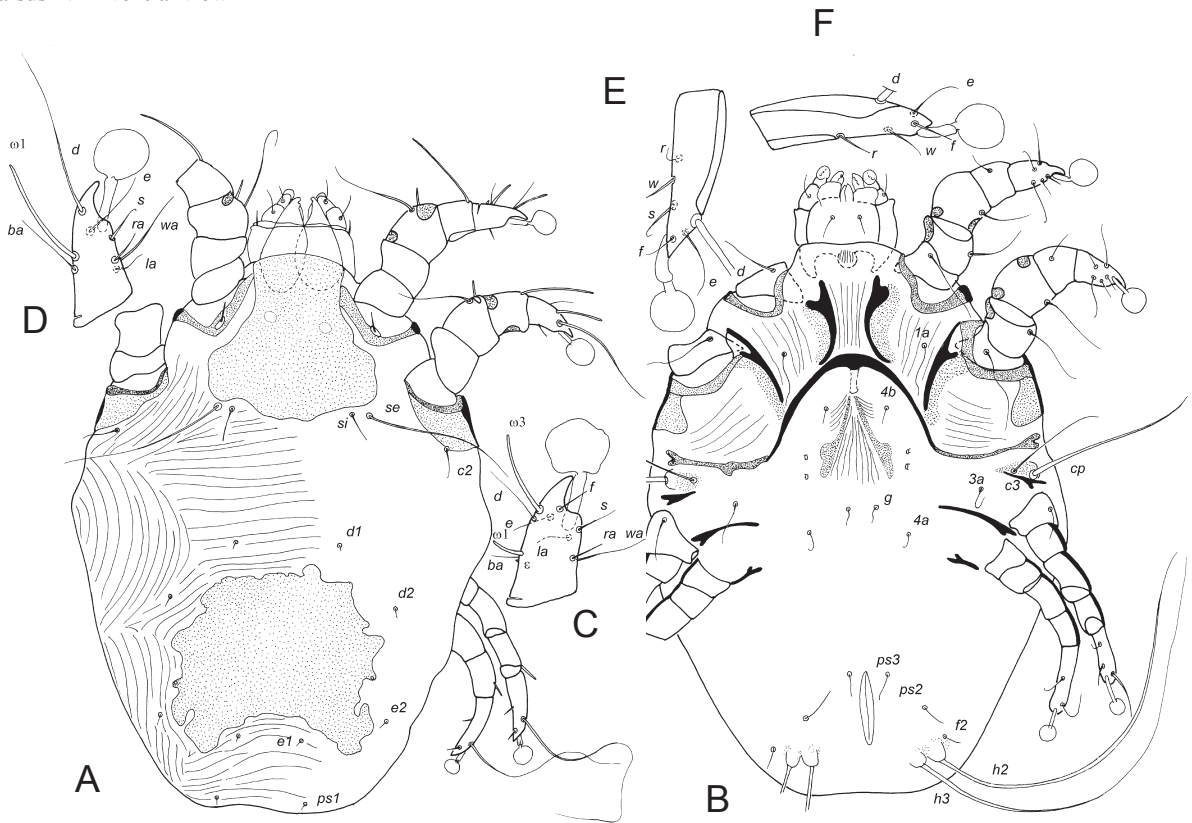


Fig. 95. *Echimyalgus belluominii* Fain, 1967, female: A — dorsal view, B — ventral view, C — tarsus I in dorsal view, D — tarsus II in dorsal view, E — tarsus III in lateral view, F — tarsus IV in lateral view.

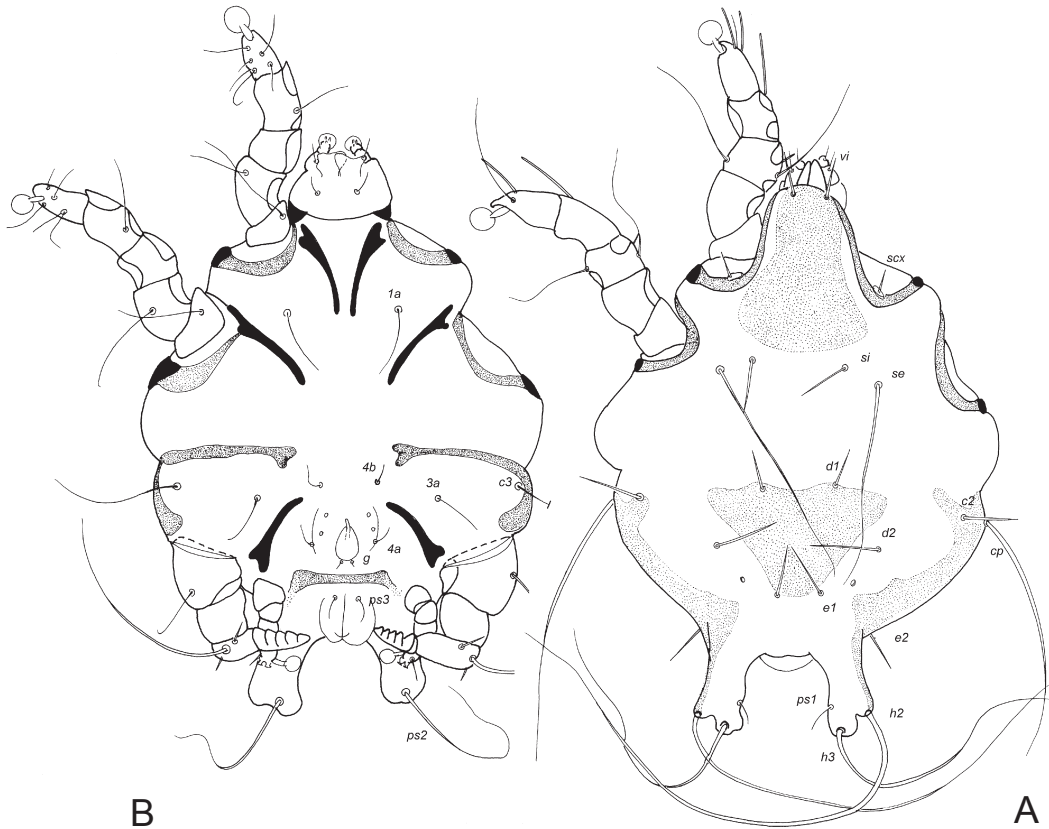


Fig. 96. *Cebalges gaudi* Fain, 1962, male: A — dorsal view, B — ventral view.

— *Female*: Setae *ps3* absent. *Male*: Pretarsi IV absent *Psorochoiropes* Fain, 1963
 11. *Female*: Tibia and tarsus III fused to each other. Pretarsi IV absent. *Male*: Opisthosomal lobes trapezoidal form. Legs IV 3 times shorter than legs III *Choriotodectes* Fain, 1975
 — *Female*: Tibia and tarsus III not fused to each other. Pretarsi IV present. *Male*: Opisthosomal lobes triangular form. Legs IV 2 times shorter than legs III
 *Chorioptes* Gervais et van Beneden, 1859
 12. *Both sexes*: Setae *scx* and *baI*, II absent. Legs I and II with short retrorse projections. *Female*: Setae *4b* present. Posterior opisthosomal margin widely rounded, without protrusions. Pretarsi III absent. *Male*: Opisthosomal lobes short, much shorter than legs IV Marsupialginae Fain, 1963 (*Marsupialges* Fain, 1963)
 — *Both sexes*: Setae *scx* and *baI*, II present. Legs I and II without projections. *Female*: Setae *4b* absent. Posterior opisthosomal margin with short protrusions. Pretarsi III present. *Male*: Opisthosomal lobes elongated, longer than legs IV Nasalialginae Fain et Nadchatram, 1979 (*Nasalialges* Fain et Nadchatram, 1979)
 13. *Both sexes*: Solenidia ω I and II situated in median or basal part of respective tarsi (in *Coendal-*

ges in apical part). Setae *baI*, II present. *Female*: Opisthosoma without lateral projections. *Male*: Paranal suckers present (absent in *Galagalges*). Tibia and tarsi of legs III not fused to each other, without ridges 19
 — *Both sexes*: Solenidia ω I and II situated in apical part of respective tarsi. Setae *baI*, II absent. *Female*: Opisthosoma with lateral projections. *Male*: Paranal suckers absent (present in *Procebalges*). Tibia and tarsi of legs III fused to each other, with ridges ... Cebalginae Fain, 1962 14
 14. *Females*: Setae *sI* and II filiform. *Male*: Spur on tarsi III absent. Ridges covering tarsus and tibia III 16
 — *Females*: Setae *sI* and II claw-like. *Male*: Spur on tarsi III present. Ridges covering only tibia III 15
 15. *Female*: Hysteronotal shield absent. Epigynum fused with apodemes Ia. Opisthosoma without lateral projections. *Male*: Opisthosomal lobes distinctly developed, their apical parts fused to each other *Schizopodalges* Fain, 1963
 — *Female*: Hysteronotal shield present. Epigynum not fused with apodemes Ia. Opisthosoma with short lateral projections. *Male*: Opisthosomal lobes very short and separated from each other
 *Alouattalges* Fain, 1963

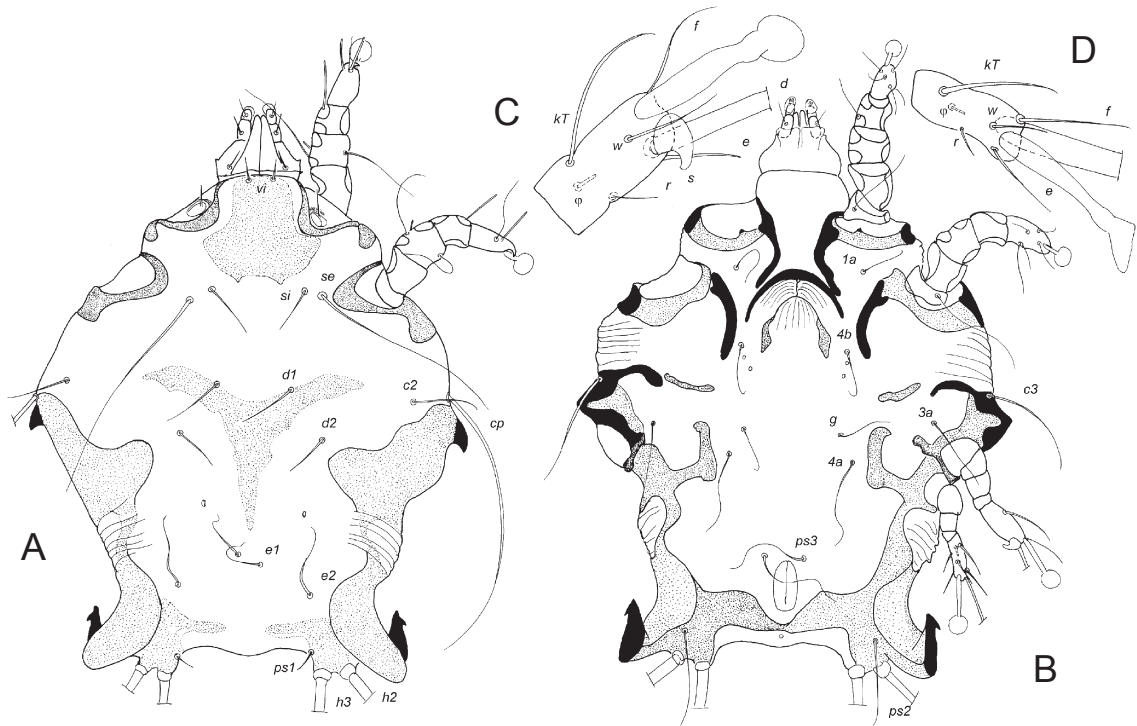


Fig. 97. *Cebalges gaudi* Fain, 1962, female: A — dorsal view, B — ventral view, C — tibia-tarsus III in ventral view, D — tibia-tarsus IV in ventral view.

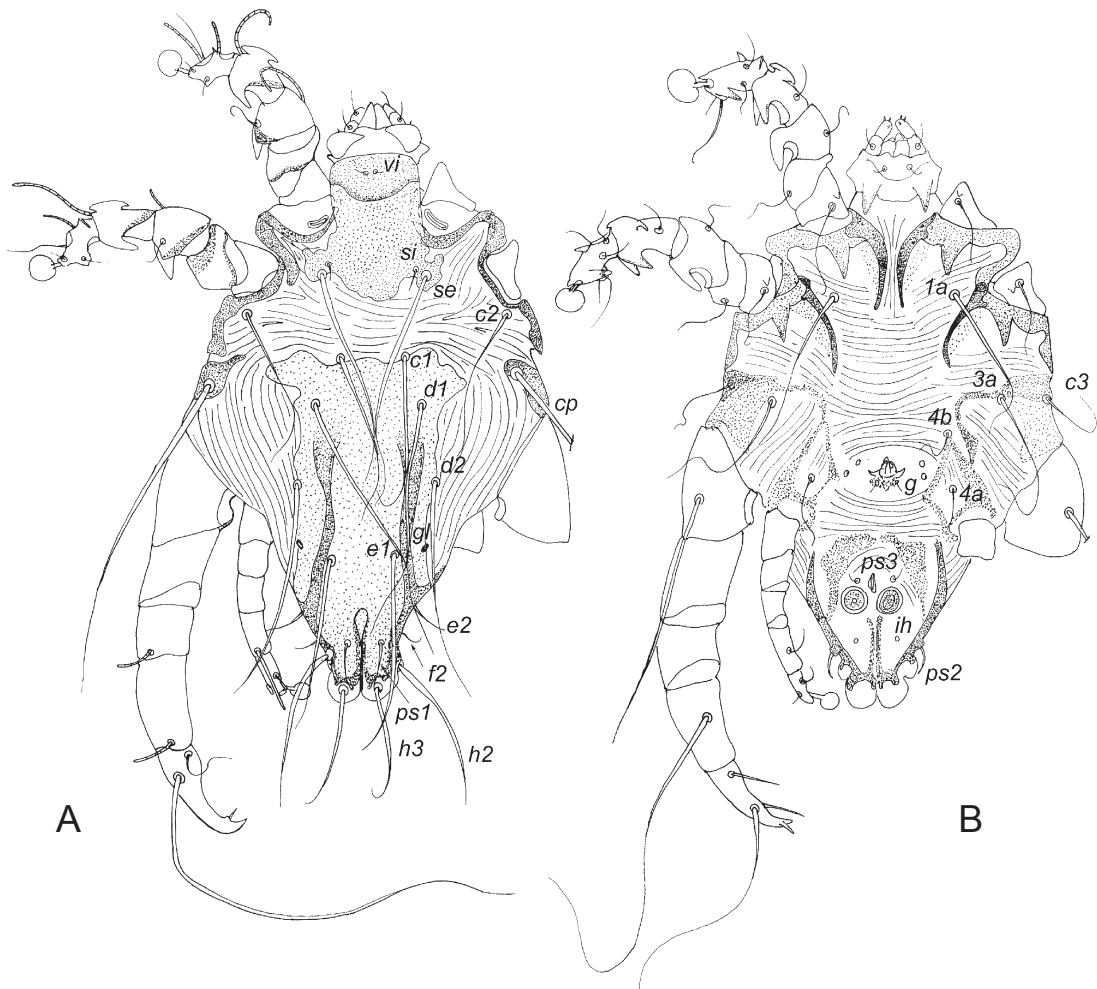


Fig. 98. *Makialges sternodons* Gaud et Till, 1957, male: A — dorsal view, B — ventral view



Fig. 99. *Makialges lepitemuri* Gaud et Till, 1957, female: A — dorsal view, B — ventral view.

16. *Both sexes*: Posterior ventral projection of subcapitulum absent or if present, pointed apically. *Female*: Epigynum not fused with apodemes Ia. Lateral opisthosomal projection curved ahead, with apical hook. *Males*: Width of opisthosomal lobes monotonous. Paranal suckers absent. Tarsi III with or without ambulacral stalk but always without ambulacral disc 17
 — *Both sexes*: Posterior ventral projection of subcapitulum present, widely rounded. *Female*: Epigynum fused with apodemes Ia. Lateral opisthosomal projection not curved, without apical hook. *Males*: Opisthosomal lobes enlarged in subapical part. Paranal suckers present. Tarsi III with ambulacral stalk and ambulacral disc
 *Procebalges* Fain, 1963
 17. *Female*: Subcapitulum without ventral projections. Tibia and tarsus III and IV fused (4 articulate segments). *Male*: Transverse sclerotized band present between anal and genital openings. Setae *f2* absent 18
 — *Female*: Subcapitulum with pair of ventral projections. Tibia and tarsus III and IV not fused (5 articulate segments). *Male*: Transverse sclerotized band absent between anal and genital openings. Setae *f2* present *Cebalgoides* Fain, 1963
 18. *Female*: Setae *ps1* present. Pretarsi III and IV present. *Male*: Subcapitulum with or without pair

of ventral projections. Setae *h3* about 2 times shorter than *h2* *Cebalges* Fain, 1962
 — *Female*: Setae *ps1* absent. Pretarsi III and IV absent. *Male*: Subcapitulum without ventral projections. Setae *h3* and *h2* subequal
 *Fonsecalges* Fain, 1962
 19. *Both sexes*: Legs I and II with retrorse protrusions. Setae *ba1* spur-like 24
 — *Both sexes*: Legs I and II without retrorse protrusions. Setae *ba1* filiform 20
 20. *Both sexes*: Pretarsi of legs III and IV present 22
 — *Both sexes*: Pretarsi of legs III and IV absent ...
 21
 21. *Both sexes*: Setae *d1* and *e1* absent. *Female*: Tibia and tarsi III and IV fused. *Male*: Dorso-apical hook absent, setae *s* and *f1*, II claw-like
 *Acaroptes* Womersley, 1953
 — *Both sexes*: Setae *d1* and *e1* present. *Female*: Tarsi III and IV strongly shortened but not fused with respective tibiae. *Male*: Dorso-apical hook present, setae *s* and *f1*, II filiform
 *Edentalges* Fonseca, 1954
 22. *Female*: Posterior margin of opisthosoma without distinct median incision. Setae *dIII* whip-like, longer than legs III. *Male*: Setae *c2*, *d1*, *d2*, *e1*, and *e2* filiform. Opisthosoma with 2 pairs of ultralong setae subequal to setae *cp*. Tarsi III

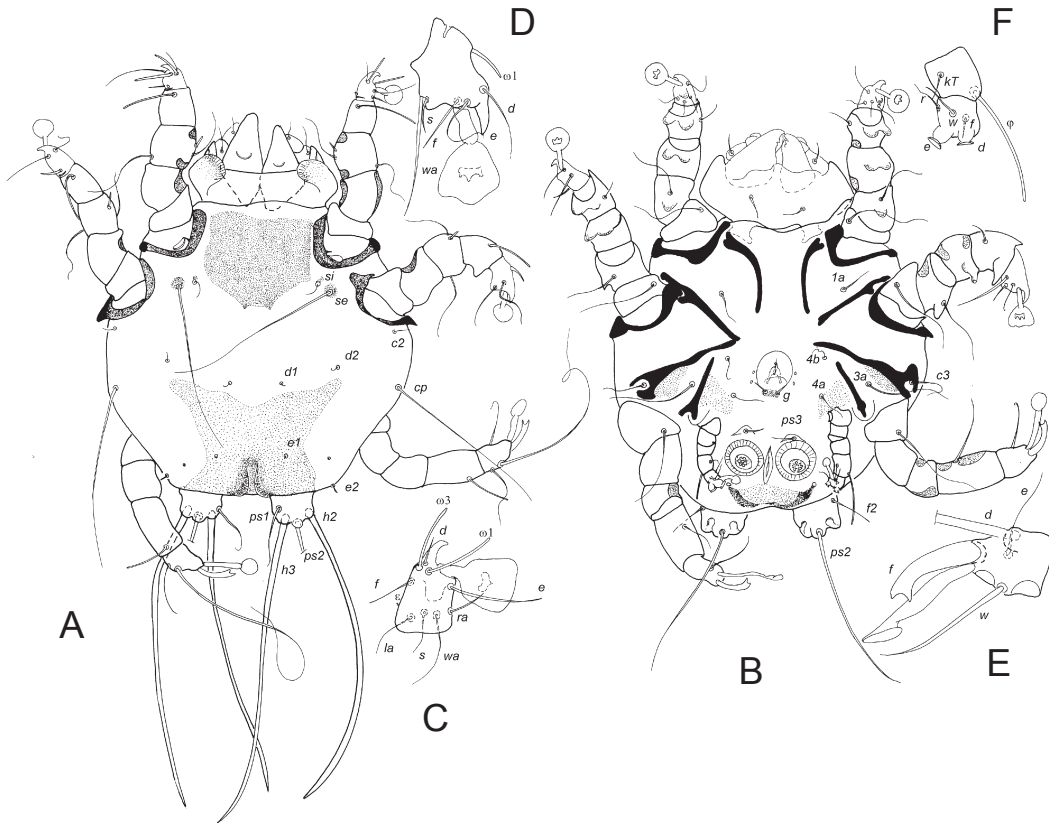


Fig. 100. *Marsupialges misonnei* Fain, 1963, male: A — dorsal view, B — ventral view, C — tarsus I in ventral view, D — tarsus II in ventral view, E — tarsus III in ventral view, F — tarsus IV in ventral view.

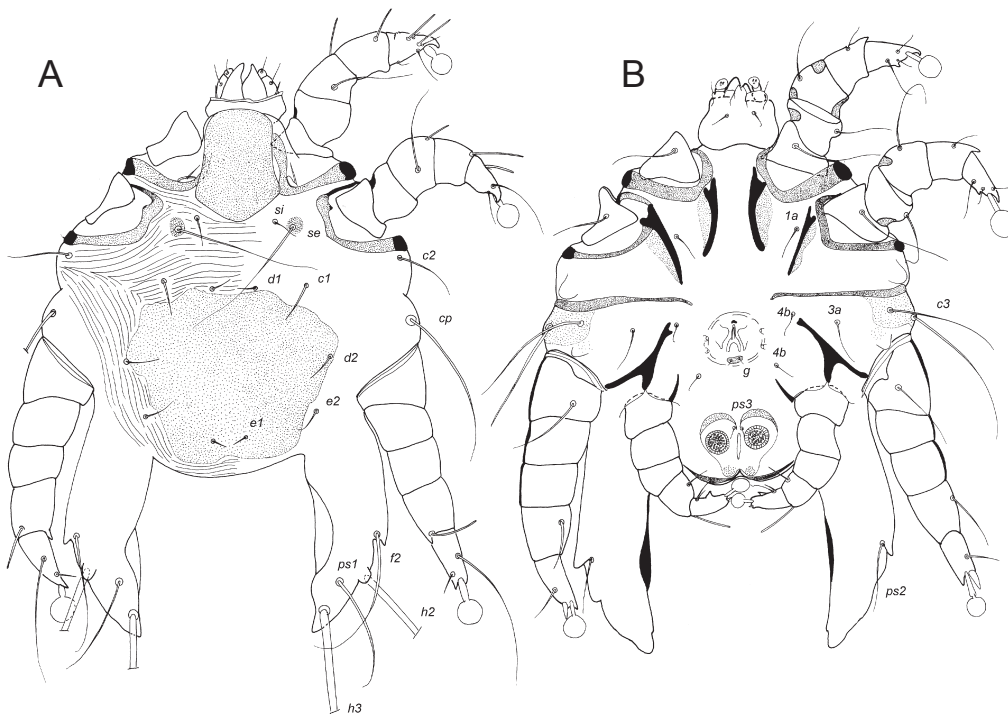


Fig. 101. *Nasaliagalges borneensis* Fain et Nadchatram, 1979, male: A — dorsal view, B — ventral view.

straight, without ventral projection near to seta *s* base 23
 — *Female*: Posterior margin of opisthosoma with distinct median incision. Setae *d*III short, shorter than tarsus III. *Male*: Setae *c*2, *d*1, *d*2, *e*1, and *e*2

slightly inflated. Opisthosoma with 1 pair of ultralong setae subequal to setae *cp*. Tarsi III slightly curved, with short ventral projection near to seta *s* base
 *Coendalges* Fain et Mendez, 1979

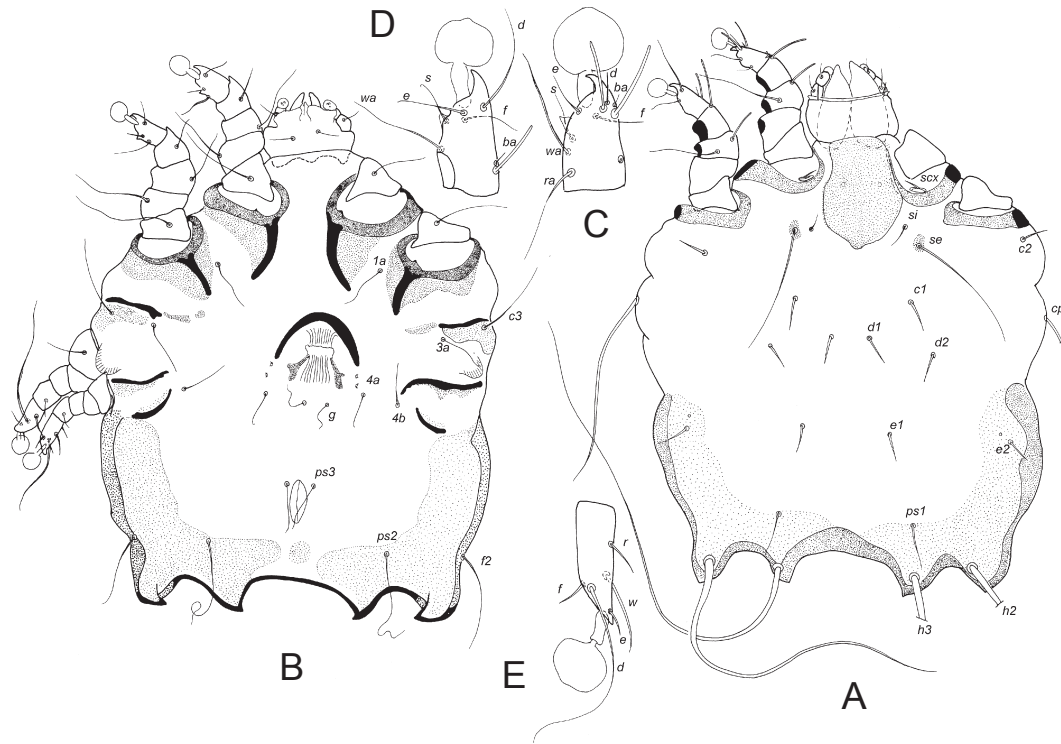


Fig. 102. *Nasalialges borneensis* Fain et Nadchatram, 1979, female: A — dorsal view, B — ventral view, C — tarsus I in dorsal view, D — tarsus II in dorsal view, E — tarsus IV in dorsal view.

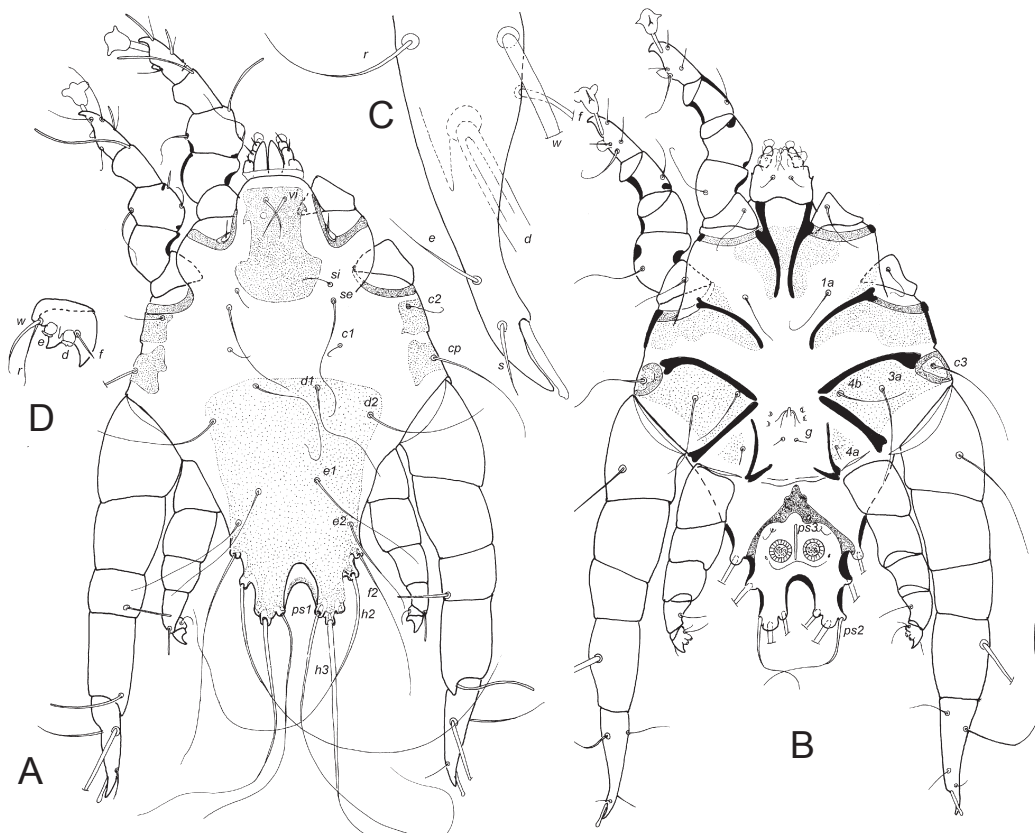


Fig. 103. *Edentalges bradypus* Fonseca, 1954, male: A — dorsal view, B — ventral view, C — tarsus III in ventral view, D — tarsus IV in dorsal view.

23. **Female:** Hysteronotal shield absent. **Male:** Paranal suckers distinctly developed, subequal to ambulacral discs of legs III. Opisthosomal lobes subdivided into 3 distinct parts each. Legs III more than 2 times longer than legs IV *Psoralges* Trouessart, 1896

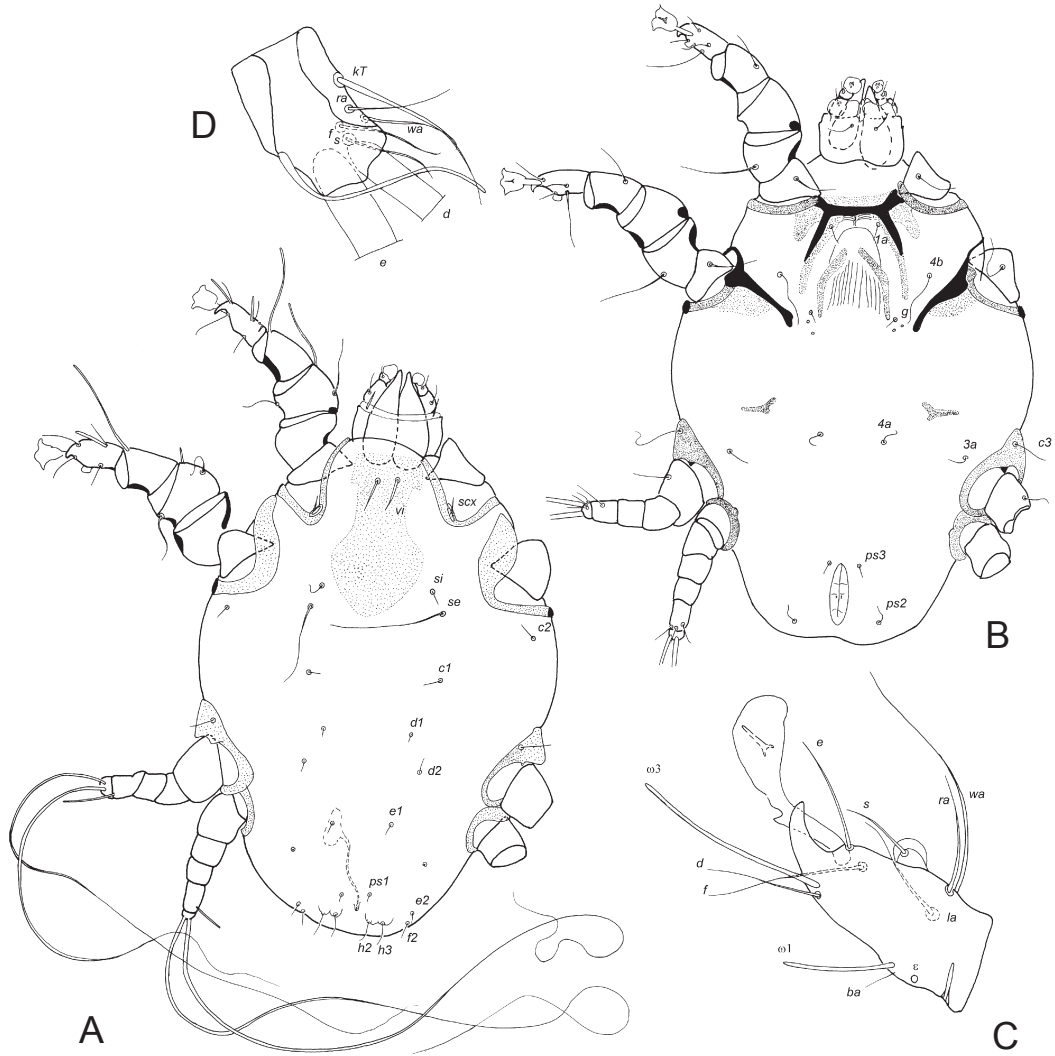


Fig. 104. *Edentalges bradypus* Fonseca, 1954, female: A — dorsal view, B — ventral view, C — tarsus III in dorsal view, D — tarsus IV in dorsal view.

— *Female*: Hysteronotal shield present. *Male*: Paranal suckers weakly developed, smaller than ambulacral discs of legs III. Opisthosomal lobes not subdivided into parts. Legs III less than 2 times longer than legs IV.
 *Myoproctalgas* Fain et Lukoschus, 1974 24. *Both sexes*: Body 1.3–2 times longer than wide; setae *d2* and *e2* present. *Male*: Paranal suckers present; legs III distinctly wider and longer than legs IV. 25
 — *Both sexes* (females are unknown but tritonymph present): Body 3 times longer than wide; setae *d2* and *e2* absent. *Male*: Paranal suckers absent. Legs III and IV subequal *Galagalges* Fain, 1963 25. *Female* (and tritonymph): Setae *ps2* anterior to level of *ps3* bases. *Male*: Latero-dorsal sclerites of hysteronotal shield present. Opisthosomal lobes distinct. Pretarsi III present. 26

— *Tritonymph* (female unknown): Setae *ps2* posterior to level of *ps3* bases. *Male*: Latero-dorsal sclerites of hysteronotal shield absent. Opisthosomal lobes weakly developed. Pretarsi III absent. *Cheirogalalgas* Fain, 1963 26. *Female*: Hysteronotal shield absent. Setae *1a* and *4a* moderately developed. *Male*: Hysteronotal shield with distinct ornamentation. Adanal shields fused into single arch-like shield. Adanal membrane distinct, bearing protuberances. Opisthosomal lobes widely separated. Setae *sIII* modified, not filiform. 27
 — *Female*: Hysteronotal shield present. Setae *1a* and *4a* whip-like. *Male*: Hysteronotal shield without ornamentation. Adanal shields separated. Adanal membrane indistinct or absent, without protuberances. Opisthosomal lobes adjacent but not fused. Setae *sIII* filiform *Makialges* Gaud et Till, 1957

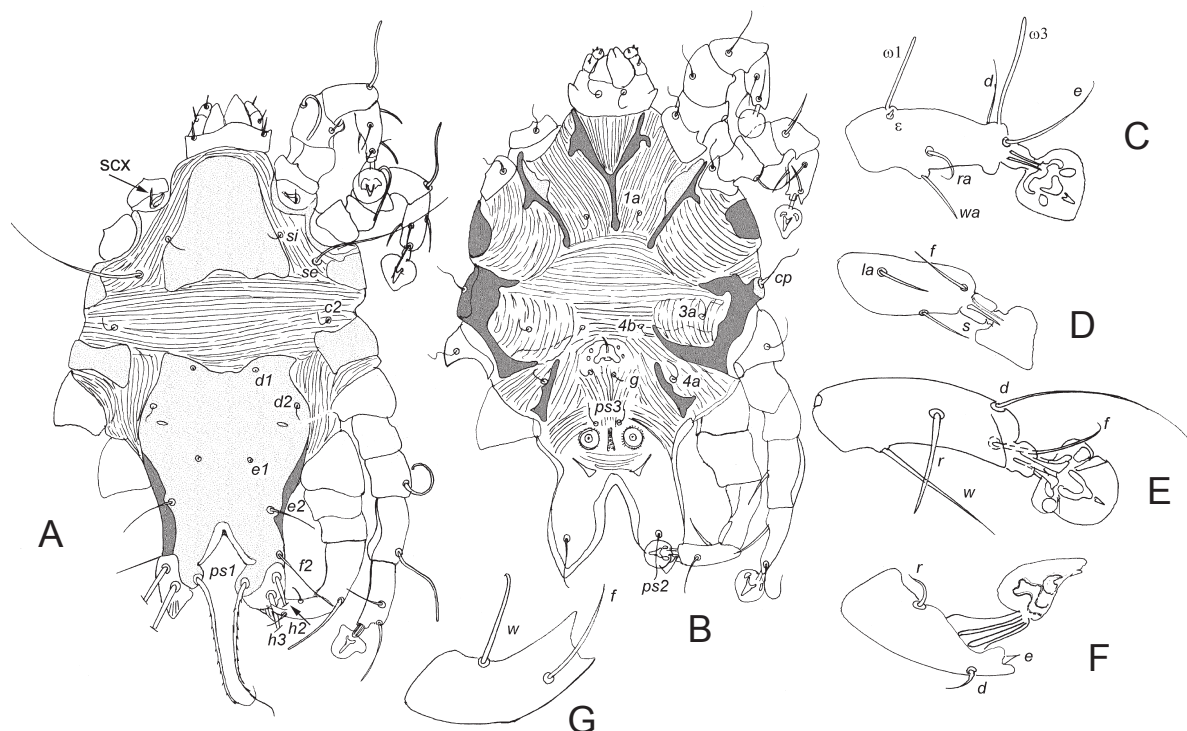


Fig. 105. *Lobalges trouessarti* Fonseca, 1954, male: A — dorsal view, B — ventral view, C — tarsus I in dorsal view, D — same in ventral view, E — tarsus III in dorsal view, F — tarsus IV in dorsal view, G — same in ventral view.

27. *Both sexes*: Coxal fields I–II without spurs. *Male*: tarsi IV without dorso-basal projection ... 28 — *Both sexes*: Coxal fields I–II with spurs. *Male*: tarsi IV with dorso-basal projection *Gaudalges* Fain, 1963
28. *Female*: Epigynum moderately developed, located between coxal fields I. Tarsi III and IV without ventral projections. *Male*: median area of hysteronotal shield without ornamentation. Tibia III with dorso-apical projection *Lemuralges* Fain, 1963
- *Female*: Epigynum large, located between coxal fields II. Tarsi III and IV with ventral projections. *Male*: Hysteronotal shield entirely ornamented. Tibia III without dorso-apical projection. *Daubentonialges* Fain, 1972

Family Lobalgidae Fain, 1965

Type genus: *Lobalges* Fonseca, 1954.
Table 18, Figs. 105–110

Diagnosis. *Both sexes*. Idiosoma dorso-ventrally flattened. Supracoxal sclerite and supracoxal opening distinct, setae *scx* present. Propodonal and hysteronotal shields present, distinctly developed. Setae *si* situated distinctly anterior to level of seta *se* bases. Apodemes Ia fused into sternum. Legs I–IV without clasping organs. Ambulacral sclerites discernible. Dorso-apical projection strongly reduced. Setae *sI* and II filiform.

Female. In Lobalginae, hysteronotal shield not subdivided. Echimytricalginae, hysteronotal shield subdivided onto unpaired central hysteronotal shield and paired opisthonorotal shields. Opisthosomal lobes present. Ovipore Y-shaped. Epigynum distinctly developed. Oviparous.

Male. In Lobalginae, legs III and IV normally developed; in Echimytricalginae, legs IV strongly reduced and their femora and genua are completely fused. Setae *dIV* and *eIV* filiform (Lobalginae) or sucker-like (Echimytricalginae).

Taxa included: 2 genera and 7 species, Lobalginae Fain, 1965 — *Lobalges* Fonseca, 1954 (1 species); Echimytricalginae Fain, 1970 — *Echimytricalges* Fain, 1970 (6 species).

Associations with hosts: inhabitants of the fur of placental mammals, Lobalginae — Pilosa (Bradypodidae, Megalonychidae) and Echimytricalginae — Rodentia (Echimyidae).

Mites of the family Lobalgidae parasitize hosts inhabiting the Neotropical region but belonging to distantly related orders, Pilosa (sloths of the families Bradypodidae and Megalonychidae) and Rodentia (spiny rats of the family Echimyidae belonging to both tree and ground-dwelling genera). The strong morphological distinctions between the lobalgid subfamilies suggests a long time since their divergence. On the other hand, their associations with such phyloge-

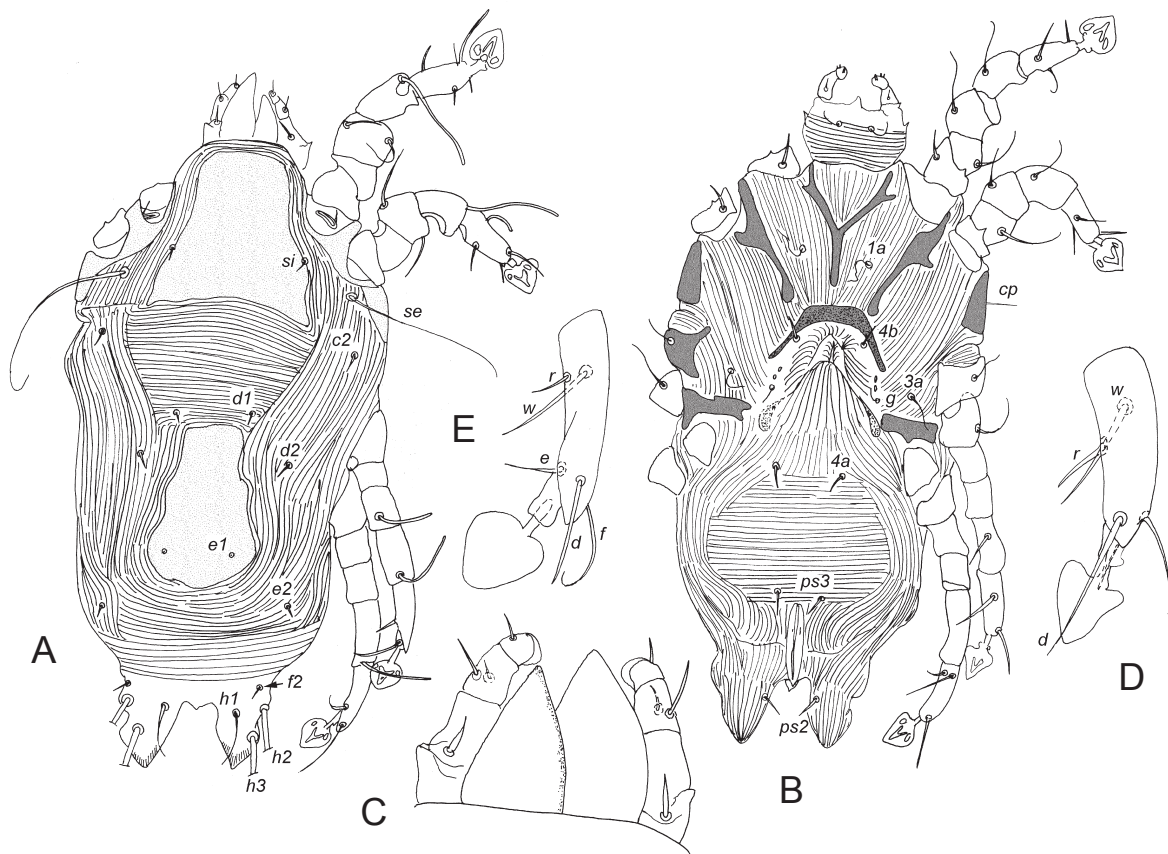


Fig. 106. *Lobalges trouessarti* Fonseca, 1954, female: A — dorsal view, B — ventral view, C — gnathosoma in dorsal view, D — tarsus III in dorsal view, E — tarsus IV in ventral view.

netically distant hosts strongly suggests secondary switching from one host group to the another (Bochkov and OConnor 2010).

Distribution. South America.

Main references. Fain (1965b) — diagnosis of Lobalgidae, Fain (1970c), Fain and Lukoschus (1970), Fain et al. (1982), Fain and Ritzi (2001) — description of new lobalgids, Bochkov and OConnor (2010) — review of Lobalgidae.

Key to genera of the family Lobalgidae
Fain, 1965

1. *Both sexes*: Subcapitulum dorsally without lobes. Setae *cp* filiform. Setae *c1* present, *c3* absent. Setae *h3* whip-like. Posterior ends of apodemes I fused into sternum. Solenidion σ III present. Setae *e*III, *s*III, *k*TIV absent. *Females*: Hysteronotum bearing one hysteronotal shield. Apodemes III short, not fused with arch of epigynum *Males*: Legs III and IV subequal, well developed; femur and genu IV not fused. Setae *d*IV and *e*IV filiform

Lobalginae Fain, 1965 (*Lobalges* Fonseca, 1954)
— *Both sexes*: Subcapitulum dorsally with pair of large lobes. Setae *cp* sword-like. Setae *c1* absent, *c3* present. Setae *h3* with foliate membrane. Poste-

rior ends of apodemes I ends free. Solenidion σ III absent. Setae *e*III, *s*III, and *k*TIV present. *Female*: Hysteronotum bearing 3 shields, unpaired central hysteronotal shield, and pair of opisthotal shields. Apodemes III distinctly developed, fused with arch of epigynum. *Male*: Legs IV strongly reduced, distinctly shorter than legs III, with fused femur and genu. Setae *d*IV and *e*IV sucker-like ...
..... Echimytricalginae Fain, 1970 (*Echimytricalges* Fain, 1970)

Subfamily incertae sedis within psoroptid complex

Subfamily Paracoroptinae Lavoipierre, 1955

Type genus: *Paracoroptes* Lavoipierre, 1955.
Table 19, Figs. 111, 112

Diagnosis. *Both sexes*. Idiosoma dorso-ventrally flattened. Supracoxal sclerite and supracoxal opening distinct, setae *scx* absent. Propodonotal and hysteronotal shields present. Bases of setae *si* and *se* situated close to each other. Apodemes Ia widely separated from each other. Legs I–IV without clasping organs. Ambulacral sclerites discernible. Dorso-apical projection absent. Setae *s*I and II angle-like, setae *ba*I, II absent.

A review of mammal-associated Psoroptidia

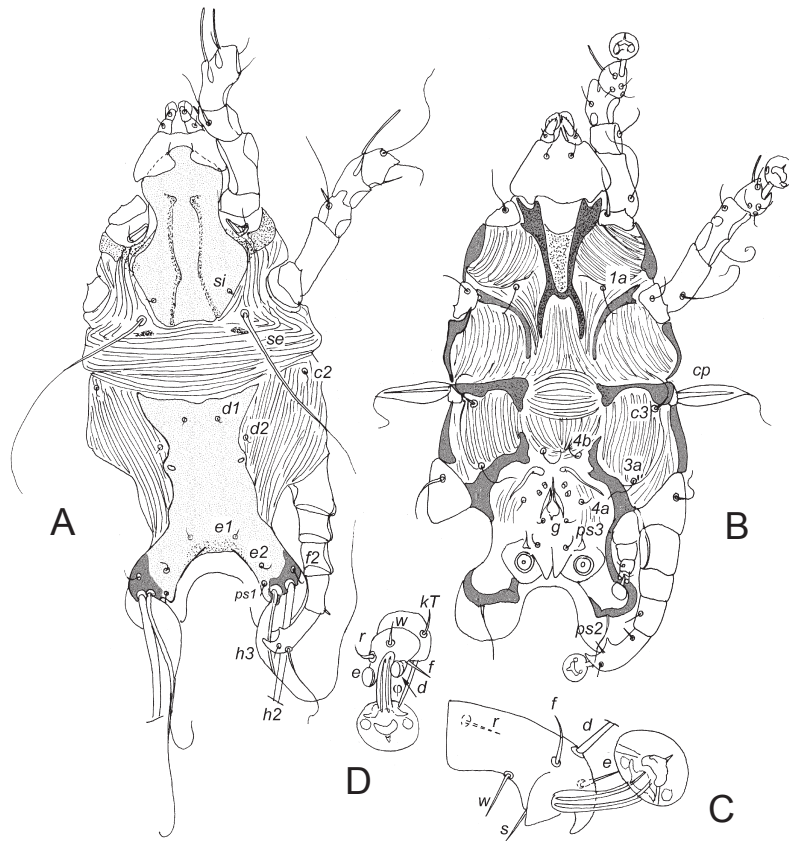


Fig. 107. *Echimytricalges guyanensis* Fain, 1970, male: A — dorsal view, B — ventral view, C — tarsus III in ventral view, D — tarsus-tibia IV in ventral view.

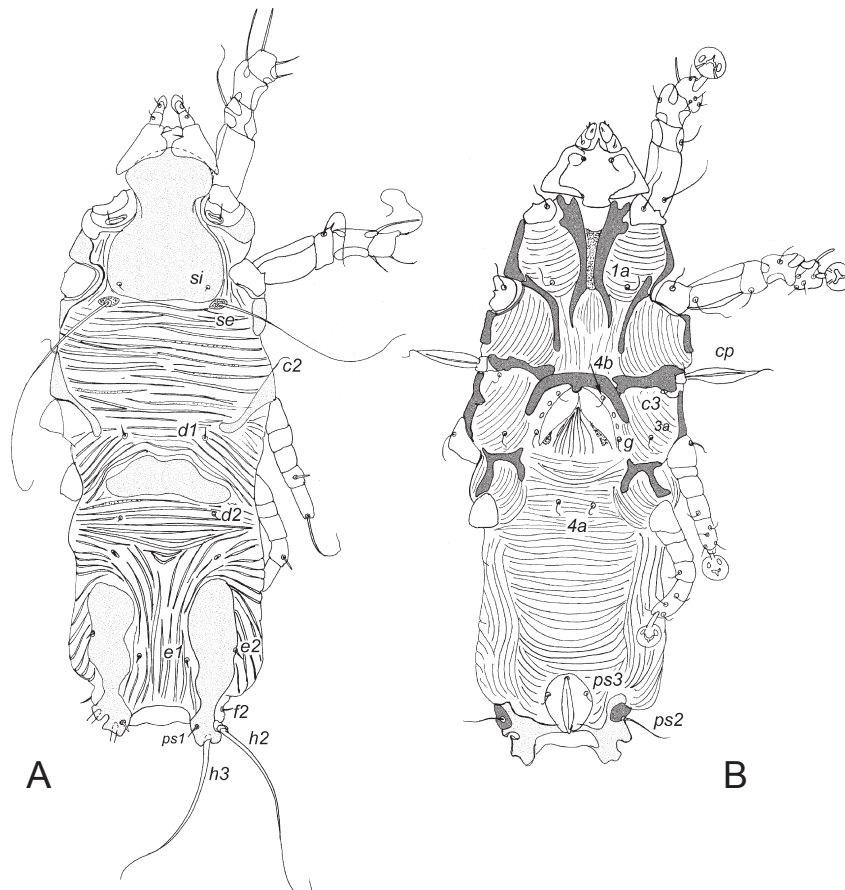


Fig. 108. *Echimytricalges guyanensis* Fain, 1970, female: A — dorsal view, B — ventral view.

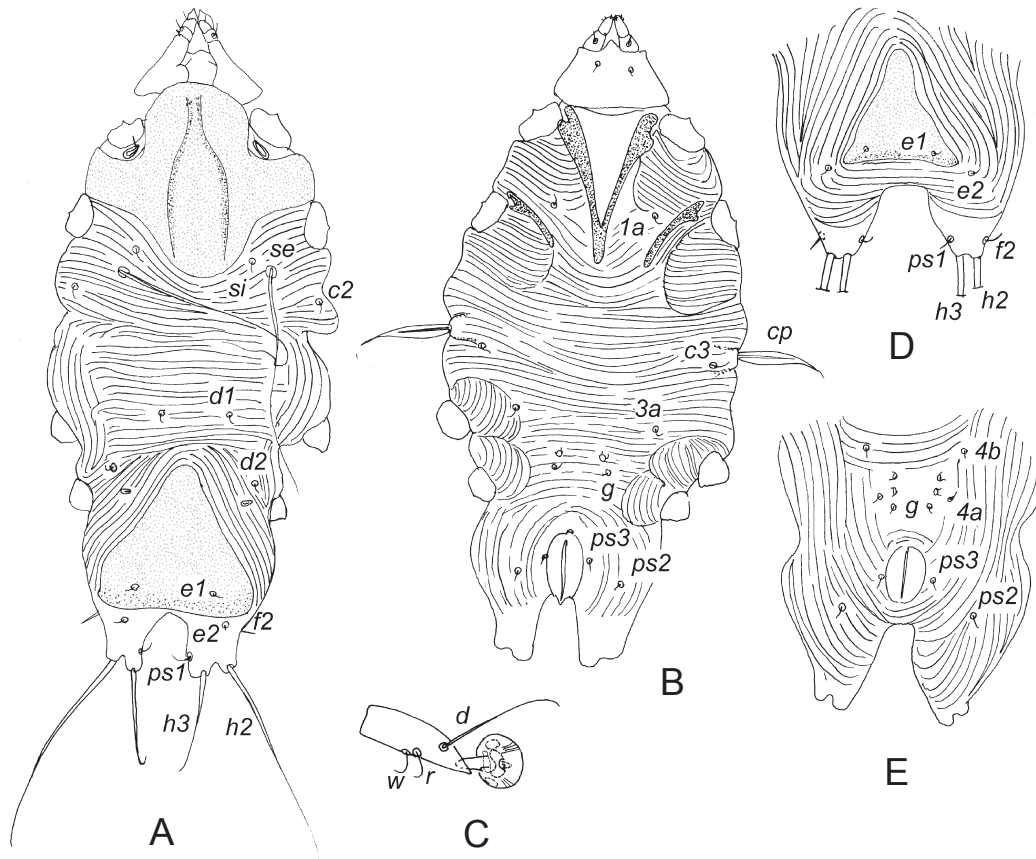


Fig. 109. *Echimytricalges hoplomys* Fain et al, 1982, larva: A — dorsal view, B — ventral view.

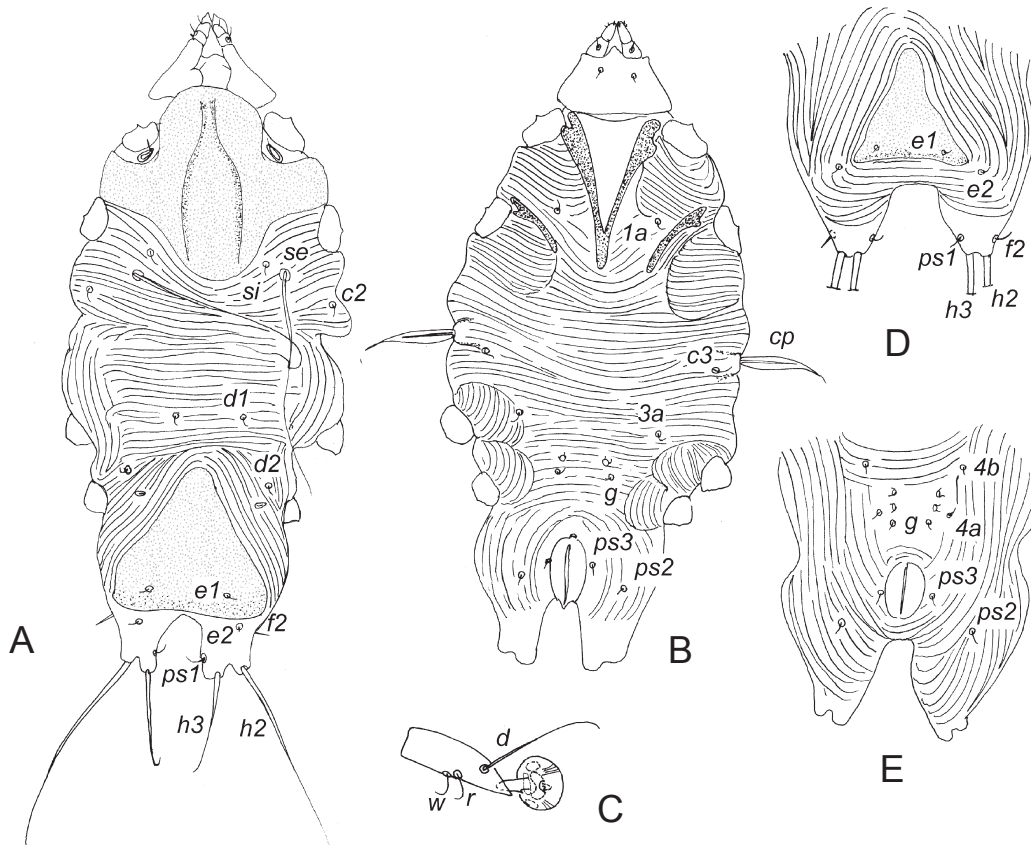


Fig. 110. *Echimytricalges hoplomys* Fain et al., 1982, protonymph (A-C): A — dorsal view, B — ventral view, C — tarsus I in dorsal view; tritonymph (D, E): D — opisthosoma in dorsal view, E — same in ventral view.

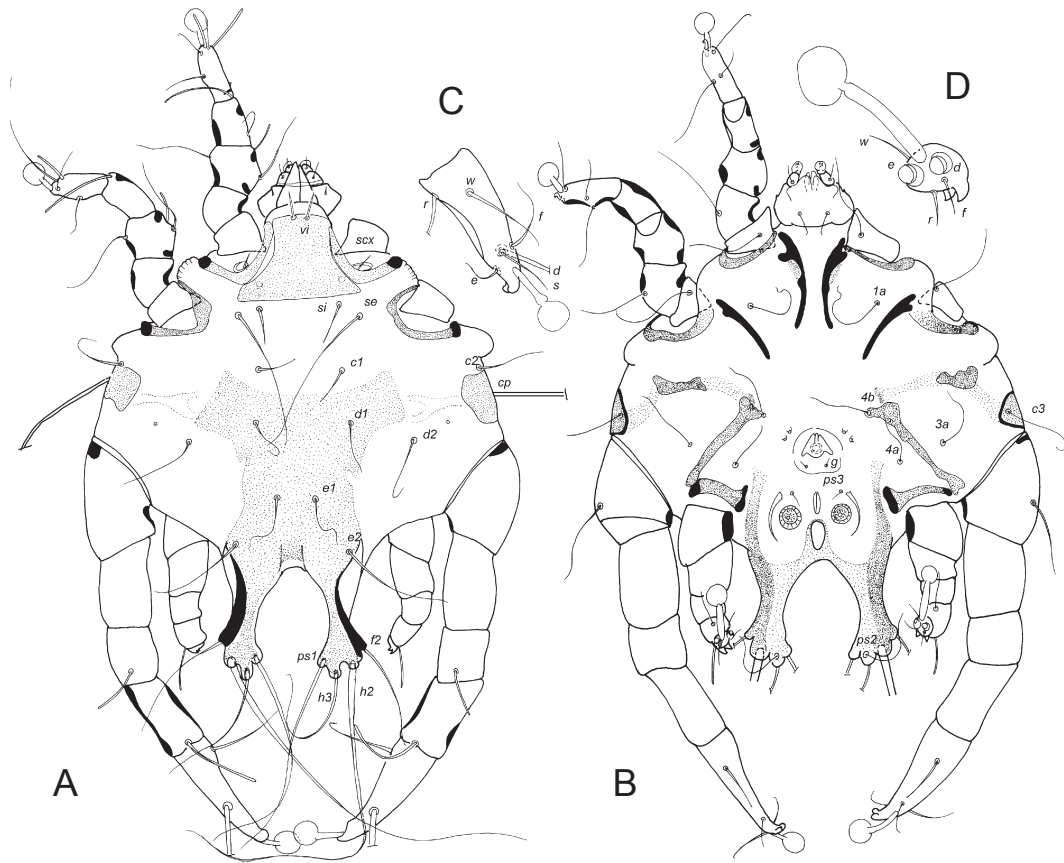


Fig. 111. *Paracoroptes allenopithecii* Fain, 1963, male: A — dorsal view, B — ventral view, C — tarsus III in ventral view, D — tarsus IV in dorsal view.

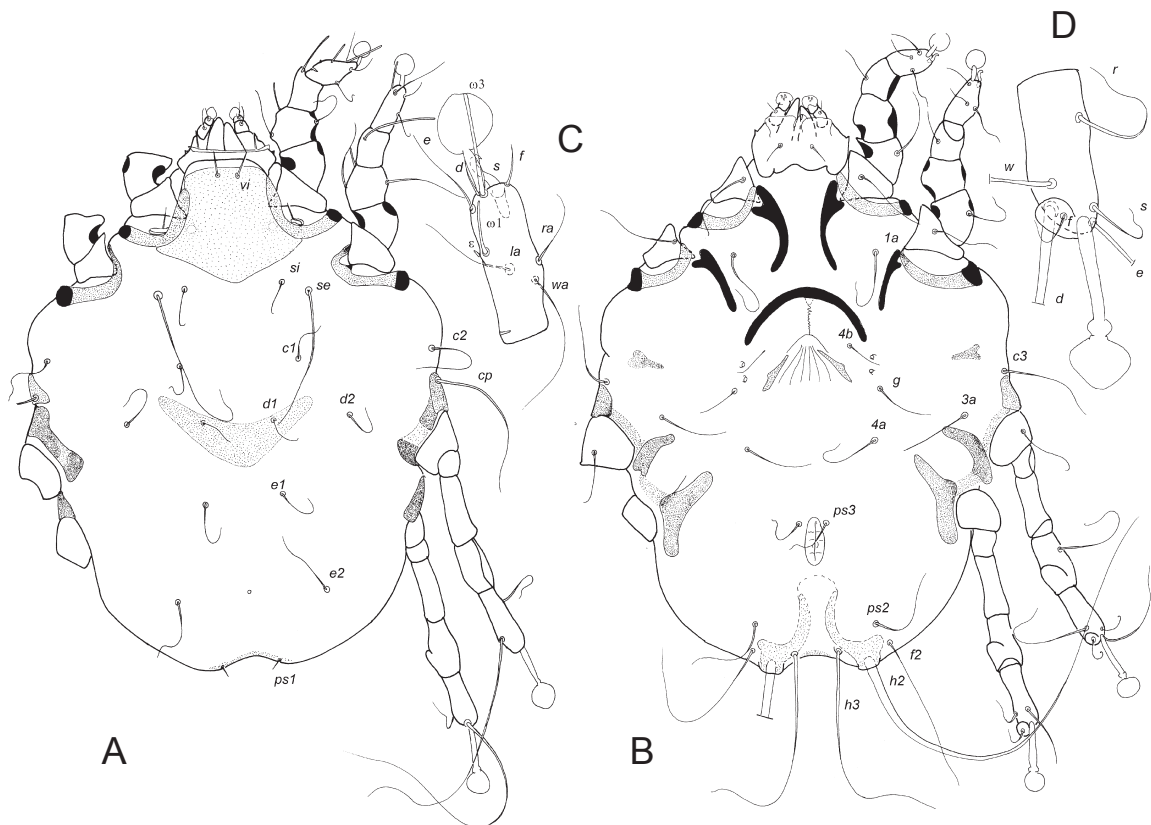


Fig. 112. *Paracoroptes allenopithecii* Fain, 1963, female: A — dorsal view, B — ventral view, C — tarsus I in dorsal view, D — tarsus III in ventral view.

Female. Opisthosomal lobes absent. Ovipore Y-shaped. Epigynum distinctly developed. Oviparous.

Male. Legs III about 2 times longer than legs IV. Setae *d*IV and *e*IV sucker-like.

Taxa included: 2 genera and 6 species, *Pangorillalges* Fain, 1962 (2 species), *Paracoroptes* Lavoipierre, 1955 (4 species).

Associations with hosts: inhabitants of the skin of Primates (Cercopithecidae, Hominidae).

Distribution. Africa.

Main references. Fain (1963) — revision of Paracoroptinae, Fain and Segerman (1978) — description of *Paracoroptes natalensis*.

Key to genera of the subfamily Paracoroptinae Lavoipierre, 1955 (both sexes)

1. Posterior margin of subcapitulum without ventral projections *Paracoroptes* Lavoipierre, 1955 — Subcapitulum dorsally with pair of distinct projections *Pangorillalges* Fain, 1962

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Table 1. Summary of taxa number, distribution and host associations of Sarcoptoidea

Family (subfamily)	Species <i>n</i>	Genera <i>n</i>	Host orders		Microhabitats	Regions
			Marsupialia	Placentalia		
Psoroptid complex						
Psoroptidae Canestrini, 1892						
Psoroptinae	21	10		Artiodactyla, Carnivora, Erinaceomorpha, Hyracoidea, Lagomorpha, Perissodactyla, Rodentia	Skin (including ear auricles)	Cosmopolite
Cebalginae Fain, 1962	7	6		Primates (Platyrrhini)	Skin	South America
Makiatginae Gaud et Till, 1957	11	6		Primates (Strepsirrhini)	Skin	Africa (including Madagascar)
Psoralginae Oudemans, 1908	9	5	Diprotodontia (Vombatidae)	Pilosa, Rodentia	Skin	Australia, South America
Nasaliatinae Fain et Nachatram, 1979	1	1		Primates (Cercopithecoidea)	Ear auricles	Asia
Marsupialginae Fain, 1963	1	1	Didelphimorphia		Skin	South America
Lobaligidae Fain, 1965						
Lobaliginae	1	1		Pilosa (Bradyrodidae and Megalonychidae)	Fur (undercoat)	South America
Echimytricalginae Fain, 1970	6	1		Rodentia (Echimyidae)	Fur (concave surfaces of spine-like hairs)	South America
Subfamilies <i>incertae sedis</i>						
Paracoroptinae Lavoipierre, 1955	6	2		Primates (Catarrhini)	Skin	Africa
Sarcoptid complex						
Sarcoptidae Murray, 1877						
Sarcoptinae	8	4		Primates, Rodentia	Superficial skin layers	Cosmopolite
Diaboloicoptinae Fain et Domtrow, 1974	3	2	Dasyuromorphia		Superficial skin layers	Australia
Teinocoptinae Fain, 1959	106	9		Carnivora, Chiroptera, Erinaceomorpha, Lagomorpha, Primates, Rodentia, Soricomorpha	Superficial skin layers	Cosmopolite
Rhyncoptidae Lawrence, 1956	12	5	Paucituberculata	Carnivora, Primates, Rodentia (Hystricidae)	Hair follicles	Eurasia, Africa, North and South America
Chirorhynchobidae Fain, 1967	3	1		Chiroptera (Phyllostomidae)	edges of wings	South America

Atopomelidae Gunther, 1942	420	46	Peramelimorphia, Diprotodontia, Dasyuromorphia, Didelphimorphia, Microbiotheria, Notorycteromorphia Paucituberculata	Afrosoricida (Tenrecidae), Carnivora, Macroscelidea, Primates, Rodentia, Soricomorpha	Fur	Asia, Africa (including Madagascar), Australia, South America (1 species in Europe, 1 species in North America and 1 cosmopolite species)	
Chirodiscidae Trouessart, 1892							
Chirodiscinae	1	1	? a small Australian marsupial		Fur	Australia	
Lemuroecinae Fain, 1968	1	1		Primates (Cheirogaleidae)	Fur	Madagascar	
Schizocoptinae Fain, 1970	2	1		Afrosoricida (Chrysochloridae)	Fur	Africa	
Labidocarpinae Gunther, 1942	226	24		Carnivora (Mustelidae), Chiroptera, Primates (Galagidae), Rodentia (Castoridae), Soricomorpha (Soricidae)	Fur (including vibrissae)	Cosmopolite	
Myocoptidae Gunther, 1942	60	5		Rodentia	Skin (Trichoecius — fur)	Cosmopolite (not recorded on Madagascar)	
Listrophoridae Megnin et Trouessart, 1884							
Listrophorinae	168	19	Didelphimorphia, Paucituberculata	Carnivora, Erinaceomorpha, Lagomorpha, Macroscelidea, Rodentia, Scandentia, Soricomorpha	Fur	Eurasia, Africa, North and South America	
Atopodontochirinae Fain et Hyland, 1972	1	1		Rodentia	Fur	North America	
Gastronyssidae Fain, 1956							
Gastronyssinae	29	7		Chiroptera	Nasal passages, eye orbits, stomach and duodenum	Cosmopolite	
Yunkeracarinae Fain, 1964	13	2		Rodentia	Nasal passages	Eurasia, Africa (including Madagascar), North and South America	
Pneumocoptidae Baker et al., 1958	4	1		Rodentia	Lungs	Europe, North America	
Lemurynyssidae Fain, 1957	4	1		Primates	Nasal passages	Africa, South America	
Sufamilies incertae sedis							
Dromiciocoptinae Fain, 1970	3	1	Didelphimorphia, Paucituberculata		Skin	South America	
Listropsoralginae Fain, 1965	7	3	Didelphimorphia, Phalangeriformes	Rodentia (Echimyidae)	Skin	Australia, South America	

Table 2. Idiosomal setation of Sarcoptoidea (maximum set)

Family/Subfamily	scx	vi	si	se	cl	c2	cp	c3	d1	d2	e1	e2	f2	h1	h2	h3	g	1a	3a	4a	4b	ps1	ps2	ps3
Psoroptid complex																								
Psoroptidae																								
Cebalginæ	+	+	+	+	-	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Makialginæ	-	+	+	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Psoralginæ	+	+	+	+	-	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Psoroptinæ	+	-	+	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Nasallialginæ	+	-	+	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	-/+	+	+	+
Marsupialginæ	-	-	+	+	-	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Lobalginæ																								
Echimytricalginæ	+	-	+	+	-	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Lobalginæ	+	-	+	+	+	+	+	-	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Subfamilies <i>incertae sedis</i>																								
Paracoroptinæ	-	+	+	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Sarcoptid complex																								
Sarcoptidae																								
Diabolicoptinæ	-	+	-	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-
Sarcoptinæ	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-
Teinocoptinæ	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-
Rhyncoptidae	-	-	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Chirohynchobiidae	-	-	-	+	+	-	-	+	-	-	-	-	-	-	-	-	+	+	+	+	+	-	+	-
Atopomelidae	+	-	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Chirodiscidae																								
Lemuroeciinæ	+	-	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Schizocoptinæ	+	-	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Labidocarpinæ	+	-	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Mycoptidae																								
Mycoptinæ	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Listrophoridae																								
Atopodontochirinae	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Listrophorinae	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+

Family/Subfamily	scx	vi	si	se	cl	c2	cp	c3	d1	d2	e1	e2	f2	h1	h2	h3	g	1a	3a	4a	4b	ps1	ps2	ps3
Gastronyssidae																								
Gastronyssinae	-	-	+	+	-	+	-	+	-	-	-	-	+	-	+	+	+	+	+	+	+	-	+	+
Yunkeracarinae	-	-	+	+	-	+	+	+	-	-	-	-	+	+	+	+	+	+	+	+	+	-	-	+
Pneumocoptidae																								
Pneumocoptinae	-	-	+	+	-	+	+	+	-	-	-	-	-	-	+	+	-	+	+	+	+	-	-	+
Lenurnyssidae																								
Lenurnyssinae	-	-	+	+	-	+	+	+	-	-	-	-	-	+	+	-	+	+	+	+	+	-	-	+
Subfamilies <i>incertae sedis</i>																								
Dromiciocoptinae	+	-	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Listropsoralginae	-	+	+	+	-	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+

- absent; + present; +/- female/male

Table 3. Leg I–II setation of Sarcoptoidea (maximum set)

Family/Subfamily	d	ba	e	f	s	la	ω1	ω3	σ1	ε	φ	wa	ra	gT	mG	cG	vF	pR
Psoroptid complex																		
Psoroptidae																		
Cebalginae	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Makialginae	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Psoralginae	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Psoroptinae	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Nasalialginae	+	+	+	+	+	-	+	+	+	+	+	+	+/-	+	+	+	+	+
Marsupialginae	+	-	+	+	+/-	+/-	+	+	+	+	+	+	+/-	+	+	+	+	+
Lobalidae																		
Echimytricalginae	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Lobalginae	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Subfamilies <i>incertae sedis</i>																		
Paracoptinae	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
Sarcoptid complex																		
Sarcoptidae																		
Diablicoptinae	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+
Sarcoptinae	-/+	+	+	+	+	+	+	+	+/-	-	+	+	+	+	+	+	+	+
Teinocoptinae	-	+	+	+	+	-	+	+	+/-	-	+	+	+	+	+	+	+	+
Rhyncoptidae	+	+	+	+	-	-	+	+	-	+	+	+	+	+	+	+	+	+

Family/Subfamily	<i>d</i>	<i>ba</i>	<i>e</i>	<i>f</i>	<i>s</i>	<i>la</i>	$\omega 1$	$\omega 3$	$\sigma 1$	ε	ϕ	<i>wa</i>	<i>ra</i>	<i>gT</i>	<i>mG</i>	<i>cG</i>	<i>vF</i>	<i>pR</i>
Chirorhynchobiidae	-	+	+	+	+	-	+	+	-	-	+	-	-	+	+	+	-	+
Atopomelidae	+	+	+	+	+	+/-	+	+	+	+	+	+	+	+	+	+	+	+
Chirodiscidae																		
Lemuroeciinae	?/+	?/-	?/+	?/+	?/-	-	+	+	-	-	+	?/+	?/+	+	+	+	+	-/+
Schizocoptinae	?/+	?/-	?/+	?/+	?/-	+/-	+	+	-	-	+	?/+	?/+	+	+	+	+	-
Labidocarpinae	-	-	-	-	-	-	+	+	-	-	+	-	-	+	+	+	+	-
Myocoptidae																		
Myocoptinae	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+
Listrophoridae																		
Aplodontochirinae	+	-	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+/-
Listrophorinae	+	-	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+/-
Gastronyssidae																		
Gastronyssinae	+	-	+	+	+	+	+	-F+M	-	+	+	+	+	+	+	+	+	-
Yunkeracarinae	+	+	+	+	+	+	+	-	-	+	+	+	+	+	+	+	+	+
Pneumocoptidae	+	+	+	+	+	-	+	-	-	-	+	+	+	+	+	+	+	-
Lemurnyssidae	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	-
Subfamilies <i>incertae sedis</i>																		
Dromiciocoptinae	+	-	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+
Listropsoralginae	+	-	+	+	+	-	+	+	+	-	+	-	+	+	+	+	+	+

+ present; - absent; F/M — female/male; / — leg I/leg II

Table 4. Leg III–IV setation of Sarcoptoidea (maximum set)

Family/Subfamily	<i>dIII</i>	<i>dIV</i>	<i>eIII</i>	<i>eIV</i>	<i>fIII</i>	<i>fIV</i>	<i>sIII</i>	<i>sIV</i>	<i>sRIII</i>	ϕIII	ϕIV	σIII	<i>wIII</i>	<i>wIV</i>	<i>rIII</i>	<i>rIV</i>	<i>kTIII</i>	<i>kTIV</i>
Psoroptid complex																		
Psoroptidae																		
Cebalginae	+	+	+	+	+	+	+	-	+	+	+	-	+	+	+	+	+	+
Makialginae	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Psoralginae	+	+	+	+	+	+	+	-	+	+	+	-	+	+	+	+	+	+
Psoroptinae	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Nasaliaalinae	+	+	+	+	+	+	-	-	+	+	+	-	+	+	+	+	+	+
Marsupialginae	+	+	+	+	+	+	-	-	+	+	+	-	+	+	+	+	+	+

Family/Subfamily	dIII	dIV	eIII	eIV	fIII	fIV	sIII	sIV	sRIII	φIII	φIV	σIII	wIII	wIV	rIII	rIV	kTIII	kTIV
Lobalgiidae																		
Echimytricalginae	+	+	+	+	+	+	+	-	+	+	+	-	+	+	+	+	+	+
Lobalginae	+	+	-	+	+	+	-	-	+	+	+	+	+	+	+	+	+	-
Subfamilies <i>incertae sedis</i>																		
Paracoroptinae	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Sarcoptid complex																		
Sarcoptidae																		
Diabolicoptinae	+	+/-	+	+/-	+	+	+	-	-	+	+	-	+	+	+	+	+	+
Sarcoptinae	+	+	+	+	+	+	+	-	-	+	+	-	+	+	+	+	+	+
Teinocoptinae	+	+/-	+/-	-	+	+	+	-	-	+	+	-	+	+	+	+	+	+
Rhyncoptidae	+	+	-	-	+	+	-	-	+	+	+	-	+	+	+	+	+	+
Chiorhynchobiidae	-	-	+	-/+	+	+	+	-	+	-	-/+	-	+	+	+	+	+	+
Atopomelidae	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Chirodiscidae																		
Lemuroeciinae	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Schizocoptinae	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Labidocarpinae	+	+	+	-	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Mycoptidae																		
Mycoptinae	+	+	+	+	+	+	-	-	+	+	+	+	+	+	+	+	+	+
Listrophoridae																		
Aplodontochirinae	+	+	+	+	+	+	-	-	+	+	+	+	+	+	+	+	+	+
Listrophorinae	+	+	+	+	+	+	-	-	+	+	+	-	+	+	+	+	+	+
Gastronyssidae																		
Gastronyssinae	+	+	+	+	+	+	+	+	-	+	+	-	+	+	+	+	+	+
Yunkeracarinae	+	+	+	+	+	+	+	+	-	+	+	-	+	+	+	+	+	+
Pneumocoptidae	+	+	+	+	+	+	+	-	-	+	+	-	+	+	+	+	+	-
Lemurynyssidae	+	+	+	+	+	+	+	+	-	+	+	-	+	+	+	+	+	+
Subfamilies <i>incertae sedis</i>																		
Dromiciocoptinae	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	+	+	+
Listropsoralginae	+	+	+	+/-	+	+	-	-	+	+	+	+	+	+	+	+	+	+

+ present; - absent; / — female/male

Host distribution of the mammal-associated Psoroptidia (Tables 5–19)

In all tables, type species of each genus and subgenus is given first in the list.

Geographical references: The European Region does not include Turkey, which is considered here under the Asiatic Region. In the Australian Region, Oceania does not include Eastern Timor and the Indonesian islands, which are considered under the Asiatic Region; New Guinea and New Zealand are specially mentioned in the parenthesis. In the African Region, Madagascar is specially mentioned in the parentheses. The South American Region includes Central America and islands of the Caribbean Sea.

* — host shift; ! — accidental host; ? — doubtful mite determination; [] — problem with host nomenclature or this host name is absent in Wilson and Reeder (2005)

Table 5. Host distribution of the family Sarcoptidae Murray, 1877

& — this species is known from representatives of more than 16 families of nine orders of therian mammals

Mite species	Host species	Host family	Host order	Distribution
Sarcoptinae Murray, 1877				
<i>Sarcoptes</i> Latreille, 1802				
<i>Sarcoptes scabiei</i> (Linnaeus, 1758)*	<i>Homo sapiens</i>	Hominidae	Primates	Cosmopolite
<i>Prosarcoptes</i> Lavoipierre, 1960				
<i>Prosarcoptes pitheci</i> (Philippe, 1948)	<i>Papio hamadryas</i>	Cercopithecidae	Primates	Africa
“	<i>Chlorocebus aethiops</i>	Cercopithecidae	Primates	Africa
<i>Prosarcoptes scaloni</i> Smiley, 1965	<i>Macaca fascicularis</i>	Cercopithecidae	Primates	Asia
“	<i>Macaca arctoides</i>	Cercopithecidae	Primates	USA
<i>Prosarcoptes talapoini</i> (Fain, 1965)	<i>Miopithecus talapoin</i>	Cercopithecidae	Primates	Africa
<i>Kutzerocoptes</i> Lavoipierre, 1970				
<i>Kutzerocoptes grunbergi</i> Lavoipierre, 1970	<i>Cebus capucinus</i>	Cebidae	Primates	South America
<i>Trixacarus</i> Sellnick, 1944				
<i>Trixacarus diversus</i> Sellnick, 1944	<i>Rattus norvegicus</i>	Muridae	Rodentia	Europe
“	<i>Rattus rattus</i>	Muridae	Rodentia	Europe
“	<i>Calomys musculinus</i>	Sigmodontidae	Rodentia	South America
<i>Trixacarus eliurus</i> Klompen, 1992	<i>Eliurus webbi</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Trixacarus caviae</i> Fain, Howell et Hyatt, 1972	<i>Cavia porcellus</i>	Caviidae	Rodentia	Cosmopolite
Teinocoptinae Fain, 1959				
<i>Teinocoptes</i> Rodhain, 1923				
<i>Teinocoptes epomophori</i> Rodhain, 1923	<i>Epomophorus wahlbergi</i>	Pteropodidae	Chiroptera	Africa
“	<i>Epomophorus labiatus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Epomophorus gambianus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Epomops franqueti</i>	Pteropodidae	Chiroptera	Africa
“	<i>Micropteropus pusillus</i>	Pteropodidae	Chiroptera	Africa
<i>Teinocoptes vandeuseni</i> Mitchell et Fain, 1963	<i>Rousettus amplexicaudatus</i>	Pteropodidae	Chiroptera	Asia
<i>Teinocoptes malayi</i> Fain et Nadchatram, 1962	<i>Macroglossus</i> sp.	Pteropodidae	Chiroptera	Asia
“	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
“	<i>Eonycteris robusta</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus celebensis</i>	Pteropodidae	Chiroptera	Asia

A review of mammal-associated Psoroptidia

<i>Teinocoptes pahangensis</i> Fain, Lukoschus et Nadchatram, 1982	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus amplexicaudatus</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
“	<i>Rousettus madagascariensis</i>	Pteropodidae	Chiroptera	Africa (Madagascar)
<i>Teinocoptes asiaticus</i> Fain et Domrow, 1961	<i>Cynopterus brachyotis</i>	Pteropodidae	Chiroptera	Asia
“	<i>Ptenochirus jagori</i>	Pteropodidae	Chiroptera	Asia
“	<i>Macroglossus</i> sp.	Pteropodidae	Chiroptera	Asia
<i>Teinocoptes astridae</i> Fain, 1959	<i>Rousettus</i> sp.	Pteropodidae	Chiroptera	Africa (Madagascar)
	<i>Rousettus madagascariensis</i>	Pteropodidae	Chiroptera	Africa (Madagascar)
<i>Teinocoptes rousetti</i> Fain, 1959	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Africa
<i>Teinocoptes strandmanni</i> Mitchell et Fain, 1963	<i>Dobsonia moluccensis</i>	Pteropodidae	Chiroptera	Asia
<i>Teinocoptes katherinae</i> Mitchell et Fain, 1963	<i>Dobsonia inermis</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
<i>Teinocoptes wilsoni</i> Mitchell et Fain, 1963	<i>Dobsonia moluccensis</i>	Pteropodidae	Chiroptera	Asia
<i>Teinocoptes aingworthi</i> Mitchell et Fain, 1963	<i>Nyctimene</i> sp.	Pteropodidae	Chiroptera	Asia
<i>Teinocoptes philippinensis</i> Klompen, 1992	<i>Harpyionycteris whiteheadi</i>	Pteropodidae	Chiroptera	Asia
<i>Teinocoptes eonycteris</i> Fain, Lukoschus et Nadchatram, 1982	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus amplexicaudatus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Africa
<i>Teinocoptes johnstoni</i> Fain, 1963	<i>Thoopterus nigrescens</i>	Pteropodidae	Chiroptera	Asia
<i>Teinocoptes haymani</i> Fain, 1963	<i>Notopteris macdonaldi</i>	Pteropodidae	Chiroptera	Australia
<i>Teinocoptes eidoloni</i> Fain, 1959	<i>Eidolon helvum</i>	Pteropodidae	Chiroptera	Africa
<i>Teinocoptes domrowi</i> Fain, 1961	<i>Pteropus conspicillatus</i>	Pteropodidae	Chiroptera	Australia
“	<i>Pteropus alecto</i>	Pteropodidae	Chiroptera	Australia
“	<i>Pteropus scapulatus</i>	Pteropodidae	Chiroptera	Australia
“	<i>Pteropus giganteus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Pteropus speciosus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Pteropus hypomelanus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Syconycteris australis</i>	Pteropodidae	Chiroptera	Australia
“	<i>Melonycteris melanops</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
“	<i>Acerodon jubatus</i>	Pteropodidae	Chiroptera	Asia
<i>Teinocoptes auricularis</i> Fain, 1959	<i>Epomophorus wahlbergi</i>	Pteropodidae	Chiroptera	Africa
“	<i>Epomophorus gambianus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Hypsignathus monstrosus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Micropteropus pusillus</i>	Pteropodidae	Chiroptera	Africa

<i>Teinocoptes ituriensis</i> Fain, 1967	<i>Lissonycteris angolensis</i>	Pteropodidae	Chiroptera	Africa
<i>Teinocoptes harpyionycteris</i> (Klompen et OConnor, 1987)	<i>Harpyionycteris whiteheadi</i>	Pteropodidae	Chiroptera	Asia
“	<i>Harpyionycteris celebensis</i>	Pteropodidae	Chiroptera	Asia
<i>Chirobia</i> Fain, 1959				
<i>Chirobia congolensis</i> Fain, 1959	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus leschenaulti</i>	Pteropodidae	Chiroptera	Asia
<i>Chirobia otophaga</i> Fain, 1959	<i>Epomophorus wahlbergi</i>	Pteropodidae	Chiroptera	Africa
“	<i>Epomophorus angolensis</i>	Pteropodidae	Chiroptera	Africa
“	<i>Epomophorus gambianus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Epomophorus labiatus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Hypsignathus monstrosus</i>	Pteropodidae	Chiroptera	Africa
<i>Chirobia thoopterus</i> Klompen, 1992	<i>Thoopterus nigrescens</i>	Pteropodidae	Chiroptera	Asia
<i>Chirobia brevior</i> (Fain, Lukoschus et Nadchatram, 1982)	<i>Chironax melanocephalus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus celebensis</i>	Pteropodidae	Chiroptera	Asia
<i>Chirobia cynopteri</i> Klompen, Lukoschus et nadchatram, 1985	<i>Macroglossus minimus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Cynopterus brachyotis</i>	Pteropodidae	Chiroptera	Asia
<i>Chirobia haplonycteris</i> Klompen, 1992	<i>Haplonycteris fischeri</i>	Pteropodidae	Chiroptera	Asia
<i>Chirobia jagori</i> Klompen, 1992	<i>Ptenochirus jagori</i>	Pteropodidae	Chiroptera	Asia
<i>Chirobia minor</i> Klompen, 1992	<i>Ptenochirus minor</i>	Pteropodidae	Chiroptera	Asia
<i>Chirobia eonycteris</i> Klompen, Lukoschus, Fain et Nadchatram, 1982	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
“	<i>Eonycteris robusta</i>	Pteropodidae	Chiroptera	Asia
<i>Chirobia rousettus</i> Klompen, 1992	<i>Rousettus amplexicaudatus</i>	Pteropodidae	Chiroptera	Asia
<i>Chirobia angolensis</i> Klompen, 1992	<i>Rousettus angolensis</i>	Pteropodidae	Chiroptera	Africa
“	<i>Myonycteris torquata</i>	Pteropodidae	Chiroptera	Africa
<i>Chirobia squamata</i> Fain, 1959	<i>Lissonycteris angolensis</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus lanosus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus madagascariensis</i>	Pteropodidae	Chiroptera	Africa (Madagascar)
<i>Cynopterocoptes</i> Klompen, 1992				
<i>Cynopterocoptes heaneyi</i> Klompen, 1992	<i>Haplonycteris fischeri</i>	Pteropodidae	Chiroptera	Asia
“	<i>Aethalops alecto</i>	Pteropodidae	Chiroptera	Asia
“	<i>Balionycteris maculata</i>	Pteropodidae	Chiroptera	Asia
“	<i>Chironax melanocephalus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Macroglossus minimus</i>	Pteropodidae	Chiroptera	Asia

A review of mammal-associated Psoroptidia

<i>Cynopteroptes heaneyi</i> Klompen, 1992	<i>Nyctimene rabori</i>	Pteropodidae	Chiroptera	Asia
“	<i>Ptenochirus jagori</i>	Pteropodidae	Chiroptera	Asia
“	<i>Ptenochirus minor</i>	Pteropodidae	Chiroptera	Asia
<i>Rousettocoptes</i> Klompen, 1992				
<i>Rousettocoptes mammophilus</i> Klompen, 1992	<i>Rousettus amplexicaudatus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus leschenaulti</i>	Pteropodidae	Chiroptera	Asia
“	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
<i>Tychosarcoptes</i> Fain, 1976				
<i>Tychosarcoptes orphanus</i> Fain, 1976	Pteropodidae sp.	Pteropodidae	Chiroptera	?
<i>Tychosarcoptes ptenochirus</i> Klompen, 1992	<i>Ptenochirus jagori</i>	Pteropodidae	Chiroptera	Asia
<i>Tychosarcoptes amamphipterinon</i> Klompen, Lukoschus, Fain et Nad- chatram, 1984	<i>Cynopterus brachyotis</i>	Pteropodidae	Chiroptera	Asia
<i>Nycteridocoptes</i> Oudemans, 1898				
<i>Nycteridocoptes poppei</i> Oudemans, 1898	<i>Vespertilio murinus</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Eptesicus serotinus</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis bechsteinii</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis myotis</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis daubentoni</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis goudoti</i>	Vespertilionidae	Chiroptera	Africa (Madagascar)
<i>Nycteridocoptes miniopteri</i> Fain, 1959	<i>Miniopterus inflatus</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Miniopterus schreibersii</i>	Vespertilionidae	Chiroptera	Europe, Africa (Madagascar)
“	<i>Miniopterus australis</i>	Vespertilionidae	Chiroptera	Asia
<i>Nycteridocoptes eyndhoveni</i> Fain, 1959	<i>Rhinolophus ferrumequinum</i>	Rhinolophidae	Chiroptera	Europe
“	<i>Rhinolophus blasii</i>	Rhinolophidae	Chiroptera	Africa
“	<i>Rhinolophus clivosus</i>	Rhinolophidae	Chiroptera	Africa
“	<i>Rhinolophus hildebrandtii</i>	Rhinolophidae	Chiroptera	Africa
“	<i>Rhinolophus fumigatus</i>	Rhinolophidae	Chiroptera	Africa
<i>Nycteridocoptes hoogstrali</i> Fain, 1961	<i>Triaenops persicus</i>	Rhinolophidae	Chiroptera	Africa
<i>Nycteridocoptes pteropi</i> Rodhain et Gedoelst, 1921	<i>Eidolon helvum</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Africa
<i>Nycteridocoptes heidemanni</i> Klompen, 1992	<i>Ptenochirus minor</i>	Pteropodidae	Chiroptera	Asia
“	<i>Ptenochirus jagori</i>	Pteropodidae	Chiroptera	Asia
“	<i>Cynopterus brachyotis</i>	Pteropodidae	Chiroptera	Asia
“	<i>Haplonycteris fischeri</i>	Pteropodidae	Chiroptera	Asia
<i>Nycteridocoptes microphallus</i> Fain, 1959	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
“	<i>Eonycteris robusta</i>	Pteropodidae	Chiroptera	Asia
“	<i>Harpyionycteris whiteheadi</i>	Pteropodidae	Chiroptera	Asia

<i>Nycteridoptes microphallus</i> Fain, 1959	<i>Rousettus celebensis</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus amplexicaudatus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Macroglossus minimus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Dobsonia exoleta</i>	Pteropodidae	Chiroptera	Asia
“	<i>Dobsonia moluccensis</i>	Pteropodidae	Chiroptera	Asia
“	<i>Pteropus pumilus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Thoopterus nigrescens</i>	Pteropodidae	Chiroptera	Asia
<i>Nycteridoptes rousetti</i> Fain, 1958	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus lanosus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousetus madagascariensis</i>	Pteropodidae	Chiroptera	Africa (Madagascar)
<i>Nycteridoptes malayi</i> Fain, 1963	<i>Aethalops alecto</i>	Pteropodidae	Chiroptera	Asia
<i>Nycteridoptes notopteris</i> Klompen, 1992	<i>Notopteris macdonaldi</i>	Pteropodidae	Chiroptera	Australia
<i>Nycteridoptes lavoipierrei</i> Fain, 1958	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Lissonycteris angolensis</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus lanosus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus madagascariensis</i>	Pteropodidae	Chiroptera	Africa (Madagascar)
<i>Nycteridoptes asiaticus</i> Fain, 1959	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
“	<i>Eonycteris robusta</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus leschenaultii</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus amplexicaudatus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Aethalops alecto</i>	Pteropodidae	Chiroptera	Asia
“	<i>Penthetor lucasi</i>	Pteropodidae	Chiroptera	Asia
<i>Nycteridoptes macrophallus</i> Fain, 1958	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus amplexicaudatus</i>	Pteropodidae	Chiroptera	Asia, Australia (New Guinea)
“	<i>Rousettus lanosus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Europe, Africa
“	<i>Rousettus madagascariensis</i>	Pteropodidae	Chiroptera	Africa (Madagascar)
“	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
“	<i>Eonycteris robusta</i>	Pteropodidae	Chiroptera	Asia
<i>Nycteridoptes orientalis</i> Fain, 1963	<i>Pteropus alecto</i>	Pteropodidae	Chiroptera	Asia
“	<i>Pteropus hypomelanus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Pteropus poliocephalus</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
<i>Nycteridoptes cynopteri</i> (Fain, 1962)	<i>Cynopterus brachyotis</i>	Pteropodidae	Chiroptera	Asia
“	<i>Ptenochirus jagori</i>	Pteropodidae	Chiroptera	Asia
“	<i>Ptenochirus minor</i>	Pteropodidae	Chiroptera	Asia

A review of mammal-associated Psoroptidia

<i>Chirophagoides</i> Fain, 1963				
<i>Chirophagoides mystacopis</i> Fain, 1963	<i>Mystacina tuberculata</i>	Mystacinidae	Chiroptera	Australia (New Zealand)
<i>Chirnyssoides</i> Fain, 1959				
<i>Chirnyssoides caparti</i> Fain, 1959	<i>Artibeus jamaicensis</i>	Phyllostomidae	Chiroptera	North and South America
“	<i>Artibeus lituratus</i>	Phyllostomidae	Chiroptera	North and South America
“	<i>Artibeus cinereus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Chiroderma salvini</i>	Phyllostomidae	Chiroptera	South America
“	<i>Desmodus rotundus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Vampyressa pusilla</i>	Phyllostomidae	Chiroptera	South America
“	<i>Platyrrhinus vittatus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Uroderma bilobatum</i>	Phyllostomidae	Chiroptera	South America
<i>Chirnyssoides amazonae</i> Fain, 1959	<i>Carollia brevicauda</i>	Phyllostomidae	Chiroptera	North and South America
“	<i>Carollia subrufa</i>	Phyllostomidae	Chiroptera	North and South America
“	<i>Carollia perspicillata</i>	Phyllostomidae	Chiroptera	South America
“	<i>Carollia castanea</i>	Phyllostomidae	Chiroptera	South America
“	<i>Glossophaga soricina</i>	Phyllostomidae	Chiroptera	North and South America
<i>Chirnyssoides vampyrops</i> Fain et Lukoschus, 1975	<i>Platyrrhinus helleri</i>	Phyllostomidae	Chiroptera	South America
“	<i>Uroderma bilobatum</i>	Phyllostomidae	Chiroptera	South America
<i>Chirnyssoides stenoderma</i> Fain et Lukoschus, 1975	<i>Stenoderma</i> sp.	Phyllostomidae	Chiroptera	South America
<i>Chirnyssoides parvisuctus</i> Fain et Lukoschus, 1975	<i>Lamproncyteris brachyotis</i>	Phyllostomidae	Chiroptera	South America
<i>Chirnyssoides phyllostomus</i> Fain et Lukoschus, 1975	<i>Phyllostomus hastatus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Lophostoma carrikeri</i>	Phyllostomidae	Chiroptera	South America
“	<i>Lophostoma silvicolum</i>	Phyllostomidae	Chiroptera	South America
<i>Chirnyssoides venezuelae</i> Fain, 1959	<i>Lophostoma brasiliense</i>	Phyllostomidae	Chiroptera	South America
“	<i>Lophostoma silvicolum</i>	Phyllostomidae	Chiroptera	South America
<i>Chirnyssoides (Noctiliocoptes)</i> Fain et Lukoschus, 1971				
<i>Chirnyssoides noctilionis</i> (Dusbabek, 1970)	<i>Noctilio leporinus</i>	Noctilionidae	Chiroptera	South America
“	<i>Noctilio albiventris</i>	Noctilionidae	Chiroptera	South America
<i>Notoedres</i> Railliet, 1893				
<i>Notoedres cati</i> (von Hering, 1838)	<i>Felis silvestris</i>	Felidae	Carnivora	Cosmopolite
“	<i>Leptailurus serval</i>	Felidae	Carnivora	Europe
“	<i>Acinonyx jubatus</i>	Felidae	Carnivora	Africa
“	<i>Lynx rufus</i>	Felidae	Carnivora	North America
“	<i>Uncia uncia</i>	Felidae	Carnivora	Asia
“	<i>Canis lupus</i>	Canidae	Carnivora	Cosmopolite
“	<i>Nasua nasua</i>	Procionidae	Carnivora	South America
<i>Notoedres cati</i> (von Hering, 1838)	<i>Herpestes javanicus</i>	Herpestidae	Carnivora	Australia (Oceania), South America

<i>Notoedres cati</i> (von Hering, 1838)	<i>Oryctolagus cuniculus</i>	Leporidae	Lagomorpha	Cosmopolite
“	* <i>Phascolarctos cinereus</i>	Phascolarctidae	Vombatiformes	Australia
<i>Notoedres musculi</i> (Kramer, 1865)	<i>Mus musculus</i>	Muridae	Rodentia	Cosmopolite
“	<i>Apodemus agrarius</i>	Muridae	Rodentia	Asia
“	<i>Apodemus flavicollis</i>	Muridae	Rodentia	Europe
<i>Notoedres oudemansi</i> Fain, 1965	<i>Rattus rattus</i>	Muridae	Rodentia	Africa
“	* <i>Atelerix albiventris</i>	Erinaceidae	Erinaceomorpha	Africa
<i>Notoedres pseudomuris</i> Lavoipierre, 1968	<i>Mus musculus</i>	Muridae	Rodentia	Asia, Africa
“	<i>Apodemus sylvaticus</i>	Muridae	Rodentia	Asia
“	<i>Microtus transcaspicus</i>	Cricetidae	Rodentia	Asia
<i>Notoedres pahangi</i> Klompen, Lukoschus et Nadchatram, 1983	<i>Rattus tiomanicus</i>	Muridae	Rodentia	Asia
“	<i>Rattus argentiventer</i>	Muridae	Rodentia	Asia
“	<i>Rattus exulans</i>	Muridae	Rodentia	Asia
“	<i>Rattus hoffmanni</i>	Muridae	Rodentia	Asia
“	<i>Niviventer fulvescens</i>	Muridae	Rodentia	Asia
“	<i>Bunomys penitus</i>	Muridae	Rodentia	Asia
“	<i>Berylmys bowersi</i>	Muridae	Rodentia	Asia
“	<i>Micaelamys namaquensis</i>	Muridae	Rodentia	Africa
“	<i>Arvicanthis niloticus</i>	Muridae	Rodentia	Africa
<i>Notoedres jamesoni</i> Lavoipierre, 1964	<i>Bandicota bengalensis</i>	Muridae	Rodentia	Asia
“	<i>Rattus annandalei</i>	Muridae	Rodentia	Asia
“	<i>Rattus argentiventer</i>	Muridae	Rodentia	Asia
“	<i>Rattus tiomanicus</i>	Muridae	Rodentia	Asia
<i>Notoedres paucipilis</i> (Lawrence, 1960)	<i>Pelomys fallax</i>	Muridae	Rodentia	Asia
<i>Notoedres muris</i> Megnin, 1877	<i>Rattus norvegicus</i>	Muridae	Rodentia	Cosmopolite
“	<i>Rattus rattus</i>	Muridae	Rodentia	Europe, Africa
“	<i>Rattus tunneyi</i>	Muridae	Rodentia	Australia
“	<i>Melomys cervinipes</i>	Muridae	Rodentia	Australia
“	<i>Arvicola amphibeus</i>	Cricetidae	Rodentia	Europe
“	<i>Cricetus cricetus</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus californicus</i>	Cricetidae	Rodentia	North America
“	<i>Holochilus brasiliensis</i>	Sigmodontidae	Rodentia	South America
“	* <i>Isoodon macrourus</i>	Perameliidae	Peramelemorpha	Australia
“	* <i>Trichosurus vulpecula</i>	Phalangeridae	Phalangeriformes	Australia
“	* <i>Erinaceus europaeus</i>	Erinaceidae	Erinaceomorpha	Australia (New Zealand)
<i>Notoedres centrifera</i> Jansen, 1963	<i>Ratufa bicolor</i>	Sciuridae	Rodentia	Asia
“	<i>Sciurus griseus</i>	Sciuridae	Rodentia	North America
“	<i>Sciurus niger</i>	Sciuridae	Rodentia	North America
“	<i>Sciurus carolinensis</i>	Sciuridae	Rodentia	North America
“	<i>Glaucomys volans</i>	Sciuridae	Rodentia	North America

A review of mammal-associated Psoroptidia

<i>Notoedres centrifera</i> Jansen, 1963	<i>Tamias striatus</i>	Sciuridae	Rodentia	North America
<i>Notoedres galagoensis</i> Fain, 1963	<i>Galago demidovii</i>	Galagidae	Primates	Africa
<i>Notoedres indicus</i> (Fain et Lukoschus, 1976)	<i>Suncus murinus</i>	Soricidae	Soricomorpha	Asia
<i>Notoedres philippinensis</i> Klompen, 1992	<i>Miniopterus tristis</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Miniopterus australis</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Miniopterus schreibersii</i>	Vespertilionidae	Chiroptera	Asia
<i>Notoedres miniopteri</i> Fain, 1959	<i>Miniopterus schreibersii</i>	Vespertilionidae	Chiroptera	Europe, Africa (Madagascar)
“	<i>Miniopterus inflatus</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Neoromicia nanus</i>	Vespertilionidae	Chiroptera	Africa
<i>Notoedres nigricans</i> Klompen, 1992	<i>Myotis nigricans</i>	Vespertilionidae	Chiroptera	South America
<i>Notoedres myoticola</i> (Fain, 1959)	<i>Myotis myotis</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis blythii</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Myotis capaccinii</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Myotis daubentonii</i>	Vespertilionidae	Chiroptera	Europe
<i>Notoedres myotis</i> (Hedeon, 1953)	<i>Myotis velifer</i>	Vespertilionidae	Chiroptera	North America
<i>Notoedres roesleri</i> Vitzthum, 1932	<i>Myotis nigricans</i>	Vespertilionidae	Chiroptera	South America
<i>Notoedris chiropteralis</i> (Trouesart, 1896)	<i>Eptesicus serotinus</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Nyctalus noctula</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Pipistrellus pipistrellus</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Pipistrellus javanicus</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Scotophilus kuhlii</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Tylonycteris pachypus</i>	Vespertilionidae	Chiroptera	Asia
<i>Notoedres schoutedeni</i> Fain, 1959	<i>Neoromicia tenuipinnis</i>	Vespertilionidae	Chiroptera	Africa
<i>Notoedres mimetilli</i> Fain, 1959	<i>Mimetillus moloneyi</i>	Vespertilionidae	Chiroptera	Africa
<i>Notoedres africanus</i> (Fain, 1959)	<i>Coleura afra</i>	Emballonuridae	Chiroptera	Africa
<i>Notoedres benoiti</i> Fain, 1959	<i>Coleura afra</i>	Emballonuridae	Chiroptera	Africa
<i>Notoedres verheyeni</i> Fain, 1959	<i>Coleura afra</i>	Emballonuridae	Chiroptera	Africa
<i>Notoedres alexfaini</i> Lavoipierre, 1968	<i>Cheiromeles torquatus</i>	Molossidae	Chiroptera	Asia
“	<i>Cheiromeles parvidens</i>	Molossidae	Chiroptera	Asia
<i>Notoedres cheiromeles</i> Fain, 1959	<i>Cheiromeles torquatus</i>	Molossidae	Chiroptera	Asia
<i>Notoedres elongatus</i> Fain, 1963	<i>Cheiromeles torquatus</i>	Molossidae	Chiroptera	Asia
“	<i>Cheiromeles parvidens</i>	Molossidae	Chiroptera	Asia
<i>Notoedres rajamanickami</i> Lavoipierre, 1968	<i>Cheiromeles torquatus</i>	Molossidae	Chiroptera	Asia
<i>Notoedres namibiensis</i> Klompen, 1992	<i>Tadarida aegyptiaca</i>	Molossidae	Chiroptera	Africa
<i>Notoedres tadaridae</i> Fain, 1959	<i>Chaerephon pumilus</i>	Molossidae	Chiroptera	Africa, including Madagascar
“	<i>Chaerephon jobensis</i>	Molossidae	Chiroptera	Australia
“	<i>Balantiopteryx plicata</i>	Molossidae	Chiroptera	Asia

<i>Notoedres tadaridae</i> Fain, 1959	<i>Mops condylurus</i>	Molossidae	Chiroptera	Africa, including Madagascar
“	<i>Mops midas</i>	Molossidae	Chiroptera	Africa (Madagascar)
“	<i>Mops mops</i>	Molossidae	Chiroptera	Asia
<i>Notoedres dewitti</i> Klompen, Lukoschus, Fain et Nadchatram, 1983	<i>Chaerophon plicata</i>	Molossidae	Chiroptera	Asia
“	<i>Chaerophon jobensis</i>	Molossidae	Chiroptera	Australia
“	<i>Mops condylurus</i>	Molossidae	Chiroptera	Africa, including Madagascar
“	<i>Mops mops</i>	Molossidae	Chiroptera	Asia
<i>Notoedres ismaili</i> Klompen, Lukoschus, Fain et Nadchatram, 1983	<i>Balantiopteryx plicata</i>	Molossidae	Chiroptera	Asia
“	<i>Mops mops</i>	Molossidae	Chiroptera	Asia
<i>Notoedres ovatus</i> Dusbabek, 1980	<i>Mops condylurus</i>	Molossidae	Chiroptera	Africa, including Madagascar
<i>Notoedres tristis</i> Fain et Marshall, 1977	<i>Chaerophon jobensis</i>	Molossidae	Chiroptera	Australia
<i>Notoedres yunkerii</i> Fain, 1962	<i>Molossus molossus</i>	Molossidae	Chiroptera	South America
<i>Notoedres (Bakeracarus)</i> Fain, 1959				
<i>Notoedres paraguayensis</i> Klompen, 1992	<i>Myotis nigricans</i>	Vespertilionidae	Chiroptera	South America
<i>Notoedres plecoti</i> Fain, 1959	<i>Plecotus auritus</i>	Vespertilionidae	Chiroptera	Europe
<i>Notoedres coreanus</i> Ah, 1975	<i>Hypsugo savii</i>	Vespertilionidae	Chiroptera	Asia
<i>Notoedres corynorhini</i> (Fain, 1961)	<i>Corynorhinus rafinesquii</i>	Vespertilionidae	Chiroptera	North America
“	<i>Corynorhinus townsendii</i>	Vespertilionidae	Chiroptera	North America
<i>Notoedres americanus</i> Klompen, 1992	<i>Myotis nigricans</i>	Vespertilionidae	Chiroptera	South America
“	<i>Myotis velifer</i>	Vespertilionidae	Chiroptera	North America
“	<i>Myotis lucifugus</i>	Vespertilionidae	Chiroptera	North America
“	<i>Myotis yumanensis</i>	Vespertilionidae	Chiroptera	North America
“	<i>Myotis volans</i>	Vespertilionidae	Chiroptera	North America
<i>Notoedres eptesicus</i> Fain et Lukoschus, 1971	<i>Eptesicus brasiliensis</i>	Vespertilionidae	Chiroptera	South America
<i>Notoedres helicothrix</i> Fain et Lukoschus, 1975	<i>Cynomops planirostris</i>	Molossidae	Chiroptera	South America
<i>Notoedres anisothrix</i> (Fain et Lukoschus, 1975)	<i>Molossus molossus</i>	Molossidae	Chiroptera	South America
<i>Notoedres lasionycteris</i> (Boyd et Bernstein, 1950)	<i>Lasionycteris noctivagans</i>	Vespertilionidae	Chiroptera	North America
<i>Notoedres lasionycteris minimus</i> (Dusbabek, 1970)	<i>Molossus molossus</i>	Molossidae	Chiroptera	South America
“	<i>Molossus rufus</i>	Molossidae	Chiroptera	South America
“	<i>Nyctinomops laticaudatus</i>	Molossidae	Chiroptera	South America
<i>Notoedres lasionycteris intermedium</i> (Dusbabek, 1970)	<i>Mormopterus minutus</i>	Molossidae	Chiroptera	South America
“	<i>Tadarida brasiliensis</i>	Molossidae	Chiroptera	South America
“	<i>Pteronotus macleayii</i>	Mormoopidae	Chiroptera	South America
“	<i>Eptesicus fuscus</i>	Vespertilionidae	Chiroptera	South America

A review of mammal-associated Psoroptidia

Diablicoptinae Fain et Domrow, 1974				
<i>Diablicoptes</i> Fain et Domrow, 1974				
<i>Diablicoptes sarcophilus</i> Fain et Domrow, 1974	<i>Sarcophilus harrisii</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Satanicoptes</i> Fain et Laurence, 1975				
<i>Satanicoptes armatus</i> Fain et Laurence, 1975	<i>Sarcophilus harrisii</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Satanicoptes phascogale</i> (Domrow et Fain, 1974)	<i>Phascogale tapoatafa</i>	Dasyuridae	Dasyuromorphia	Australia

Table 6. Host distribution of the family Rhyncoptidae Lawrence, 1956

Mite species	Host species	Host family	Host order	Distribution
<i>Rhyncoptes</i> Lawrence, 1956				
<i>Rhyncoptes recurvidens</i> Lawrence, 1956	<i>Hystrix africae australis</i>	Hystricidae	Rodentia	Africa
<i>Rhyncoptes anastosi</i> (Fain, 1962)	<i>Leontopithecus rosalia</i>	Cebidae	Primates	South America
“	<i>Saguinus oedipus</i>	Cebidae	Primates	South America
<i>Rhyncoptes cebi</i> (Fain, 1964)	<i>Cebus albifrons</i>	Cebidae	Primates	South America
<i>Rhyncoptes cercopitheci</i> (Fain, 1964)	<i>Cercopithecus campbelli</i>	Cercopithecidae	Primates	Africa
<i>Rhyncoptes grabberi</i> Klompen, 1989	<i>Macaca mulatta</i>	Cercopithecidae	Primates	Asia
<i>Saimirioptes</i> Fain, 1968				
<i>Saimirioptes paradoxus</i> Fain, 1968	<i>Saimiri sciureus</i>	Cebidae	Primates	South America
<i>Saimirioptes hershkovitzi</i> OConnor, 1987	<i>Cebus apella</i>	Cebidae	Primates	South America
<i>Audycoptes</i> Lavoipierre, 1964				
<i>Audycoptes greeri</i> Lavoipierre, 1964	<i>Saimiri sciureus</i> or <i>boliviensis</i>	Cebidae	Primates	South America
<i>Audycoptes lawrencei</i> Lavoipierre, 1964	<i>Saimiri sciureus</i> or <i>boliviensis</i>	Cebidae	Primates	South America
<i>Ursicoptes</i> Fain et Johnston, 1970				
<i>Ursicoptes americanus</i> Fain et Johnston, 1970	<i>Ursus americanus</i>	Ursidae	Carnivora	North America
“	<i>Ursus maritimus</i>	Ursidae	Carnivora	Europe
<i>Ursicoptes procioni</i> Fain et Wilson, 1979	<i>Procyon lotor</i>	Procionidae	Carnivora	North America
<i>Caenolestocoptes</i> Fain et Lukoschus, 1976				
<i>Caenolestocoptes inca</i> Fain et Lukoschus, 1976	<i>Lestoros inca</i>	Caenolestidae	Paucituberculata	South America

Table 7. Host distribution of the family Chirorhynchobiidae Fain, 1968

Mite species	Host species	Host family	Host order	Distribution
<i>Chirorhynchobia</i> Fain, 1968				
<i>Chirorhynchobia urodermae</i> Fain, 1968	<i>Uroderma bilobatum</i>	Phyllostomidae	Chiroptera	South America
<i>Chirorhynchobia matsoni</i> Yunker, 1970	<i>Anoura geoffroyi</i>	Phyllostomidae	Chiroptera	South America
<i>Chirorhynchobia glossophaga</i> Bochkov, Klompen et OConnor, 2008	<i>Glossophaga soricina</i>	Phyllostomidae	Chiroptera	South America

Table 8. Host distribution of the family Atopomelinae Gunther, 1942

Data by Fain et al. 1986 are mostly omitted because of the numerous accidental records: “p. 371: Il semble donc certain que dans la plupart des cas les hôtes qui sont renseignés ici sont “accidentels” et sont le résultat d'une contamination survenue au cours du transport des hôtes”.

Mite species	Host species	Host family	Host order	Distribution
<i>Atopomelus</i> Trouessart, 1918				
<i>Atopomelus locusta</i> Trouessart, 1918	<i>Neotetracus sinensis</i>	Erinaceidae	Erinaceomorpha	Asia
“	<i>Hylomys suillus</i>	Erinaceidae	Erinaceomorpha	Asia
<i>Atopomelus hylomys</i> Bochkov, Klimov et OConnor, 2005	<i>Hylomys suillus</i>	Erinaceidae	Erinaceomorpha	Asia
<i>Atopomelus priapus</i> Bochkov, Klimov et OConnor, 2005	<i>Neotetracus sinensis</i>	Erinaceidae	Erinaceomorpha	Asia
<i>Atopomelus crocidurae</i> Fain et Lukoschus, 1977	<i>Crocidura attenuata</i>	Soricidae	Soricomorpha	Asia
“	<i>Crocidura monticola</i>	Soricidae	Soricomorpha	Asia
“	<i>Crocidura foetida</i>	Soricidae	Soricomorpha	Asia
“	<i>Crocidura malayana</i>	Soricidae	Soricomorpha	Asia
“	<i>Crocidura mindorus</i>	Soricidae	Soricomorpha	Asia
“	<i>Crocidura grayi</i>	Soricidae	Soricomorpha	Asia
“	<i>Crocidura beatus</i>	Soricidae	Soricomorpha	Asia
<i>Atopomelus talpae</i> Fain, Lukoschus et Cauwenberge, 1973	<i>Talpa romana</i>	Talpidae	Soricomorpha	Asia
<i>Cubanochirus</i> Fain, 1970				
<i>Cubanochirus maximus</i> Fain, 1970	<i>Solenodon cubanus</i>	Solenodontidae	Soricomorpha	South America
<i>Micropotamogalichus</i> Fain, 1970				
<i>Micropotamogalichus congoensis</i> Fain, 1970	<i>Micropotamogale ruwenzorii</i>	Tenrecidae	Afrosoricida	Africa
<i>Listrocarpus</i> Fain, 1967				
<i>Listrocarpus logothrix</i> Fain, 1967	<i>Lagothrix lagotricha</i>	Atelidae	Primates	South America
<i>Listrocarpus cebi</i> Fain, 1972	<i>Cebus albifrons</i>	Cebidae	Primates	South America
<i>Listrocarpus alouatta</i> Fain, 1972	<i>Alouatta macconnelli</i>	Atelidae	Primates	South America
“	<i>Alouatta palliata</i>	Atelidae	Primates	South America
<i>Listrocarpus saimirii</i> Fain, 1967	<i>Saimiri sciureus</i>	Cebidae	Primates	South America
“	<i>Pithecia monachus</i>	Pitheciidae	Primates	South America
“	<i>Saguinus midas</i>	Cebidae	Primates	South America
<i>Listrocarpus surinamensis</i> Fain, 1972	<i>Saimiri sciureus</i>	Cebidae	Primates	South America
<i>Listrocarpus anurus</i> Fain, 1972	<i>Saimiri sciureus</i>	Cebidae	Primates	South America
<i>Listrocarpus cosgrovei</i> Fain, 1972	<i>Callimico goeldii</i>	Cebidae	Primates	South America
“	<i>Saguinus melanoleucus</i>	Cebidae	Primates	South America
“	<i>Saimiri sciureus</i>	Cebidae	Primates	South America
<i>Listrocarpus hapalei</i> Fain, 1967	<i>Callithrix jacchus</i>	Cebidae	Primates	South America
<i>Listrocarpus capucinus</i> Troyo, Solano et Calderon-Alvares, 2004	<i>Cebus capucinus</i>	Cebidae	Primates	South America
<i>Listrocarpus costaricensis</i> Troyo, Solano et Calderon-Alvares, 2004	<i>Saimiri oerstedii</i>	Cebidae	Primates	South America
<i>Listrocarpus spinifer</i> Fain et Lukoschus, 1976	<i>Caluromys lanatus</i>	Didelphidae	Didelphimorphia	South America
<i>Lemuroptes</i> Lawrence, 1958				
<i>Lemuroptes primarius</i> Lawrence, 1958	<i>Galago alleni</i>	Galagidae	Primates	Africa

A review of mammal-associated Psoroptidia

<i>Lemuroptes bursatus</i> Fain, 1972	<i>Otolemur crassicaudatus</i>	Galagidae	Primates	Africa
“	<i>Otolemur crassicaudatu</i>	Galagidae	Primates	Africa
<i>Lemuroptes zumpti</i> Fain, 1972	<i>Otolemur crassicaudatu</i>	Galagidae	Primates	Africa
<i>Lemuroptes attenuatus</i> Fain, 1972	<i>Galago demidoff</i>	Galagidae	Primates	Africa
<i>Lemuroptes potto</i> Fain, 1972	<i>Perodicticus potto</i>	Lorisidae	Primates	Africa
<i>Austrochirus</i> Womersley, 1943				
<i>Austrochirus queenslandicus</i> Womersley, 1943	Phalangeridae sp.	Phalangeridae	Diprotodontia	Australia
“	<i>Isoodon obesulus</i>	Peramelidae	Peramelimorphia	Australia
“	<i>Isoodon macrourus</i>	Peramelidae	Peramelimorphia	Australia
“	<i>Perameles nasuta</i>	Peramelidae	Peramelimorphia	Australia
<i>Austrochirus squamiferus</i> Fain, 1970	<i>Perameles nasuta</i>	Peramelidae	Peramelimorphia	Australia
<i>Austrochirus armatus</i> Fain, 1972	<i>Perameles nasuta</i>	Peramelidae	Peramelimorphia	Australia
“	<i>Isoodon obesulus</i>	Peramelidae	Peramelimorphia	Australia
<i>Austrochirus peroryctes</i> Fain et Bochkov, 2003	<i>Peroryctes raffrayana</i>	Peramelidae	Peramelimorphia	Australia
<i>Austrochirus perameles</i> Fain, 1971	<i>Isoodon macrourus</i>	Peramelidae	Peramelimorphia	Australia (New Guinea)
<i>Austrochirus dorreensis</i> Fain et Lukoschus, 1981	<i>Perameles bougainville</i>	Peramelidae	Peramelimorphia	Australia
<i>Austrochirus</i> (<i>Austrochiroides</i>) Fain, 1970				
<i>Austrochirus mcmillani</i> Domrow, 1961	Peramelidae sp.	Peramelidae	Peramelimorphia	Australia (New Guinea)
“	<i>Echymipera kalubu</i>	Peramellidae	Peramelimorphia	Australia (New Guinea)
“	<i>Peroryctes raffrayana</i>	Peramelidae	Peramelimorphia	Australia (New Guinea)
“	<i>Perameles nasuta</i>	Peramelidae	Peramelimorphia	Australia
“	<i>Anisomys imitator</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Melomys rufescens</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Austrochirus womersleyi</i> Fain, 1972	<i>Perameles moresbyensis</i>	Peramelidae	Peramelimorphia	Australia (New Guinea)
“	<i>Peroryctes raffrayana</i>	Peramelidae	Peramelimorphia	Australia (New Guinea)
<i>Austrochirus dubininae</i> Fain et Bochkov, 2003	<i>Isoodon macrourus</i>	Peramelidae	Peramelimorphia	Australia (New Guinea)
<i>Sclerochiroopsis</i> Fain, 1972				
<i>Sclerochiroopsis filmeri</i> (Domrow, 1960)	<i>Macrotis lagotis</i>	Thylacomyidae	Peramelimorphia	Australia
<i>Petrogalochirus</i> Fain, 1970				
<i>Petrogalochirus tasmaniensis</i> Fain, 1970	<i>Petrogale penicillata</i>	Macropodidae	Diprotodontia	Australia
<i>Petrogalochirus dycei</i> (Domrow, 1960)	<i>Trichosurus vulpecula</i>	Phalangeridae	Diprotodontia	Australia
“	<i>Trichosurus caninus</i>	Phalangeridae	Diprotodontia	Australia
<i>Petrogalochirus macropus</i> Fain et Domrow, 1974	<i>Macropus rufogriseus</i>	Macropodidae	Diprotodontia	Australia
<i>Koalachirus</i> Fain, 1970				
<i>Koalachirus perkinsi</i> (Domrow, 1958)	<i>Phascolarctos cinereus</i>	Phascolarctidae	Diprotodontia	Australia

<i>Phalangerobia</i> Fain, 1972				
<i>Phalangerobia ovata</i> Fain, 1972	<i>Phalanger intercastellanus</i>	Phalangeridae	Diprotodontia	Australia (New Guinea)
<i>Atellana</i> Domrow, 1968				
<i>Atellana papilio</i> Domrow, 1968	<i>Trichosurus vulpecula</i>	Phalangeridae	Diprotodontia	Australia
“	<i>Trichosaurus caninus</i>	Phalangeridae	Diprotodontia	Australia
<i>Petaurobia</i> Fain, 1971				
<i>Petaurobia papuana</i> Fain, 1971	<i>Petaurus breviceps</i>	Phalangeridae	Diprotodontia	Australia (New Guinea)
<i>Petaurobia dactylopsila</i> Fain, 1971	<i>Dactylopsila trivirgata</i>	Phalangeridae	Diprotodontia	Australia
<i>Petaurobia australiana</i> Fain et Lukoschus, 1981	<i>Petaurus breviceps</i>	Phalangeridae	Diprotodontia	Australia
<i>Campylochirus</i> Trouessart, 1893				
<i>Campylochirus chelopus</i> Trouessart, 1893	<i>Pseudocheirus peregrinus</i>	Phalangeridae	Diprotodontia	Australia
<i>Campylochirus brevipenis</i> Fain et Domrow, 1976	<i>Pseudocheirus peregrinus</i>	Phalangeridae	Diprotodontia	Australia
<i>Campylochirus antechinus</i> Fain, 1971	<i>Antechinus stuartii</i>	Dasyuridae	Dasyuromorpha	Australia
“	<i>Antechinus flavipes</i>	Dasyuridae	Dasyuromorpha	Australia
“	<i>Dasyurus viverrinus</i>	Dasyuridae	Dasyuromorpha	Australia
“	<i>Petauroides volans</i>	Pseudocheiridae	Diprotodontia	Australia
<i>Campylochirus stenophallus</i> Fain et Domrow, 1976	<i>Petauroides volans</i>	Pseudocheiridae	Diprotodontia	Australia
<i>Campylochirus pseudocheirus</i> Fain, 1972	<i>Pseudochirulus forbesi</i>	Pseudocheiridae	Diprotodontia	Australia (New Guinea)
“	<i>Pseudochirulus mayeri</i>	Pseudocheiridae	Diprotodontia	Australia
<i>Campylochirus caparti</i> Fain, 1974	<i>Pseudocheirus</i> sp.	Pseudocheiridae	Diprotodontia	Australia (New Guinea)
<i>Campylochirus petauricola</i> Fain, 1972	<i>Petaurus breviceps</i>	Petauridae	Diprotodontia	Australia (New Guinea)
“	<i>Petaurus breviceps</i>	Petauridae	Diprotodontia	Australia
<i>Campylochirus brevicepsicola</i> Fain et Lukoschus, 1981	<i>Petaurus breviceps</i>	Petauridae	Diprotodontia	Australia
<i>Campylochiropsis</i> Fain, 1972				
<i>Campylochiropsis dolichurus</i> (Fain, 1972)	<i>Petauroides volans</i>	Pseudocheirida	Diprotodontia	Australia
<i>Campylochiropsis missonei</i> Fain, 1974	<i>Pseudocheirus</i> sp.	Pseudocheirida	Diprotodontia	Australia (New Guinea)
<i>Campylochiropsis micrura</i> Fain et Domrow, 1974	<i>Pseudocheirus peregrinus</i>	Phalangeridae	Diprotodontia	Australia
<i>Distoechurobia</i> Fain, 1972				
<i>Distoechurobia anomala</i> Fain, 1972	<i>Distoechurus pennatus</i>	Acrobatidae	Diprotodontia	Australia (New Guinea)
<i>Sclerochiroides</i> Fain, 1970				
<i>Sclerochiroides mirabilis</i> Fain, 1970	<i>Peroryctes raffrayana</i>	Peramelidae	Peramelemorphia	Australia (New Guinea)
“	<i>Microperoryctes longicauda</i>	Peramelidae	Peramelemorphia	Australia
“	<i>Petauroides volans</i>	Pseudocheirida	Diprotodontia	Australia
<i>Austrobius</i> Fain, 1971				
<i>Austrobius alatus</i> Fain, 1971	<i>Myrmecobius fasciatus</i>	Myrmecobiidae	Dasyuromorphia	Australia

A review of mammal-associated Psoroptidia

<i>Austrobius alatus</i> Fain, 1971	<i>Sminthopsis fuliginosus</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Atopomelopsis</i> Fain, 1972 stat. res.				
<i>Atopomelopsis longipilis</i> (Fain, 1971)	<i>Macrotis leucura</i>	Thylacomyidae	Peramelemorpha	Australia
<i>Dasyurochirus</i> Fain, 1970				
<i>Dasyurochirus biscutatus</i> Fain, 1970	<i>Sminthopsis murina</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Antechinomys laniger</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Dasyurochirus sminthopsis</i> (Womersley, 1954)	<i>Sminthopsis crassicaudata</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Sminthopsis leucopus</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Sminthopsis murina</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Dasyuroides byrnei</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Dasyurochirus trouessarti</i> (Domrow, 1961)	<i>Antechinus godmani</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Antechinus flavipes</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Dasyurus hallucatus</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Dasyurochirus australis</i> Fain, 1972	<i>Antechinus minimus</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Antechinus flavipes</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Antechinus stuartii</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Phascogale calura</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Dasyurochirus intercalatus</i> Fain, 1972	<i>Sminthopsis murina</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Antechinus flavipes</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Dasyurochirus inermis</i> Fain, 1971	<i>Micromurexia habbema</i>	Dasyuridae	Dasyuromorphia	Australia (New Guinea)
<i>Dasyurochirus squamatus</i> (Fain, 1972)	<i>Antechinus flavipes</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Antechinus swainsonii</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Phascogale</i> sp.	Dasyuridae	Dasyuromorphia	Australia
<i>Dasyurochirus longissimus</i> Fain et Lukoschus, 1981	<i>Acrobates pygmaeus</i>	Acrobatidae	Diprotodontia	Australia
<i>Dasyurochirus major</i> Fain et Domrow, 1973	<i>Dasyurus maculatus</i>	Dasyuridae	Dasyuromorphia	Australia (New Guinea)
<i>Dasyurochirus radovskyi</i> Fain, 1975	<i>Dasyurus albopunctatus</i>	Dasyuridae	Dasyuromorphia	Australia (New Guinea)
<i>Dasyurochirus longicaudatus</i> Fain et Lukoschus, 1981	<i>Antechinus godmani</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Dasyurochirus tapoatafa</i> Fain et Lukoschus, 1981	<i>Phascogale tapoatafa</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Dasyurochirus granulipes</i> Fain et Lukoschus, 1981	<i>Sminthopsis granulipes</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Dasyurochirus leprosus</i> Fain, 1971	<i>Myrmecobius fasciatus</i>	Myrmecobiidae	Dasyuromorphia	Australia
“	<i>Neophascogale lorentzi</i>	Dasyuridae	Dasyuromorphia	Australia (New Guinea)
<i>Dasyurochirus</i> (<i>Dasyurochiroides</i>) Fain, 1972				
<i>Dasyurochirus gracilis</i> Fain, 1972	<i>Antechinus hageni</i>	Dasyuridae	Dasyuromorphia	Australia (New Guinea)
<i>Labidopygus</i> Fain et Domrow, 1973				
<i>Labidopygus australiensis</i> Fain et Domrow, 1973	<i>Micromurexia habema</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Labidopygus squamatus</i> Fain et Lukoschus, 1981	<i>Antechinus swainsonii</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Antechinus flavipes</i>	Dasyuridae	Dasyuromorphia	Australia

<i>Scolonoticus</i> Fain, 1971				
<i>Scolonoticus brevis</i> Fain, 1971	<i>Antechinus flavipes</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Scolonoticus medius</i> Fain, 1972	<i>Sminthopsis crassicaudata</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Antechinus stuartii</i>	Dasyuridae	Dasyuromorphia	Australia
“	<i>Acrobates pygmaeus</i>	Acrobatidae	Diprotodontia	Australia
<i>Scolonoticus petaurus</i> Fain, 1972	<i>Petaurus breviceps</i>	Petauridae	Diprotodontia	Australia
<i>Scolonoticus quasinudus</i> Fain et Lukoschus, 1981	<i>Petaurus breviceps</i>	Petauridae	Diprotodontia	Australia
<i>Cytostethum</i> Domrow, 1956				
<i>Cytostethum promeces</i> Domrow, 1956	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum trachypyx</i> Domrow, 1956	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum charactum</i> Domrow, 1956	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum pseudocharactum</i> Domrow, 1956	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum nanophyes</i> Domrow, 1956	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum domrowi</i> Fain, 1972	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum mollisoni</i> Domrow, 1961	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
“	<i>Macropus dorsalis</i>	Macropodidae	Diprotodontia	Australia
<i>Cytostethum potorous</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum mediostriatum</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum procharactum</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum mesocharactum</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum tubiferum</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum pseudotubiferum</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum neotubiferum</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum toloomense</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum hamatum</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum gracile</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum dendriticum</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum postscutatum</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum spinulatum</i> Fain et Domrow, 1974	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum macropus</i> Fain, 1971	<i>Potorous tridactylus</i>	Potoridae	Diprotodontia	Australia
“	<i>Macropus rufogriseus</i>	Macropodidae	Diprotodontia	Australia
“	<i>Macropus dorsalis</i>	Macropodidae	Diprotodontia	Australia
<i>Cytostethum nudum</i> Fain, 1972	<i>Myrmecobius fasciatus</i>	Myrmecobiidae	Dasyuromorphia	Australia
<i>Cytostethum (Metacytostethum)</i> Fain, 1971				
<i>Cytostethum thylogale</i> Fain, 1971	<i>Thylogale</i> sp.	Macropodidae	Diprotodontia	Australia
“	<i>Bettongia gaimardi</i>	Potoridae	Diprotodontia	Australia

A review of mammal-associated Psoroptidia

<i>Cytostethum prosquamatum</i> Fain et Domrow, 1974	<i>Thylogale brunii</i>	Macropodidae	Diprotodontia	Australia
<i>Cytostethum mediosquamatum</i> Fain et Domrow, 1974	<i>Thylogale brunii</i>	Macropodidae	Diprotodontia	Australia (New Guinea)
“	<i>Dorcopsulus vanheurni</i>	Macropodidae	Diprotodontia	Australia (New Guinea)
<i>Cytostethum postsquamatum</i> Fain et Domrow, 1974	<i>Thylogale billardierii</i>	Macropodidae	Diprotodontia	Australia (New Guinea)
<i>Cytostethum asquamatum</i> Fain, 1975	<i>Dorcopsulus</i> sp.	Macropodidae	Diprotodontia	Australia (New Guinea)
“	<i>Dorcopsulus vanheurni</i>	Macropodidae	Diprotodontia	Australia (New Guinea)
<i>Cytostethum wilsoni</i> Fain, 1975	<i>Dorcopsulus vanheurni</i>	Macropodidae	Diprotodontia	Australia (New Guinea)
<i>Cytostethum murmurens</i> Fain, 1975	<i>Dorcopsulus vanheurni</i>	Macropodidae	Diprotodontia	Australia (New Guinea)
<i>Cytostethum euphallus</i> Fain et Domrow, 1974	<i>Thylogale stigmatica</i>	Macropodidae	Diprotodontia	Australia
<i>Cytostethum mydenense</i> Fain et Domrow, 1974	<i>Thylogale billardierii</i>	Macropodidae	Diprotodontia	Australia
<i>Cytostethum thetis</i> Fain et Domrow, 1974	<i>Thylogale thetis</i>	Macropodidae	Diprotodontia	Australia
<i>Cytostethum inerme</i> Fain et Domrow, 1974	<i>Thylogale billardierii</i>	Macropodidae	Diprotodontia	Australia
<i>Cytostethum intermedium</i> Fain et Domrow, 1974	<i>Bettongia gaimardi</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum tasmaniense</i> Fain et Domrow, 1974	<i>Bettongia gaimardi</i>	Potoridae	Diprotodontia	Australia
“	<i>Bettongia penicillata</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum clibanarius</i> Domrow, 1961	<i>Aepyprymnus rufescens</i>	Potoridae	Diprotodontia	Australia
<i>Cytostethum longitarsus</i> Fain, 1972	<i>Echymipera kalubu</i>	Peramelidae	Peramelemorphia	Australia (New Guinea)
<i>Cytostethum wallabia</i> Fain et Domrow, 1984	<i>Wallabia bicolor</i>	Macropodidae	Diprotodontia	Australia
<i>Neolabidocarpus</i> Gunther, 1942				
<i>Neolabidocarpus buloloensis</i> (Gunther, 1940)	<i>Thylogale coxenii</i>	Macropodidae	Diprotodontia	Australia (New Guinea)
<i>Procytostethum</i> Fain, 1971				
<i>Procytostethum parvum</i> (Domrow, 1961)	<i>Hypsiprymnodon moschatus</i>	Hypsiprymnodontidae	Diprotodontia	Australia
<i>Notoryctobia</i> Fain et Lukoschus, 1981				
<i>Notoryctobia notoryctes</i> Fain et Lukoschus, 1981	<i>Notoryctes typhlops</i>	Notoryctidae	Notoryctero-morphia	Australia
<i>Dromiciolichus</i> Fain, 1970				
<i>Dromiciolichus chiliensis</i> Fain, 1970	<i>Dromiciops gliroides</i>	Microbiotheriidae	Microbiotheria	South America
<i>Prodidelphoecius</i> Fain, 1976				
<i>Prodidelphoecius euphallus</i> Fain, 1976	<i>Monodelphis scalops</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius</i> Fain, 1970				
<i>Didelphoecius paranensis</i> Fain, 1970	<i>Monodelphis dimidiata</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius monodelphis</i> Fain, 1970	<i>Monodelphis brevicaudata</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius incisus</i> Fain, 1970	<i>Monodelphis brevicaudata</i>	Didelphidae	Didelphimorphia	South America

<i>Didelphoecius colombianus</i> Fain, 1970	<i>Didelphis aurita</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius colombianus manaosensis</i> Fain, 1979	<i>Caluromys philander</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius scutatus</i> Fain, 1976	<i>Caluromys philander</i>	Didelphidae	Didelphimorphia	South America
“	<i>Caluromys derbianus</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius didelphicola</i> Fain, 1970	<i>Didelphis aurita</i>	Didelphidae	Didelphimorphia	South America
“	<i>Marmosa murina</i>	Didelphidae	Didelphimorphia	South America
“	<i>Micoureus demerarae</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius philander</i> Fain, 1970	<i>Caluromys philander</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius surinamensis</i> Fain, 1976	<i>Caluromys philander</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius dissimilis</i> Fain, 1976	<i>Caluromys philander</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius capirangensis</i> Fain, 1970	<i>Marmosa</i> sp.	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius marmosae</i> Fain, 1970	<i>Marmosa</i> sp.	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius palmeriensis</i> Fain, 1976	<i>Gracilinanus microtarsus</i>	Didelphidae	Didelphimorphia	South America
“	<i>Gracilinanus agilis</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius coalescens</i> Fain, 1979	<i>Marmosa</i> sp.	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius guyanensis</i> Fain, 1970	<i>Marmosa murina</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius squamatus</i> Fain, 1976	<i>Thylamis pusillus</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius inversus</i> Fain, 1976	<i>Marmosa robinsoni</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius venezuelensis</i> Fain, 1979	<i>Marmosa murina</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius callipygus</i> Fain, 1976	<i>Monodelphis breviceaudata</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphoecius validus</i> Fain, Zanatta-Coutincho et Fonseca, 1996	<i>Metachirus nudicaudatus</i>	Didelphidae	Didelphimorphia	South America
<i>Metadidelphoecius</i> Fain, 1979				
<i>Metadidelphoecius peruvianus</i> Fain, 1979	<i>Marmosops impavidus</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphilichus</i> Fain, 1970				
<i>Didelphilichus serrifer</i> Fain, 1970	<i>Didelphis aurita</i>	Didelphidae	Didelphimorphia	South America
“	<i>Didelphis marsupialis</i>	Didelphidae	Didelphimorphia	North and South America
“	<i>Didelphis albiventris</i>	Didelphidae	Didelphimorphia	South America
<i>Didelphilichus philander</i> Fain, 1970	<i>Philander opossum</i>	Didelphidae	Didelphimorphia	South America
<i>Metachiroecius</i> Fain, Zanatta-Coutincho et Fonseca, 1996				
<i>Metachiroecius brasiliensis</i> Fain, Zanatta-Coutincho et Fonseca, 1996	<i>Metachirus nudicaudatus</i>	Didelphidae	Didelphimorphia	South America
<i>Caenolestolichus</i> Fain et Bochkov, 2003				
<i>Caenolestolichus lukoschusi</i> Fain et Bochkov, 2003	<i>Caenolestes fuliginosus</i>	Caenolestidae	Paucituberculata	South America
<i>Murichirus</i> Fain, 1971				
<i>Murichirus enoplus</i> (Domrow, 1956)	<i>Hydromys chrysogaster</i>	Muridae	Rodentia	Australia
“	<i>Anisomys imitator</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Rattus niobe</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Rattus fuscipes</i>	Muridae	Rodentia	Australia
“	<i>Melomys lutillus</i>	Muridae	Rodentia	Australia

A review of mammal-associated Psoroptidia

<i>Murichirus enoplus</i> (Domrow, 1956)	<i>Melomys cervinipes</i>	Muridae	Rodentia	Australia
“	<i>Hyomys goliath</i> or <i>Mallomys rothschildi</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Uromys caudimaculatus</i>	Muridae	Rodentia	Australia
<i>Murichirus pogonomys</i> Fain, 1972	<i>Chiruromys vates</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus melomys</i> Fain, 1972	<i>Melomys rufescens</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Uromys [brunyii]</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Rattus praetor</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus leopoldi</i> Fain, 1974	<i>Paramelomys platyops</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus scorteus</i> Fain, 1974	<i>Paramelomys platyops</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus coriaceus</i> Fain, 1974	<i>Rattus exulans</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Paramelomys platyops</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus coriaceus gratus</i> Fain, 1974	<i>Paramelomys mollis</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Paramelomys rubex</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Paramelomys moncktoni</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus longior</i> Fain, 1974	<i>Paramelomys platyops</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus coxatus</i> Fain, 1977	<i>Paramelomys rubex</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Paramelomys moncktoni</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus simplex</i> Fain, 1977	<i>Paramelomys rubex</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus quasimelomys</i> Fain et Lukoschus, 1981	<i>Melomys</i> sp.	Muridae	Rodentia	Australia
“	<i>Melomys cervinipes</i>	Muridae	Rodentia	Australia
<i>Murichirus lobatitarsis</i> Fain et Lukoschus, 1981	<i>Melomys</i> sp.	Muridae	Rodentia	Australia
“	<i>Melomys cervinipes</i>	Muridae	Rodentia	Australia
<i>Murichirus vermicularis</i> (Fain et Lukoschus, 1981)	<i>Melomys cervinipes</i>	Muridae	Rodentia	Australia
<i>Murichirus maximus</i> Fain, 1976	<i>Uromys anak</i>	Muridae	Rodentia	Australia
<i>Murichirus alatus</i> Fain et Lukoschus, 1981	<i>Notomys alexis</i>	Muridae	Rodentia	Australia
“	<i>Pseudomys hermannsburgensis</i>	Muridae	Rodentia	Australia
<i>Murichirus dorsostriatus</i> Fain et Lukoschus, 1981	<i>Notomys alexis</i>	Muridae	Rodentia	Australia
<i>Murichirus dorsoscutatus</i> Fain et Lukoschus, 1981	<i>Notomys alexis</i>	Muridae	Rodentia	Australia
<i>Murichirus notomys</i> Fain, 1972	<i>Notomys mitchellii</i>	Muridae	Rodentia	Australia
<i>Murichirus zyzomys</i> Fain et Lukoschus, 1981	<i>Zyzomys argurus</i>	Muridae	Rodentia	Australia

<i>Murichirus mastacomys</i> Fain et Lukoschus, 1981	<i>Mastacomys fuscus</i>	Muridae	Rodentia	Australia
<i>Murichirus moschati</i> (Domrow, 1961)	<i>Hypsiprymnodon moschatus</i>	Hypsiprymnodontidae	Diprotodontia	Australia
<i>Murichirus parahydromys</i> Fain, 1971	<i>Parahydromys asper</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Hydromys chrysogaster</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus pseudohydromys</i> Fain, 1971	<i>Pseudohydromys murinus</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Murichirus ornatus</i> Fain, 1972	<i>Pseudochirulus forbesi</i>	Phalangeridae	Diprotodontia	Australia (New Guinea)
<i>Murochirus anabiotus</i> Domrow, 1992	<i>Pseudocheirus peregrinus</i>	Phalangeridae	Diprotodontia	Australia
<i>Microchirus petaurus</i> Fain et Lukoschus, 1981	<i>Petaurus breviceps</i>	Petauridae	Diprotodontia	Australia
<i>Chirodiscoides</i> Hirst, 1917				
<i>Chirodiscoides caviae</i> Hirst, 1917	<i>Cavia porcellus</i>	Caviidae	Rodentia	Cosmopolite
“	<i>Cavia aperea</i>	Caviidae	Rodentia	South America
“	<i>Microcavia australis</i>	Caviidae	Rodentia	South America
“	<i>Galea musteloides</i>	Caviidae	Rodentia	South America
<i>Chirodiscoides galeae</i> Fain, 1972	<i>Galea spixii</i>	Caviidae	Rodentia	South America
<i>Chirodiscoides didelphicola</i> Fain, 1970	! <i>Didelphis albiventris</i>	Didelphidae	Didelphimorphia	South America
“	<i>Kannabateomys amblyonyx</i>	Echimyidae	Rodentia	South America
<i>Chirodiscoides inglesae</i> Fain, 1970	! <i>Lycalopex gymnocercus</i>	Canidae	Carnivora	South America
“	<i>Proechimys guyannensis</i>	Echimyidae	Rodentia	South America
<i>Chirodiscoides interruptus</i> Fain, 1970	<i>Proechimys guyannensis</i>	Echimyidae	Rodentia	South America
“	! <i>Caluromys philander</i>	Didelphidae	Didelphimorphia	South America
<i>Chirodiscoides cercomys</i> Fain, 1970	<i>Trichomys apereoides</i>	Echimyidae	Rodentia	South America
<i>Chirodiscoides echimys</i> Fain, 1970	<i>Phyllomys thomasi</i>	Echimyidae	Rodentia	South America
<i>Chirodiscoides proechimys</i> Fain, 1972	<i>Proechimys guyannensis</i>	Echimyidae	Rodentia	South America
“	<i>Proechimys brevicauda</i>	Echimyidae	Rodentia	South America
“	<i>Proechimys cuvieri</i>	Echimyidae	Rodentia	South America
“	<i>Proechimys quadruplicatus</i>	Echimyidae	Rodentia	South America
<i>Chirodiscoides euryzgomatomys</i> Fain, 1972	<i>Euryzgomatomys spinosus</i>	Echimyidae	Rodentia	South America
<i>Chirodiscoides bolivianus</i> Fain, 1976	<i>Proechimys</i> sp.	Echimyidae	Rodentia	South America
“	<i>Proechimys semispinosus</i>	Echimyidae	Rodentia	South America
<i>Chirodiscoides ascuamatus</i> Fain et Lukoschus, 1976	! <i>Marmosa murina</i>	Didelphidae	Didelphimorphia	South America
<i>Isothricola</i> Fain, 1970				
<i>Isothricola appendiculatus</i> Fain, 1970	<i>Isothrix bistrata</i>	Echimyidae	Rodentia	South America
<i>Isothricola amazonicus</i> Fain, 1970	<i>Isothrix bistrata</i>	Echimyidae	Rodentia	South America
<i>Isothricola bolivianus</i> Fain, 1970	<i>Proechimys</i> sp.	Echimyidae	Rodentia	South America

A review of mammal-associated Psoroptidia

<i>Isothricola bolivianus</i> Fain, 1970	<i>Proechimys semispinosus</i>	Echimyidae	Rodentia	South America
<i>Isothricola coniformis</i> Fain, 1970	<i>Proechimys guyannensis</i>	Echimyidae	Rodentia	South America
<i>Isothricola ovatus</i> Fain, 1970	<i>Proechimys guyannensis</i>	Echimyidae	Rodentia	South America
<i>Myocastorobia</i> Fain, 1975				
<i>Myocastorobia myocastor</i> (Fain, 1970)	<i>Myocastor coypus</i>	Myocastoridae	Rodentia	North and South America
<i>Oryzomysia</i> Fain, 1970				
<i>Oryzomysia longilobata</i> Fain, 1970	<i>Oryzomys</i> sp.	Cricetidae	Rodentia	South America
<i>Oryzomysia peruviana</i> Fain, 1970	<i>Oryzomys nitidus</i>	Cricetidae	Rodentia	South America
<i>Oryzomysia peruviana punctulata</i> Fain, 1979	<i>Oecomys trinitatis</i>	Cricetidae	Rodentia	South America
<i>Oryzomysia peruviana boliviana</i> Fain, 1979	<i>Oryzomys</i> sp.	Cricetidae	Rodentia	South America
<i>Oryzomysia peruviana costaricensis</i> Fain, 1979	<i>Nectomys</i> sp.	Cricetidae	Rodentia	South America
<i>Oryzomysia oryzomys</i> (Radford, 1954)	<i>Oryzomys palustris</i>	Cricetidae	Rodentia	South America
“	<i>Holochilus brasiliensis</i>	Cricetidae	Rodentia	South America
“	<i>Zygodontomys brevicauda</i>	Cricetidae	Rodentia	South America
<i>Oryzomysia oryzomys brevior</i> Fain, 1979	<i>Melanomys calignosus</i>	Cricetidae	Rodentia	South America
<i>Oryzomysia oxymycterus</i> Fain, 1976	<i>Oxymycterus quaestor</i>	Cricetidae	Rodentia	South America
“	<i>Ichthyomys hydrobates</i>	Cricetidae	Rodentia	South America
<i>Oryzomysia lukoschusi</i> Fain, 1976	<i>Nectomys squamipes</i>	Cricetidae	Rodentia	South America
<i>Oryzomysia emarginata</i> Fain, 1979	<i>Holochilus brasiliensis</i>	Cricetidae	Rodentia	South America
“	<i>Nectomys squamipes</i>	Cricetidae	Rodentia	South America
“	<i>Oligoryzomys longicaudatus</i>	Cricetidae	Rodentia	South America
<i>Oryzomysia striata</i> Fain, 1979	<i>Thomasomys ischyurus</i>	Cricetidae	Rodentia	South America
<i>Oryzomysia neacomys</i> Fain, 1979	<i>Neacomys spinosus</i>	Cricetidae	Rodentia	South America
<i>Euryzygomysia</i> Fain, 1979				
<i>Euryzygomysia catharinae</i> Fain, 1979	<i>Euryzygomatomys spinosus</i>	Echimyidae	Rodentia	South America
<i>Capromysia</i> Fain, 1979				
<i>Capromysia elongatus</i> (Fain, 1970)	<i>Mesocapromys nanus</i>	Capromyidae	Rodentia	South America
“	<i>Mesocapromys melanurus</i>	Muridae	Rodentia	South America
“	<i>Mysateles prehensilis</i>	Capromyidae	Rodentia	South America
“	<i>Capromys pilorides</i>	Capromyidae	Rodentia	South America
<i>Capromylichus</i> Fain, 1970				
<i>Capromylichus cubanus</i> Fain, 1970	<i>Mesocapromys nanus</i>	Capromyidae	Rodentia	South America
“	<i>Mesocapromys auritus</i>	Capromyidae	Rodentia	South America
<i>Plagiodontochirus</i> Fain, 1976				
<i>Plagiodontochirus nudus</i> Fain, 1976	<i>Plagiodontia aedium</i>	Capromyidae	Rodentia	South America
<i>Domingoecius</i> Fain et Lukoschus, 1979				
<i>Domingoecius cruzi</i> Fain et Lukoschus, 1979	<i>Plagiodontia aedium</i>	Capromyidae	Rodentia	South America
<i>Bathyergolichus</i> Fain, 1970				

<i>Bathyergolichus bathyergians</i> (Radford, 1939)	<i>Bathyergus suillus</i>	Bathyergidae	Rodentia	Africa
<i>Bathyergolichus zumpti</i> (Lawrence, 1956)	<i>Georychus capensis</i>	Bathyergidae	Rodentia	Africa
<i>Bathyergolichus heliophobius</i> Fain, 1970	<i>Heliophobius argenteocinereus</i>	Bathyergidae	Rodentia	Africa
<i>Bathyergolichus hottentotus</i> Fain, 1986 (= <i>Bathyergolichus cryptomys</i> Fain et Bochkov, 2003, syn. nov.)	<i>Cryptomys hottentotus</i>	Bathyergidae	Rodentia	Africa
<i>Listrophoroides</i> Hirst, 1923				
<i>Listrophoroides aethiopicus</i> Hirst, 1923	<i>Cricetomys gambianus</i>	Nesomyidae	Rodentia	Africa
“	<i>Cricetomys ansorgei</i>	Nesomyidae	Rodentia	Africa
<i>Listrophoroidea dasymys</i> Radford, 1942	<i>Dasymys incommutus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides oenomys</i> Fain, 1972	<i>Oenomys hypoxanthus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides spectabilis</i> Fain, Hart et Rahm, 1986	<i>Thamnomys</i> sp.	Muridae	Rodentia	Africa
<i>Listrophoroides borneoensis</i> Fain, 1970	<i>Sundamys muelleri</i>	Muridae	Rodentia	Asia
“	<i>Rattus argentiventer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides ptilocercus</i> Fain, 1970	<i>Sundamys muelleri</i>	Muridae	Rodentia	Asia
<i>Listrophoroides longiventris</i> Fain, 1976	<i>Rattus</i> sp.	Muridae	Rodentia	Asia
<i>Listrophoroides mindanensis</i> Fain et Lukoschus, 1981	<i>Batomys salomonseni</i>	Muridae	Rodentia	Asia
<i>Listrophoroides batomys</i> Bochkov et OConnor, 2006	<i>Batomys granti</i>	Muridae	Rodentia	Asia
<i>Listrophoroides pahangi</i> Fain, 1974	<i>Leopoldamys sabanus</i>	Muridae	Rodentia	Asia
“	<i>Leopoldamys edwardsi</i>	Muridae	Rodentia	Asia
<i>Listrophoroides hongkongensis</i> Fain, 1974	<i>Niviventer fulvescens</i>	Muridae	Rodentia	Asia
<i>Listrophoroides cremoriventer</i> Fain, 1976	<i>Niviventer cremoriventer</i>	Muridae	Rodentia	Asia
“	<i>Niviventer langbianis</i>	Muridae	Rodentia	Asia
<i>Listrophoroides rapit</i> Bochkov, OConnor et Gorog, 2004	<i>Niviventer rapit</i>	Muridae	Rodentia	Asia
<i>Listrophoroides bilineatus</i> Fain, 1975	<i>Rattus andamanensis</i>	Muridae	Rodentia	Asia
<i>Listrophoroides bowersi</i> Fain, 1979	<i>Berylmys bowersi</i>	Muridae	Rodentia	Asia
“	<i>Dacnomys millardi</i>	Muridae	Rodentia	Asia
<i>Listrophoroides brachypyx</i> Fain, 1974	<i>Rattus exulans</i>	Muridae	Rodentia	Asia
“	<i>Rattus tiomanicus</i>	Muridae	Rodentia	Asia
“	<i>Rattus tanezumi</i>	Muridae	Rodentia	Asia
“	<i>Rattus mindorensis</i>	Muridae	Rodentia	Asia
<i>Listrophoroides infraluteus</i> Fain, 1976	<i>Sundamys infraluteus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides laosensis</i> Fain, 1979	<i>Leopoldamys edwardsi</i>	Muridae	Rodentia	Asia
<i>Listrophoroides pakistanicus</i> Fain et Hyland, 1980	<i>Millardia meltada</i>	Muridae	Rodentia	Asia
“	<i>Mus musculus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides exilis</i> Fain et Hyland, 1980	<i>Bandicota bengalensis</i>	Muridae	Rodentia	Asia
<i>Listrophoroides pseudolongiventris</i> Fain, 1976	<i>Rattus</i> sp.	Muridae	Rodentia	Asia

A review of mammal-associated Psoroptidia

<i>Listrophoroides crunomys</i> Bochkov et OConnor, 2006	<i>Crunomys suncooides</i>	Muridae	Rodentia	Asia
"	<i>Crunomys melanius</i>	Muridae	Rodentia	Asia
<i>Listrophoroides insularis</i> Bochkov et OConnor, 2006	<i>Chrotomys whiteheadi</i>	Muridae	Rodentia	Asia
"	<i>Chrotomys silaceus</i>	Muridae	Rodentia	Asia
"	<i>Carpomys phaeurus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides chrotomys</i> Bochkov et OConnor, 2006	<i>Chrotomys gonzalesi</i>	Muridae	Rodentia	Asia
"	<i>Chrotomys mindorensis</i>	Muridae	Rodentia	Asia
"	<i>Bullimus luzonicus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides malayi</i> Fain, 1970	<i>Maxomys alticola</i>	Muridae	Rodentia	Asia
"	<i>Maxomys ochraceiventer</i>	Muridae	Rodentia	Asia
"	<i>Maxomys baeodon</i>	Muridae	Rodentia	Asia
<i>Listrophoroides alticola</i> Fain, 1976	<i>Maxomys alticola</i>	Muridae	Rodentia	Asia
"	<i>Maxomys ochraceiventer</i>	Muridae	Rodentia	Asia
"	<i>Maxomys baeodon</i>	Muridae	Rodentia	Asia
<i>Listrophoroides hipposideros</i> Fain, 1979	<i>Maxomys ochraceiventer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides forcicatus</i> Fain, 1974	<i>Maxomys alticola</i>	Muridae	Rodentia	Asia
"	<i>Maxomys ochraceiventer</i>	Muridae	Rodentia	Asia
"	<i>Maxomys baeodon</i>	Muridae	Rodentia	Asia
<i>Listrophoroides kinobaluensis</i> Fain, 1976	<i>Maxomys whiteheadi</i>	Muridae	Rodentia	Asia
<i>Listrophoroides inas</i> Bochkov et OConnor, 2005	<i>Maxomys inas</i>	Muridae	Rodentia	Asia
<i>Listrophoroides musschenbroekii</i> Bochkov et OConnor, 2005	<i>Maxomys musschenbroekii</i>	Muridae	Rodentia	Asia
<i>Listrophoroides maculatissimus</i> Fain, 1979	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
<i>Listrophoroides biexcavatus</i> Fain, 1979	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
<i>Listrophoroides cocoensis</i> Fain, 1976	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
<i>Listrophoroides moi</i> Bochkov et OConnor, 2005	<i>Maxomys moi</i>	Muridae	Rodentia	Asia
<i>Listrophoroides palawanensis</i> Fain, 1976	<i>Maxomys panglima</i>	Muridae	Rodentia	Asia
<i>Listrophoroides inflatus</i> Bochkov et OConnor, 2005	<i>Maxomys inflatus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides hemistriatus</i> Fain, 1976	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
"	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
<i>Listrophoroides bifidus</i> Fain, 1976	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
"	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides rajah</i> Fain, 1974	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
"	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides toxophallus</i> Fain, 1976	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
"	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides femoratus</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia

A. V. Bochkov

<i>Listrophoroides femoratus</i> Bochkov et OConnor, 2005	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
<i>Listrophoroides eudrilus</i> Fain, 1976	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
“	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
<i>Listrophoroides neobifidus</i> Fain, 1976	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
“	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
<i>Listrophoroides ancophallus</i> Fain, 1976	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
“	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides stenophallus</i> Fain, 1981	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
“	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
<i>Listrophoroides iphiophallus</i> Fain, 1976	<i>Maxomys rajah</i>	Muridae	Rodentia	Asia
“	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides birmanicus</i> Fain, 1976	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides bellulus</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides cambodiensis</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides khmeri</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides maxomys</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides megaphallus</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides helenae</i> Bochkov et Mironov, 2001	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides parafemoratus</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides musseri</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides pseudorajah</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides gorogae</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides surifer</i> Bochkov et OConnor, 2005	<i>Maxomys surifer</i>	Muridae	Rodentia	Asia
<i>Listrophoroides (Paklistrophoroides)</i> Fain et Hyland, 1980				
<i>Listrophoroides legadilla</i> Radford, 1947	<i>Mus fernandoni</i>	Muridae	Rodentia	Asia
<i>Listrophoroides orbitalis</i> Fain, 1976	<i>Mus fernandoni</i>	Muridae	Rodentia	Asia
<i>Listrophoroides decoratus</i> Fain et Hyland, 1980	<i>Meriones hurrianae</i>	Muridae	Rodentia	Asia
<i>Listrophoroides scutalis</i> Fain, 1979	<i>Leopoldamys edwardsi</i>	Muridae	Rodentia	Asia
<i>Listrophoroides (Marquesania)</i> Womersley, 1943				
<i>Listrophoroides cucullatus</i> (Trouessart, 1893)	<i>Rattus norvegicus</i>	Muridae	Rodentia	Cosmopolite (Tropical and Subtropical Regions)
“	<i>Rattus rattus</i>	Muridae	Rodentia	Cosmopolite (Tropical and Subtropical Regions)

A review of mammal-associated Psoroptidia

<i>Listrophoroides cucullatus</i> (Trouessart, 1893)	<i>Rattus tanezumi</i>	Muridae	Rodentia	Cosmopolite (Tropical Regions)
“	<i>Rattus annandalei</i>	Muridae	Rodentia	Asia
“	<i>Rattus argentiventer</i>	Muridae	Rodentia	Asia
“	<i>Rattus hoffmanni</i>	Muridae	Rodentia	Asia
“	<i>Rattus tiomanicus</i>	Muridae	Rodentia	Asia
“	<i>Rattus xanthurus</i>	Muridae	Rodentia	Asia
“	<i>Bunomys coelestis</i>	Muridae	Rodentia	Asia
“	<i>Stenomys niobe</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Listrophoroides papuanus</i> Fain, 1970	<i>Conilurus penicillatus</i>	Muridae	Rodentia	Australia
“	<i>Pseudomys nanus</i>	Muridae	Rodentia	Australia
“	<i>Rattus leucopus</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Listrophoroides obliquelineatus</i> Fain, 1975	<i>Rattus leucopus</i>	Muridae	Rodentia	Australia, including New Guinea
<i>Listrophoroides mordax</i> Bochkov et Fain, 2003	<i>Rattus mordax</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Rattus leucopus</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Listrophoroides crenatus</i> Fain, 1975	<i>Melomys rufescens</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Paramelomys moncktoni</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Rattus nitidus</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Listrophoroides interpolatus</i> Fain, 1975	<i>Stenomys niobe</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Rattus verecundus</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Paramelomys moncktoni</i>	Muridae	Rodentia	Australia (New Guinea)
“	<i>Maxomys hellwaldii</i>	Muridae	Rodentia	Asia
“	<i>Maxomys dollmani</i>	Muridae	Rodentia	Asia
“	<i>Maxomys</i> sp. A	Muridae	Rodentia	Asia
“	<i>Maxomys</i> sp. B	Muridae	Rodentia	Asia
<i>Listrophoroides melomys</i> Bochkov et Fain, 2003	<i>Paramelomys moncktoni</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Listrophoroides queenslandicus</i> (Womersley, 1943)	<i>Rattus sordidus</i>	Muridae	Rodentia	Australia
“	<i>Rattus fuscipes</i>	Muridae	Rodentia	Australia
“	<i>Rattus villosissimus</i>	Muridae	Rodentia	Australia
“	<i>Rattus tunneyi</i>	Muridae	Rodentia	Australia
“	<i>Rattus leucopus</i>	Muridae	Rodentia	Australia (New Guinea)
<i>Listrophoroides australiae</i> Fain, 1970	<i>Rattus lutreolus</i>	Muridae	Rodentia	Australia
“	<i>Rattus fuscipes</i>	Muridae	Rodentia	Australia
<i>Listrophoroides postsquamatus</i> Fain, 1976	<i>Rattus everetti</i>	Muridae	Rodentia	Asia
“	<i>Rattus</i> sp. A.	Muridae	Rodentia	Asia
“	<i>Apomys</i> sp. D.	Muridae	Rodentia	Asia

A. V. Bochkov

<i>Listrophoroides postsquamatus</i> Fain, 1976	<i>Rattus hoffmanni</i>	Muridae	Rodentia	Asia
“	<i>Maxomys hellwaldii</i>	Muridae	Rodentia	Asia
<i>Listrophoroides lativentris</i> Fain, 1981	<i>Sundamys muelleri</i>	Muridae	Rodentia	Asia
<i>Listrophoroides sculpturatus</i> Fain, Nadchatram et Lukoschus, 1981	<i>Sundamys muelleri</i>	Muridae	Rodentia	Asia
<i>Listrophoroides dominator</i> Fain, 1981	<i>Paruromys dominator</i>	Muridae	Rodentia	Asia
“	<i>Rattus xanthurus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides echiothrix</i> Bochkov, OConnor et Gorog, 2005	<i>Echiothrix centrosa</i>	Muridae	Rodentia	Asia
<i>Listrophoroides lenothrix</i> Bochkov, OConnor et Gorog, 2005	<i>Lenothrix canus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides pseudolativentris</i> Bochkov, OConnor et Gorog, 2005	<i>Lenothrix canus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides paracucullatus</i> Bochkov et OConnor, 2006	<i>Rattus exulans</i>	Muridae	Rodentia	Asia
“	<i>Apomys</i> sp. D	Muridae	Rodentia	Asia
<i>Listrophoroides postsquamatus</i> Bochkov et OConnor, 2006	<i>Rattus everetti</i>	Muridae	Rodentia	Asia
“	<i>Rattus</i> sp. A	Muridae	Rodentia	Asia
“	<i>Apomys</i> sp. D	Muridae	Rodentia	Asia
<i>Listrophoroides apomys</i> Bochkov et OConnor, 2006	<i>Apomys datae</i>	Muridae	Rodentia	Asia
“	<i>Apomys microdon</i>	Muridae	Rodentia	Asia
“	<i>Apomys musculus</i>	Muridae	Rodentia	Asia
“	<i>Apomys</i> sp. B	Muridae	Rodentia	Asia
“	<i>Apomys</i> sp. C	Muridae	Rodentia	Asia
<i>Listrophoroides heaneyi</i> Bochkov et OConnor, 2006	<i>Apomys hylocetes</i>	Muridae	Rodentia	Asia
“	<i>Apomys insignis</i>	Muridae	Rodentia	Asia
“	<i>Apomys microdon</i>	Muridae	Rodentia	Asia
<i>Listrophoroides faini</i> Bochkov et OConnor, 2006	<i>Apomys datae</i>	Muridae	Rodentia	Asia
“	<i>Apomys</i> sp.	Muridae	Rodentia	Asia
“	<i>Carpomys phaeurus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides limnomys</i> Bochkov et OConnor, 2006	<i>Limnomys bryophilus</i>	Muridae	Rodentia	Asia
“	<i>Tarsomys apoensis</i>	Muridae	Rodentia	Asia
<i>Listrophoroides bullimus</i> Bochkov et OConnor, 2006	<i>Bullimus gamay</i>	Muridae	Rodentia	Asia
<i>Listrophoroides luzonicus</i> Bochkov et OConnor, 2006	<i>Bullimus luzonicus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides corpuzrarosae</i> Bochkov et OConnor, 2006	<i>Bullimus bagobus</i>	Muridae	Rodentia	Asia
<i>Listrophoroides (Olistrophoroides)</i> Fain, 1972				
<i>Listrophoroides lemniscomys</i> Radford, 1940	<i>Lemniscomys striatus</i>	Muridae	Rodentia	Africa
“	<i>Lemniscomys rosalia</i>	Muridae	Rodentia	Africa
“	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
“	<i>Mus minutoides</i>	Muridae	Rodentia	Africa
“	<i>Lophuromys aquilus</i>	Muridae	Rodentia	Africa

A review of mammal-associated Psoroptidia

<i>Listrophoroides womersleyi</i> Lawrence, 1951	<i>Myotomys sloggetti</i>	Muridae	Rodentia	Africa
“	<i>Otomys irroratus</i>	Muridae	Rodentia	Africa
“	<i>Otomys tropicalis</i>	Muridae	Rodentia	Africa
<i>Listrophoroides otomys striatus</i> Fain, 1972	<i>Otomys</i> sp.	Muridae	Rodentia	Africa
<i>Listrophoroides caudatus</i> Fain, 1970	<i>Lemniscomys striatus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides pelomys</i> Fain, 1972	<i>Pelomys fallax</i>	Muridae	Rodentia	Africa
<i>Listrophoroides yaoundeensis</i> Fain, 1972	<i>Hybomys univittatus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides hybomys</i> Fain, 1970	<i>Hybomys univittatus</i>	Muridae	Rodentia	Africa
“	<i>Deomys ferrugineus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides hybomys scalaris</i> Fain, 1972	<i>Hybomys univittatus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides grammomys</i> Fain, 1970	<i>Grammomys dolichurus</i>	Muridae	Rodentia	Africa
“	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
“	<i>Oenomys hypoxanthus</i>	Muridae	Rodentia	Africa
“	<i>Gerbilliscus dichrura</i>	Muridae	Rodentia	Africa
<i>Listrophoroides auricularis</i> Fain, 1970	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
<i>Listrophoroides deomys</i> Fain, 1970	<i>Deomys ferrugineus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides africanus</i> Radford, 1944	<i>Mastomys erythroleucus</i>	Muridae	Rodentia	Africa
“	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Pelomys minor</i>	Muridae	Rodentia	Africa
<i>Listrophoroides africanus praomys</i> Fain, 1972	<i>Praomys tullbergi</i>	Muridae	Rodentia	Africa
“	<i>Praomys morio</i>	Muridae	Rodentia	Africa
<i>Listrophoroides scutatus</i> Fain, 1970	<i>Hylomyscus alleni</i>	Muridae	Rodentia	Africa
“	<i>Uranomys ruddi</i>	Muridae	Rodentia	Africa
“	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
<i>Listrophoroides dephomys</i> Fain, 1970	<i>Dephomys defua</i>	Muridae	Rodentia	Africa
<i>Listrophoroides punctulatus</i> Fain, 1970	<i>Colomys goslingi</i>	Muridae	Rodentia	Africa
<i>Listrophoroides stochomys</i> Fain, 1972	<i>Stochomys longicaudatus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides camerounensis</i> Fain, 1972	<i>Hybomys univittatus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides quadratus</i> Fain, 1972	<i>Stochomys longicaudatus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides conifer</i> Fain, 1972	<i>Stochomys longicaudatus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides kivuanus</i> Fain, Hart et rahm, 1986	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
<i>Listrophoroides irangiensis</i> Fain, Hart et rahm, 1986	<i>Hybomys univittatus</i>	Muridae	Rodentia	Africa
“	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
<i>Listrophoroides thallomys</i> Fain et Bochkov, 2003	<i>Thallomys paedulcus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides oenomiphilus</i> Fain et Bochkov, 2003	<i>Oenomys hypoxanthus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides (Afrolistrophoroides)</i> Fain, 1972				
<i>Listrophoroides mastomys</i> Radford, 1940	<i>Mastomys erythroleucus</i>	Muridae	Rodentia	Africa

<i>Listrophoroides mastomys</i> Radford, 1940	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Thallomys paeulculus</i>	Muridae	Rodentia	Africa
“	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
“	<i>Lemniscomys striatus</i>	Muridae	Rodentia	Africa
“	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
“	<i>Mus minutoides</i>	Muridae	Rodentia	Africa
“	<i>Oenomys hypoxanthus</i>	Muridae	Rodentia	Africa
“	<i>Pelomys minor</i>	Muridae	Rodentia	Africa
“	<i>Lophuromys aquilus</i>	Muridae	Rodentia	Africa
“	<i>Gerbilliscus leucogaster</i>	Muridae	Rodentia	Africa
<i>Listrophoroides teinophallus</i> Fain, 1970	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Pelomys minor</i>	Muridae	Rodentia	Africa
<i>Listrophoroides radfordi</i> Fain, 1970	<i>Praomys tullbergi</i>	Muridae	Rodentia	Africa
<i>Listrophoroides scambophallus</i> Fain, 1970	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Praomys tullbergi</i>	Muridae	Rodentia	Africa
“	<i>Otomys</i> sp.	Muridae	Rodentia	Africa
<i>Listrophoroides treptophallus</i> Fain, 1970	<i>Dephomyys defua</i>	Muridae	Rodentia	Africa
<i>Listrophoroides hylomyscus</i> Fain, 1970	<i>Hylomyscus alleni</i>	Muridae	Rodentia	Africa
<i>Listrophoroides leggada</i> Fain, 1970	<i>Mus musculoides</i>	Muridae	Rodentia	Africa
<i>Listrophoroides benoiti</i> Fain, 1970	<i>Colomys goslingi</i>	Muridae	Rodentia	Africa
<i>Listrophoroides gaudi</i> Fain, 1972	<i>Praomys tullbergi</i>	Muridae	Rodentia	Africa
<i>Listrophoroides vittatus</i> Fain, 1970	<i>Hybomys univittatus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides dubininae</i> Fain, 1972	<i>Dendromys melanotis</i>	Nesomyidae	Rodentia	Africa
“	<i>Dendromys mesomelas</i>	Nesomyidae	Rodentia	Africa
<i>Listrophoroides malacomys</i> Fain, 1970	<i>Malacomys edwardsi</i>	Muridae	Rodentia	Africa
<i>Listrophoroides notatus</i> Fain, 1970	<i>Malacomys edwardsi</i>	Muridae	Rodentia	Africa
<i>Listrophoroides leptolobus</i> Fain, 1972	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
<i>Listrophoroides bilobatus</i> Fain, 1970	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
<i>Listrophoroides serratus</i> Fain, 1970	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
<i>Listrophoroides obliquistriatus</i> Fain, 1970	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
<i>Listrophoroides furcatus</i> Fain, 1970	Muridae sp.	Muridae	Rodentia	Africa
<i>Listrophoroides machadoi</i> Fain, 1972	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
“	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Lophuromys aquilus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides angolensis</i> Fain, 1972	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
<i>Listrophoroides sigmophallus</i> Fain, 1972	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
<i>Listrophoroides attenuatus</i> Fain, 1972	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
“	<i>Lophuromys aquilus</i>		Rodentia	Africa
<i>Listrophoroides carrissoensis</i> Fain, 1972	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
<i>Listrophoroides verruculosus</i> Fain, Hart et Rahm, 1986	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa

A review of mammal-associated Psoroptidia

<i>Listrophoroides verruculosus</i> Fain, Hart et Rahm, 1986	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
<i>Listrophoroides myomyscus</i> Fain et Bochkov, 2003	<i>Praomys daltoni</i>	Muridae	Rodentia	Africa
<i>Listrophoroides colomys</i> Fain et Bochkov, 2003	<i>Colomys goslingi</i>	Muridae	Rodentia	Africa
<i>Listrophoroides paraleggada</i> Fain et Bochkov, 2003	<i>Mus musculoides</i>	Muridae	Rodentia	Africa
<i>Listrophoroides prionomys</i> Fain et Bochkov, 2004	<i>Prionomys batesi</i>	Muridae	Rodentia	Africa
<i>Listrophoroides (Arboricolichus)</i> Fain, 1972				
<i>Listrophoroides adherens</i> (Trouessart, 1893)	<i>Anomalurus derbianus</i>	Anomaluridae	Rodentia	Africa
<i>Listrophoroides anomaluri</i> Fain, 1972	<i>Anomalurus derbianus</i>	Anomaluridae	Rodentia	Africa
<i>Listrophoroides idiuri</i> Fain, 1972	<i>Idiurus macrotis</i>	Anomaluridae	Rodentia	Africa
<i>Listrophoroides verheyeni</i> Fain, 1970	<i>Heliosciurus rufobrachium</i>	Sciuridae	Rodentia	Africa
<i>Listrophoroides funisciuri</i> Fain, 1970	<i>Funisciurus</i> sp.	Sciuridae	Rodentia	Africa
<i>Listrophoroides monticola</i> Fain, Hart et Rahm, 1986	<i>Funisciurus carruthesi</i>	Sciuridae	Rodentia	Africa
<i>Listrophoroides graphiuri</i> Fain, 1970	<i>Graphiurus nagtglasii</i>	Gliridae	Rodentia	Africa
<i>Listrophoroides claviglis</i> Fain, 1970	<i>Graphiurus murinus</i>	Gliridae	Rodentia	Africa
<i>Listrophoroides schoutedeni</i> Fain, 1967	<i>Galago demidoff</i>	Galagidae	Primates	Africa
“	<i>Galago demidoff</i>	Galagidae	Primates	Africa
“	<i>Galago demidoff</i>	Galagidae	Primates	Africa
<i>Listrophoroides (Colistrophoroides)</i> Fain, 1972				
<i>Listrophoroides uromomys</i> Fain, 1970	<i>Uromomys ruddi</i>	Muridae	Rodentia	Africa
“	<i>Acomys johannis</i>	Muridae	Rodentia	
<i>Listrophoroides (Crilistrophoroides)</i> Fain, 1972				
<i>Listrophoroides cricetomys</i> Fain, 1970	<i>Cricetomys emini</i>	Nesomyidae	Rodentia	Africa
<i>Listrophoroides (Pedetochirus)</i> Fain, 1972				
<i>Listrophoroides pedetochirus</i> Fain, 1972	<i>Pedetes capensis</i>	Pedetidae	Rodentia	Africa
<i>Listrophoroides (Macroscelistrophoroides)</i> Fain et Bochkov, 2003				
<i>Listrophoroides petrodromi</i> Fain et Bochkov, 2003	<i>Petrodromus tetradactylus</i>	Macroscelididae	Macroscelidea	Africa
<i>Listrophoroides (Alistrophoroides)</i> Fain, 1972				
<i>Listrophoroides mediosquamatus</i> Fain, 1970	<i>Dephomys defua</i>	Muridae	Rodentia	Africa
“	<i>Hybomys univittatus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides pachyphallus</i> Fain, 1970	<i>Hybomys univittatus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides hulstaerti</i> Fain, 1970	<i>Oenomys hypoxanthus</i>	Muridae	Rodentia	Africa
<i>Listrophoroides aethomys</i> Fain et Bochkov, 2003	<i>Aethomys bocagei</i>	Muridae	Rodentia	Africa
<i>Listrophoroides marginatus</i> Fain, 1970	<i>Microgale cowani</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
“	<i>Microgale talazaci</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
“	<i>Microgale pusilla</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
“	<i>Microgale thomasi</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)

<i>Listrophoroides marginatus</i> Fain, 1970	<i>Gymnuromys roberti</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides uilenbergi</i> Fain, 1970	<i>Microgale</i> sp.	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides scleropygus</i> Fain, 1970	<i>Microgale</i> sp.	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides nesogale</i> Fain, 1970	<i>Microgale dobsoni</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides pusillus</i> Fain et Lukoschus, 1976	<i>Microgale pusilla</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides papillosus</i> Fain et Lukoschus, 1976	<i>Microgale longicaudata</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides sikorai</i> Fain et Lukoschus, 1976	<i>Microgale thomasi</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides (Eulistrophoroides)</i> Fain, 1976				
<i>Listrophoroides paulianus</i> Fain, 1976	<i>Brachyuromys betsileoensis</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides minutus</i> Fain, 1976	<i>Eliurus penicillatus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides legendrei</i> Fain, 1976	<i>Macrotarsomys bastardi</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides tenuis</i> Fain, 1976	<i>Macrotarsomys bastardi</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides (Pallistrophoroides)</i> Fain, 1976				
<i>Listrophoroides scapulatus</i> Fain, 1970	<i>Eliurus penicillatus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
“	<i>Nesomys rufus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides coxatus</i> Fain, 1970	<i>Eliurus penicillatus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
“	<i>Macrotarsomys bastardi</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides sigmoidus</i> Fain, 1976	<i>Macrotarsomys bastardi</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides similis</i> Fain, 1970	<i>Macrotarsomys bastardi</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides arciferus</i> Fain, 1970	<i>Macrotarsomys bastardi</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides crassipes</i> Fain, 1976	<i>Eliurus penicillatus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides annulatus</i> Fain, 1970	<i>Eliurus penicillatus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides eliuri</i> Fain, 1970	<i>Eliurus penicillatus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides subquadratus</i> Fain, 1976	<i>Eliurus penicillatus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides tarsatus</i> Fain, 1976	<i>Gymnuromys roberti</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides fuitivus</i> Fain, 1976	<i>Gymnuromys roberti</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides gymnuromys</i> Fain, 1976	<i>Gymnuromys roberti</i>	Nesomyidae	Rodentia	Africa (Madagascar)
“	<i>Eliurus penicillatus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides ambiguous</i> Fain, 1970	<i>Nesomys rufus</i>	Nesomyidae	Rodentia	Africa (Madagascar)

A review of mammal-associated Psoroptidia

<i>Listrophoroides nesomys</i> Fain, 1976	<i>Nesomys rufus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides hypogeomys</i> Fain, 1970	<i>Hypogeomys antimena</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides madagascariensis</i> Fain, 1970	<i>Brachyuromys betsileoensis</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides callipygus</i> Fain et Lukoschus, 1976	<i>Brachyuromys betsileoensis</i>	Nesomyidae	Rodentia	Africa (Madagascar)
<i>Listrophoroides (Madlistrophoroides)</i> Fain, 1972				
<i>Listrophoroides fortis</i> Fain, 1972	! <i>Brachyuromys betsileoensis</i>	Nesomyidae	Rodentia	Africa (Madagascar)
“	! <i>Nesomys rufus</i>	Nesomyidae	Rodentia	Africa (Madagascar)
“	<i>Oryzorictes tetradactylus</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
“	<i>Microgale cowani</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides microgale</i> Fain, 1970	<i>Microgale</i> sp.	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides lukoschusi</i> Fain, 1976	<i>Microgale cowani</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
“	<i>Microgale owani</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides imerinensis</i> Fain et Lukoschus, 1976	<i>Microgale thomasi</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides uniformis</i> Fain et Lukoschus, 1976	<i>Oryzorictes hova</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
“	<i>Microgale cowani</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
“	<i>Microgale longicaudata</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
“	<i>Limnogale mergulus</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides calcaratus</i> Fain et Lukoschus, 1976	<i>Microgale longicaudata</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides (Bilistrophoroides)</i> Fain, 1976				
<i>Listrophoroides curtus</i> Fain et Lukoschus, 1976	<i>Limnogale mergulus</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Listrophoroides (Lemurlistrophoroides)</i> Fain, 1972				
<i>Listrophoroides pauliani</i> Lawrence, 1959	<i>Lepilemur ruficaudatus</i>	Lepilemuridae	Primates	Africa (Madagascar)
<i>Listrophoroides lawrencei</i> Fain, 1970	<i>Lepilemur ruficaudatus</i>	Lepilemuridae	Primates	Africa (Madagascar)
<i>Listrophoroides dauphinensis</i> Fain, 1970	<i>Lepilemur ruficaudatus</i>	Lepilemuridae	Primates	Africa (Madagascar)
“	<i>Hapalemur griseus</i>	Lemuridae	Primates	Africa (Madagascar)
“	<i>Avahi laniger</i>	Indriidae	Primates	Africa (Madagascar)
<i>Listrophoroides squamosus</i> Lawrence, 1959	<i>Lepilemur mustelinus</i>	Lepilemuridae	Primates	Africa (Madagascar)
<i>Listrophoroides inopinatus</i> Fain, 1976	<i>Lepilemur mustelinus</i>	Lepilemuridae	Primates	Africa (Madagascar)
<i>Listrophoroides lepilemur</i> Fain et Bochkov, 2003	<i>Lepilemur leucopus</i>	Lepilemuridae	Primates	Africa (Madagascar)

<i>Listrophoroides trilineatus</i> Fain, 1970	<i>Eulemur coronatus</i>	Lemuridae	Primates	Africa (Madagascar)
<i>Listrophoroides gracilis</i> Fain, 1970	<i>Cheirogaleus</i> sp.	Gheirogaleidae	Primates	Africa (Madagascar)
<i>Listrophoroides mediostrictus</i> Fain, 1970	<i>Cheirogaleus</i> sp.	Gheirogaleidae	Primates	Africa (Madagascar)
<i>Listrophoroides cheirogaleus</i> Fain, 1970	<i>Cheirogaleus</i> sp.	Gheirogaleidae	Primates	Africa (Madagascar)
<i>Listrophoroides tenullus</i> Fain, 1970	<i>Cheirogaleus</i> sp.	Gheirogaleidae	Primates	Africa (Madagascar)
<i>Listrophoroides eupleres</i> Fain, 1970	* <i>Eupleres goudotii</i>	Eupleridae	Carnivora	Africa (Madagascar)
“	<i>Cheirogaleus</i> sp.	Gheirogaleidae	Primates	Africa (Madagascar)
<i>Listrophoroides angulatus</i> Fain, 1970	<i>Eupleres goudotii</i>	Eupleridae	Carnivora	Africa (Madagascar)
<i>Listrophoroides (Belistrophoroides)</i> Fain, 1976				
<i>Listrophoroides amplius</i> Fain, 1970	<i>Eulemur coronatus</i>	Lemuridae	Primates	Africa (Madagascar)
<i>Centetesia</i> Lawrence, 1966				
<i>Centetesia tiptoni</i> Lawrence, 1955	<i>Hemicentetes semispinosus</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
“	<i>Hemicentetes nigriceps</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Centetesia tesellata</i> Lawrence, 1955	<i>Hemicentetes semispinosus</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Tenrecobia</i> Lawrence, 1955				
<i>Tenrecobia pauliana</i> Lawrence, 1955	<i>Setifer setosus</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
“	<i>Echinops telfairi</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)
<i>Tenrecobia tenrec</i> Fain, 1970	<i>Tenrec ecaudatus</i>	Tenrecidae	Afrosoricida	Africa (Madagascar)

Table 9. Host distribution of the family Chirodiscidae Trouessart, 1892

Mite species	Host species	Host family	Host order	Distribution
Chirodiscinae Trouessart, 1892				
<i>Chirodiscus</i> Trouessart et Neumann, 1889				
<i>Chirodiscus amplexans</i> Trouessart et Neumann, 1889	Unknown marsupial host			Australia
Labidocarpinae Gunther, 1942				
Labidocarpini Fain, 1971				
<i>Adentocarpus</i> Fain, 1972				
<i>Adentocarpus pipistrelli</i> (Fain, 1970)	<i>Hypsugo crassulus</i>	Vespertilionidae	Chiroptera	Africa
<i>Afrolabidocarpus</i> Fain, 1970				
<i>Afrolabidocarpus longiscutatus</i> Fain, 1970	<i>Hipposideros ruber</i>	Hipposideridae	Chiroptera	Africa
“	<i>Coleura afra</i>	Hipposideridae	Chiroptera	Africa
“	<i>Hipposideros</i> sp.	Hipposideridae	Chiroptera	Australia (New Guinea)
“	<i>Hipposideros ater</i>	Hipposideridae	Chiroptera	Australia (New Guinea)
<i>Afrolabidocarpus breviscutatus</i> Fain, 1970	<i>Hipposideros ruber</i>	Hipposideridae	Chiroptera	Africa

A review of mammal-associated Psoroptidia

<i>Afrolabidocarpus medioscutatus</i> Fain, 1972	<i>Hipposideros</i> sp.	Hipposideridae	Chiroptera	Australia (New Guinea)
<i>Afrolabidocarpus longipis</i> Fain, 1980	<i>Hipposideros galeritus</i>	Hipposideridae	Chiroptera	Asia
<i>Afrolabidocarpus vietnamensis</i> Fain, 1976	<i>Hipposideros pratti</i>	Hipposideridae	Chiroptera	Asia
<i>Alabidocarpus</i> Ewing, 1929				
<i>Alabidocarpus megalonyx</i> Trouessart, 1895	<i>Rhinolophus ferrumequinum</i>	Rhinolophidae	Chiroptera	Europe
“	<i>Rhinolophus zuluensis</i>	Rhinolophidae	Chiroptera	Africa
“	<i>Rhinolophus hildebrandtii</i>	Rhinolophidae	Chiroptera	Africa
“	<i>Rhinolophus fumigatus</i>	Rhinolophidae	Chiroptera	Africa
<i>Alabidocarpus diceratops</i> Lawrence, 1959	<i>Rhinolophus capensis</i>	Rhinolophidae	Chiroptera	Africa
“	<i>Rhinolophus euryale</i>	Rhinolophidae	Chiroptera	Europe
<i>Alabidocarpus vietnamensis</i> Fain, 1976	<i>Rhinolophus macrotis</i>	Rhinolophidae	Chiroptera	Asia
“	<i>Rhinolophus arcuatus</i>	Rhinolophidae	Chiroptera	Asia
“	<i>Rhinolophus shameli</i>	Rhinolophidae	Chiroptera	Asia
<i>Alabidocarpus scutellatus</i> Fain, 1982	<i>Rhinolophus creaghi</i>	Rhinolophidae	Chiroptera	Asia
<i>Alabidocarpus minor</i> (Rollinat et Trouessart, 1897)	<i>Rhinolophus ferrumequinum</i>	Rhinolophidae	Chiroptera	Europe
“	<i>Rhinolophus hipposideros</i>	Rhinolophidae	Chiroptera	Europe
“	<i>Rhinolophus blasii</i>	Rhinolophidae	Chiroptera	Europe
<i>Alabidocarpus yandinae</i> Domrow et Moorhouse, 1975	<i>Rhinolophus megaphyllus</i>	Rhinolophidae	Chiroptera	Australia
<i>Alabidocarpus spinatarsus</i> Fain, 1971	<i>Hipposideros ruber</i>	Hipposideridae	Chiroptera	Africa
<i>Alabidocarpus lankadivae</i> Fain, 1976	<i>Hipposideros lankadiva</i>	Hipposideridae	Chiroptera	Asia
<i>Alabidocarpus laoensis</i> Fain, 1976	<i>Hipposideros</i> sp.	Hipposideridae	Chiroptera	Asia
<i>Alabidocarpus calcaratus</i> Lawrence, 1952	<i>Myotis tricolor</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Myotis blythii</i>	Vespertilionidae	Chiroptera	Eurasia
“	<i>Myotis oxygnathus</i>	Vespertilionidae	Chiroptera	Eurasia
“	<i>Myotis emarginatus</i>	Vespertilionidae	Chiroptera	Europe
<i>Alabidocarpus longipilus</i> Pinichpongse, 1963	<i>Myotis yumanensis</i>	Vespertilionidae	Chiroptera	North America
“	<i>Myotis californicus</i>	Vespertilionidae	Chiroptera	North America
“	<i>Myotis volans</i>	Vespertilionidae	Chiroptera	North America
“	<i>Myotis nigricans</i>	Vespertilionidae	Chiroptera	South America
<i>Alabidocarpus intercalatus</i> Fain, 1971	<i>Myotis myotis</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis daubentonii</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis emarginatus</i>	Vespertilionidae	Chiroptera	Asia
<i>Alabidocarpus miniopteri</i> Fain, 1971	<i>Miniopterus schreibersii</i>	Vespertilionidae	Chiroptera	Africa
<i>Alabidocarpus recurvus</i> (Womersley, 1943)	Chiroptera sp.		Chiroptera	Australia
“	<i>Nyctophilus geoffroyi</i>	Vespertilionidae	Chiroptera	Australia
“	<i>Chaerephon jobensis</i>	Molossidae	Chiroptera	Australia
“	<i>Scotorepens greyi</i>		Chiroptera	Australia

<i>Alabidocarpus recurvus</i> (Womersley, 1943)	<i>Vespadeus douglasorum</i>	Vespertilionidae	Chiroptera	Australia
“	<i>Vespadeus pumilus</i>	Vespertilionidae	Chiroptera	Australia
<i>Alabidocarpus chalinolobi</i> Fain et Lukoschus, 1981	<i>Chalinolobus gouldii</i>	Vespertilionidae	Chiroptera	Australia
<i>Alabidocarpus parvulus</i> Fain et Lukoschus, 1981	<i>Vespadeus douglasorum</i>	Vespertilionidae	Chiroptera	Australia
“	<i>Eptesicus pumilus</i>	Vespertilionidae	Chiroptera	Australia
<i>Alabidocarpus nodulosus</i> Fain, 1979	<i>Miniopterus paululus</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Miniopterus australis</i>	Vespertilionidae	Chiroptera	Asia
<i>Alabidocarpus fudjii</i> Wada, 1967	<i>Miniopterus schreibersii</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Miniopticrus australis</i>	Vespertilionidae	Chiroptera	Australia (New Guinea)
<i>Alabidocarpus eptesicus</i> Fain, 1970	<i>Neoromicia tenuipinnis</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Eptesicus nilssonii</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Kerivoula cuprosa</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Neoromicia nanus</i>	Vespertilionidae	Chiroptera	Africa
<i>Alabidocarpus nyctali</i> Fain, 1971	<i>Nyctalus noctula</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Plecotus auritus</i>	Vespertilionidae	Chiroptera	Europe
<i>Alabidocarpus kerivoula</i> Fain, 1971	<i>Kerivoula smithii</i>	Vespertilionidae	Chiroptera	Africa
<i>Alabidocarpus scotophilus</i> Fain, 1972	<i>Scotophilus kuhlii</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Scotophilus nigrita</i>	Vespertilionidae	Chiroptera	Asia
<i>Alabidocarpus parryae</i> Fain, 1976	<i>Myotis moluccarum</i>	Vespertilionidae	Chiroptera	Australia
<i>Alabidocarpus glauconycteris</i> Fain, 1976	<i>Glauconycteris argentata</i>	Vespertilionidae	Chiroptera	Africa
<i>Alabidocarpus nycteris</i> Fain, 1970	<i>Nycteris</i> sp.	Nycteridae	Chiroptera	Africa
“	<i>Nycteris grandis</i>	Nycteridae	Chiroptera	Africa
<i>Alabidocarpus kivuensis</i> Fain, 1971	<i>Nycteris hispida</i>	Nycteridae	Chiroptera	Africa
<i>Alabidocarpus molossicola</i> Fain, 1970	<i>Chaerephon ansorgei</i>	Molossidae	Chiroptera	Africa
<i>Alabidocarpus saccopteryx</i> Fain, 1970	<i>Saccopteryx bilineata</i>	Emballonuridae	Chiroptera	South America
<i>Alabidocarpus taphozous</i> Fain, 1971	<i>Taphozous perforatus</i>	Emballonuridae	Chiroptera	Africa
<i>Alabidocarpus laviae</i> Fain, 1976	<i>Lavia frons</i>	Megadermatidae	Chiroptera	Africa
<i>Alabidocarpus hilli</i> Fain, 1976	Megadermatidae sp.	Megadermatidae	Chiroptera	Africa
<i>Alabidocarpus octodens</i> Fain, 1972	<i>Megaderma spasma</i>	Megadermatidae	Chiroptera	Asia
<i>Alabidocarpus furmani</i> Pinichpongse, 1963	<i>Anoura geoffroyi</i>	Phyllostomidae	Chiroptera	South America
“	<i>Anoura caudifer</i>	Phyllostomidae	Chiroptera	South America
“	<i>Carollia perspicillata</i>	Phyllostomidae	Chiroptera	South America
“	<i>Carollia brevicauda</i>	Phyllostomidae	Chiroptera	South America
“	<i>Glossophaga longirostris</i>	Phyllostomidae	Chiroptera	South America
“	<i>Glossophaga soricina</i>	Phyllostomidae	Chiroptera	South America
“	<i>Pteronotus parnellii</i>	Phyllostomidae	Chiroptera	South America
“	<i>Platyrrhinus helleri</i>	Phyllostomidae	Chiroptera	South America
<i>Alabidocarpus jonesi</i> McDaniel, 1971	<i>Platyrrhinus helleri</i>	Phyllostomidae	Chiroptera	South America
<i>Alabidocarpus nicaraguae</i> McDaniel, 1971	<i>Uroderma bilobatum</i>	Phyllostomidae	Chiroptera	South America

A review of mammal-associated Psoroptidia

<i>Alabidocarpus nicaraguae</i> McDaniel, 1971	<i>Uroderma magnirostrum</i>	Phyllostomidae	Chiroptera	South America
<i>Alabidocarpus guyanensis</i> Fain, 1972	<i>Artibeus cinereus</i>	Phyllostomidae	Chiroptera	South America
<i>Alabidocarpus rousetti</i> Fain, 1970	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Eidolon helvum</i>	Pteropodidae	Chiroptera	Africa
“	<i>Dobsonia inermis</i>	Pteropodidae	Chiroptera	Asia
“	<i>Rousettus amplexicaudatus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
<i>Asiolabidocarpus</i> Fain, 1972				
<i>Asiolabidocarpus bougainvilleensis</i> Fain, 1972	<i>Hipposideros</i> sp.	Hipposideridae	Chiroptera	Australia (New Guinea)
“	<i>Hipposideros calcaratus</i>	Hipposideridae	Chiroptera	Asia, Australia
“	<i>Aselliscus tricuspoidatus</i>	Hipposideridae	Chiroptera	Asia, Australia
<i>Asiolabidocarpus hipposideros</i> Fain, 1979	<i>Hipposideros armiger</i>	Hipposideridae	Chiroptera	Asia
<i>Asiolabidocarpus megadermae</i> Fain, 1972	<i>Megaderma spasma</i>	Megadermatidae	Chiroptera	Asia
<i>Dentocarpus</i> Dusbabek et Cruz, 1966				
<i>Dentocarpus silvai</i> Dusbabek et Cruz, 1966	<i>Chaerephon major</i>	Molossidae	Chiroptera	South America
“	<i>Molossus molossus</i>	Molossidae	Chiroptera	South America
“	[<i>Molossus</i> or <i>Eumops</i>] <i>ater</i>	Molossidae	Chiroptera	South America
“	<i>Cynomops planirostris</i>	Molossidae	Chiroptera	South America
<i>Dentocarpus silvai eumopsicola</i> Fain, 1973	<i>Eumops auripendulus</i>	Molossidae	Chiroptera	South America
<i>Dentocarpus macrotrichus</i> Dusbabek et Cruz, 1966	<i>Tadarida brasiliensis</i>	Molossidae	Chiroptera	South America (Cuba)
<i>Dentocarpus aitkeni</i> (Pinichpongse, 1963)	<i>Chaerephon major</i>	Molossidae	Chiroptera	South America
“	<i>Molossus molossus</i>	Molossidae	Chiroptera	South America
“	[<i>Molossus</i> or <i>Eumops</i>] <i>ater</i>	Molossidae	Chiroptera	South America
<i>Dentocarpus chaerephon</i> Fain, 1970	<i>Chaerephon pumilus</i>	Molossidae	Chiroptera	Africa
“	<i>Chaerephon jobensis</i>	Molossidae	Chiroptera	Australia
<i>Dentocarpus eumops</i> Fain, 1972	<i>Eumops auripendulus</i>	Molossidae	Chiroptera	South America
<i>Dentocarpus orientalis</i> Fain, 1976	<i>Chaerephon johorensis</i>	Molossidae	Chiroptera	Asia
<i>Dentocarpus mimon</i> Fain, 1976	<i>Mimon bennettii</i>	Molossidae	Chiroptera	South America
<i>Dentocarpus medius</i> (Fain, 1970)	<i>Eptesicus brasiliensis</i>	Vespertilionidae	Chiroptera	South America
<i>Dentocarpus tenuis</i> (Fain, 1970)	<i>Coleura afra</i>	Emballonuridae	Chiroptera	Africa
“	<i>Myopterus whitleyi</i>	Molossidae	Chiroptera	Africa
“	<i>Taphozous</i> sp.	Emballonuridae	Chiroptera	Africa
“	<i>Chaerephon</i> sp.	Molossidae	Chiroptera	Africa
<i>Dentocarpus exiguus</i> Fain, 1971	<i>Coleura afra</i>	Emballonuridae	Chiroptera	Africa
<i>Dentocarpus lukoschusi</i> (Fain, 1970)	<i>Micronycteris megalotis</i>	Phyllostomidae	Chiroptera	South America
<i>Dentocarpus taphozous</i> Fain, 1972	<i>Saccolaimus saccolaimus</i>	Emballonuridae	Chiroptera	South America

<i>Dentocarpus notopterus</i> (Fain, 1976)	<i>Notopterus macdonaldi</i>	Pteropodidae	Chiroptera	Australia (New Hebrides)
<i>Dentocarpus notopterus</i> (Fain, 1976)	<i>Notopterus neocaledonica</i>	Pteropodidae	Chiroptera	Australia
<i>Dentocarpus borneoensis</i> Fain, 1979	<i>Rhinolophus creaghi</i>	Rhinolophidae	Chiroptera	Asia
“	<i>Pteropus admiralitatum</i>	Pteropodidae	Chiroptera	Australia
<i>Dentocarpus novaeguineae</i> (Fain, 1976)	<i>Nyctimene albiventer</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
“	<i>Nyctimene major</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
“	<i>Nyctimene aello</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
“	<i>Nyctimene draconilla</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
“	<i>Nyctimene [bougainvillei]</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
<i>Labidocarpellus</i> Fain, 1976				
<i>Labidocarpellus nyctimene</i> Fain, 1976	<i>Nyctimene albivente</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
<i>Labidocarpellus kimberleyensis</i> (Fain et Lukoschus, 1981)	<i>Vespadeus douglasorum</i>	Vespertilionidae	Chiroptera	Australia
<i>Labidocarpellus selangorensis</i> Fain, 1976	<i>Dyacopterus spadiceus</i>	Pteropodidae	Chiroptera	Asia
<i>Labidocarpellus phyllodermae</i> (Fain, 1976)	<i>Phylloderma stenops</i>	Phyllostomidae	Chiroptera	South America
<i>Labidocarpellus abyssinicus</i> (Fain, 1976)	<i>Otomops martiensseni</i>	Molossidae	Chiroptera	Africa
<i>Labidocarpellus chropterus</i> (Fain, 1976)	<i>Chropterus auritus</i>	Molossidae	Chiroptera	South America
<i>Labidocarpellus peropteryx</i> (Fain, 1972)	<i>Peropteryx kappleri</i>	Emballonuridae	Chiroptera	South America
<i>Labidocarpellus guyanensis</i> (Fain, 1973)	<i>Peropteryx kappleri</i>	Emballonuridae	Chiroptera	South America
<i>Labidocarpellus cynopterus</i> Fain, 1976	<i>Cynopterus brachyotis</i>	Pteropodidae	Chiroptera	Asia
<i>Labidocarpellus eonycteris</i> Fain, 1976	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
<i>Labidocarpellus dobsonia</i> (Fain, 1975)	<i>Dobsonia moluccensis</i>	Pteropodidae	Chiroptera	Australia (New Guinea)
<i>Eurolabidocarpus</i> Fain et Aellen, 1994				
<i>Eurolabidocarpus helveticus</i> Fain et Aellen, 1994	<i>Myotis daubentonii</i>	Vespertilionidae	Chiroptera	Europe
<i>Glossophagocarus</i> Cruz, 1973				
<i>Glossophagocarus cubanus</i> Cruz, 1973	<i>Monophyllus redmani</i>	Phyllostomidae	Chiroptera	South America
<i>Labidocarpoides</i> Fain, 1970				
<i>Labidocarpoides congoensis</i> (Fain, 1970)	<i>Rhinolophus hildebrandtii</i>	Rhinolophidae	Chiroptera	Africa
<i>Labidocarpoides guineae</i> Fain, 1971	<i>Rhinolophus landeri</i>	Rhinolophidae	Chiroptera	Africa
<i>Labidocarpoides grandior</i> Fain, 1979	<i>Rhinolophus luctus</i>	Rhinolophidae	Chiroptera	Asia
<i>Labidocarpoides uluensis</i> Fain, 1979	<i>Rhinolophus luctus</i>	Rhinolophidae	Chiroptera	Asia
<i>Labidocarpoides hipposideros</i> (Fain, 1970)	<i>Hipposideros caffer</i>	Hipposideridae	Chiroptera	Africa
“	<i>Hipposideros ruber</i>	Hipposideridae	Chiroptera	Africa

A review of mammal-associated Psoroptidia

<i>Labidocarpoides hipposideros ceylanicus</i> Fain, 1976	<i>Rhinolophus beddomei</i>	Rhinolophidae	Chiroptera	Asia
<i>Labidocarpus</i> Trouessart, 1895				
<i>Labidocarpus rollinatti</i> Trouessart, 1895	<i>Rhinolophus hipposideros</i>	Rhinolophidae	Chiroptera	Europe
“	<i>Rhinolophus ferrumequinum</i>	Rhinolophidae	Chiroptera	Europe
“	<i>Rhinolophus mehelyi</i>	Rhinolophidae	Chiroptera	Europe
“	<i>Rhinolophus clivus</i>	Rhinolophidae	Chiroptera	Africa
“	<i>Rhinolophus capensis</i>	Rhinolophidae	Chiroptera	Africa
<i>Labidocarpus minor</i> Trouessart, 1897	<i>Rhinolophus ferrumequinum</i>	Rhinolophidae	Chiroptera	Europe
<i>Labidocarpus selangorensis</i> Fain, 1979	<i>Rhinolophus luctus</i>	Rhinolophidae	Chiroptera	Asia
<i>Labidocarpus refulgens</i> Fain, 1979	<i>Rhinolophus lepidus</i>	Rhinolophidae	Chiroptera	Asia
“	<i>Rhinolophus euryotis</i>	Rhinolophidae	Chiroptera	Asia
<i>Labidocarpus rhinolophi</i> Fain, 1976	<i>Rhinolophus malayanus</i>	Rhinolophidae	Chiroptera	Asia
<i>Labidocarpus hongkongensis</i> Fain, 1982	<i>Rhinolophus pusillus</i>	Rhinolophidae	Chiroptera	Asia
<i>Labidocarpus acuminatus</i> Fain, 1982	<i>Rhinolophus acuminatus</i>	Rhinolophidae	Chiroptera	Asia
<i>Labidocarpus vittatus</i> (Fain, 1976)	<i>Rhinolophus malayanus</i>	Rhinolophidae	Chiroptera	Asia
<i>Labidocarpus australiensis</i> Fain et Lukoschus, 1981	<i>Hipposideros ater</i>	Hipposideridae	Chiroptera	Australia
”	<i>Hipposideros bicolor</i>	Hipposideridae	Chiroptera	Asia
<i>Labidocarpus laviae</i> Fain, 1970	<i>Lavia frons</i>	Megadermatidae	Chiroptera	Africa
<i>Labidocarpus formosanus</i> Fain, 1979	<i>Coelops frithii</i>	Hipposideridae	Chiroptera	Asia
<i>Labidocarpus dossuarius</i> McDaniel, 1972	<i>Chaerephon major</i>	Molossidae	Chiroptera	South America
<i>Lawrenceocarpus</i> Dusbabek et Cruz, 1966				
<i>Lawrenceocarpus micropilus</i> Dusbabek et Cruz, 1966	<i>Pteronotus quadridens</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus dusbabeki</i> Cruz, 1969	<i>Brachyphylla nana</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus puertoricensis</i> Cruz, Tamsitt et Valdiviego, 1974	<i>Brachyphylla cavernarum</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus lobus</i> McDaniel, 1971	<i>Carollia perspicillata</i>	Phyllostomidae	Chiroptera	South America
“	<i>Brachyphylla cavernarum</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus moormops</i> Gruz, 1974	<i>Mormoops blainvillei</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus mimon</i> Fain, 1970	<i>Mimon bennettii</i>	Phyllostomidae	Chiroptera	South America
“	<i>Mimon crenulatum</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus micronycteris</i> Fain, 1976	<i>Micronycteris nicefori</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus jamaicensis</i> Fain, 1976	<i>Mormoops blainvillei</i>	Mormoopidae	Chiroptera	South America
<i>Lawrenceocarpus planirostris</i> Fain, 1976	<i>Mormoops megalophylla</i>	Mormoopidae	Chiroptera	South America
<i>Lawrenceocarpus chilonycteris</i> Fain, 1981	<i>Chilonycteris fuliginosus</i>	Phyllostomidae	Chiroptera	South America

<i>Lawrenceocarpus chilonycteris</i> Fain, 1981	<i>Chilonycteris</i> sp.	Phyllostomidae	Chiroptera	South America
“	<i>Pteronotus parnellii</i>	Phyllostomidae	Chiroptera	South America
“	<i>Pteronotus personatus</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus phyllostomus</i> McDaniel, 1972	<i>Phyllostomus elongatus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Micronycteris hirsuta</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus puertoricensis</i> Cruz, Tamsitt et Valdiviero, 1974	<i>Brachyphylla cavernarum</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus delacruzii</i> Guerrero, 1995	<i>Tonatia bidens</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus faini</i> Guerrero, 1995	<i>Phyllostomus discolor</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus fudeci</i> Guerrero, 1995	<i>Lonchophylla thomasi</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus machadoallison</i> Guerrero, 1995	<i>Lonchorhina aurita</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus ojustii</i> Guerrero, 1995	<i>Mimon crenulatum</i>	Phyllostomidae	Chiroptera	South America
<i>Lawrenceocarpus terramaris</i> Guerrero, 1995	<i>Carollia perspicillata</i>	Phyllostomidae	Chiroptera	South America
<i>Megadermicolus</i> Fain, 1971				
<i>Megadermicolus yunkerii</i> Fain, 1971	<i>Cardioderma cor</i>	Megadermatidae	Chiroptera	Africa
<i>Olabidocarpus</i> Lawrence, 1948				
<i>Olabidocarpus belsorum</i> (Van Endhoven, 1940)	<i>Myotis myotis</i>	Vespertilionidae	Chiroptera	Eurasia
“	<i>Myotis oxygnathus</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis capaccinii</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis daubentonii</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis brandtii</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis mystacinus</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis adversus</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Myotis tricolor</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Myotis blythii</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Myotis goudoti</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Kerivoula smithii</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Rhinolophus hipposideros</i>	Hipposideridae	Chiroptera	Europe
<i>Olabidocarpus squamosus</i> Fain, 1971	<i>Kerivoula cuprosa</i>	Vespertilionidae	Chiroptera	Africa
<i>Olabidocarpus miniopteri</i> Fain, 1982	<i>Miniopterus inflatus</i>	Vespertilionidae	Chiroptera	Africa
<i>Olabidocarpus eptesicus</i> Fain, 1970	<i>Eptesicus brasiliensis</i>	Vespertilionidae	Chiroptera	South America
<i>Olabidocarpus plecoti</i> Fain, 1971	<i>Plecotus auritus</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis velifer</i>	Vespertilionidae	Chiroptera	North America
<i>Olabidocarpus kreceki</i> Cruz, 1973	<i>Eptesicus fuscus</i>	Vespertilionidae	Chiroptera	South America
<i>Olabidocarpus myoticola</i> Fain, 1970	<i>Myotis albescens</i>	Vespertilionidae	Chiroptera	South America
“	<i>Myotis nigricans</i>	Vespertilionidae	Chiroptera	South America
“	<i>Myotis</i> sp.	Vespertilionidae	Chiroptera	Asia
<i>Olabidocarpus orientalis</i> Fain, 1976	<i>Myotis moluccarum</i>	Vespertilionidae	Chiroptera	Australia
“	<i>Myotis adversus</i>	Vespertilionidae	Chiroptera	Asia
<i>Olabidocarpus americanus</i> McDaniel et Lawrence, 1964	<i>Lasiurus intermedius</i>	Vespertilionidae	Chiroptera	North America
“	<i>Lasiurus borealis</i>	Vespertilionidae	Chiroptera	South America
“	<i>Lasiurus cinereus</i>	Vespertilionidae	Chiroptera	Australia

A review of mammal-associated Psoroptidia

<i>Olabidocarpus whitakeri</i> McDaniel et Coffman, 1970	<i>Myotis austroriparius</i>	Vespertilionidae	Chiroptera	North America
<i>Olabidocarpus chalinolobi</i> Fain, 1979	<i>Chalinolobus gouldii</i>	Vespertilionidae	Chiroptera	Australia
<i>Olabidocarpus malayi</i> Fain, 1970	<i>Pipistrellus stenopterus</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Pipistrellus angulatus</i>	Vespertilionidae	Chiroptera	Asia
“	<i>Vespadelus douglasorum</i>	Vespertilionidae	Chiroptera	Australia
<i>Olabidocarpus nyctinomops</i> Fain, 1976	<i>Nyctinomops laticaudatus</i>	Molossidae	Chiroptera	South America
<i>Olabidocarpus lawrencei</i> McDaniel et Coffman, 1970	<i>Tadarida brasiliensis</i>	Molossidae	Chiroptera	North America
<i>Olabidocarpus kelantanensis</i> Fain, 1976	<i>Chaerephon johorensis</i>	Molossidae	Chiroptera	Asia
<i>Olabidocarpus triaenops</i> Fain, 1976	<i>Triaenops persicus</i>	Hipposideridae	Chiroptera	Africa
<i>Olabidocarpus nycteris</i> Fain, 1976	<i>Nycteris hispida</i>	Nycteridae	Chiroptera	Africa
<i>Olabidocarpus taphozous</i> Fain, 1976	<i>Saccolaimus peli</i>	Emballonuridae	Chiroptera	Africa
“	<i>Taphozous mauritanus</i>	Emballonuridae	Chiroptera	Africa
<i>Olabidocarpus tanganyikensis</i> (Pinichpongse, 1963)	<i>Coleura afra</i>	Emballonuridae	Chiroptera	Africa
“	<i>Coleura afra</i>	Emballonuridae	Chiroptera	Africa
<i>Olabidocarpus cynomops</i> Fain, 1972	<i>Cynomops planirostris</i>	Molossidae	Chiroptera	South America
<i>Olabidocarpus otomops</i> Fain, 1970	<i>Otomops martiensseni</i>	Molossidae	Chiroptera	Africa
<i>Olabidocarpus africanus</i> Fain, 1970	<i>Lavia frons</i>	Megadermatidae	Chiroptera	Africa
<i>Parakosa</i> McDaniel et Lawrence, 1962				
<i>Parakosa tadarida</i> Dusbabek et Cruz, 1966 (= <i>rectipes</i> Pinichpongse, 1963)	<i>Nyctinomops laticaudatus</i>	Molossidae	Chiroptera	North America
“	<i>Molossus molossus</i>	Molossidae	Chiroptera	South America
“	[<i>Eumops</i> or <i>Molossus</i>] <i>ater</i>	Molossidae	Chiroptera	South America
“	<i>Molossus pretiosus</i>	Molossidae	Chiroptera	South America
“	<i>Molossus molossus</i>	Molossidae	Chiroptera	South America
“	<i>Molossus currentium</i>	Molossidae	Chiroptera	South America
“	<i>Eumops auripendulus</i>	Molossidae	Chiroptera	South America
“	<i>Sturnira lilium</i>	Phyllostomidae	Chiroptera	South America
“	<i>Glossophaga longirostris</i>	Phyllostomidae	Chiroptera	South America
“	<i>Carollia brevicauda</i>	Phyllostomidae	Chiroptera	South America
“	<i>Noctilio leporinus</i>	Noctilionidae	Chiroptera	South America
<i>Parakosa flexipes</i> (Pinichpongse, 1963)	<i>Chaerephon major</i>	Molossidae	Chiroptera	South America
“	<i>Molossus molossus</i>	Molossidae	Chiroptera	South America
“	<i>Molossus aztecus</i>	Molossidae	Chiroptera	South America
“	<i>Molossus pretiosus</i>	Molossidae	Chiroptera	South America
“	[<i>Eumops</i> or <i>Molossus</i>] <i>ater</i>	Molossidae	Chiroptera	South America
“	<i>Molossus currentium</i>	Molossidae	Chiroptera	South America
“	<i>Promops centralis</i>	Molossidae	Chiroptera	South America
“	<i>Artibeus jamaicensis</i>	Phyllostomidae	Chiroptera	South America
“	<i>Enchisthensis hartii</i>	Phyllostomidae	Chiroptera	South America
“	<i>Glossophaga longirostris</i>	Phyllostomidae	Chiroptera	South America

<i>Parakosa flexipes</i> (Pinichpongse, 1963)	<i>Noctilio leporinus</i>	Noctilionidae	Chiroptera	South America
<i>Parakosa asiatica</i> Fain, 1976	<i>Chaerephon johorensis</i>	Molossidae	Chiroptera	Asia
<i>Parakosa mops</i> (Fain, 1970)	<i>Mops condylurus</i>	Molossidae	Chiroptera	Africa
<i>Parakosa indica</i> Fain, 1976	<i>Otomops wroughtoni</i>	Molossidae	Chiroptera	Asia
<i>Parakosa mormopterus</i> Fain, 1976	<i>Mormopterus beccarii</i>	Molossidae	Chiroptera	Asia
<i>Parakosa philippinensis</i> Fain, 1976	<i>Rousettus amplexicaudatus</i>	Pteropodidae	Chiroptera	Asia
<i>Paralabidocarpus</i> Pinichpongse, 1963				
<i>Paralabidocarpus artibeus</i> Pinichpongse, 1963	<i>Artibeus lituratus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Artibeus cinereus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Artibeus jamaicensis</i>	Phyllostomidae	Chiroptera	South America
“	<i>Platyrrhinus helleri</i>	Phyllostomidae	Chiroptera	South America
“	<i>Stenoderma</i> sp.	Phyllostomidae	Chiroptera	South America
“	<i>Sturnira lilium</i>	Phyllostomidae	Chiroptera	South America
“	<i>Stenoderma rufum</i>	Phyllostomidae	Chiroptera	South America
“	<i>Ardops nichollsi</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus foxi</i> Cruz, Tamsitt et Valdivieso, 1974	<i>Artibeus jamaicensis</i>	Phyllostomidae	Chiroptera	South America
“	<i>Phyllops falcatus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Stenoderma rufum</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus stenodermi</i> Cruz, Tamsitt et Valdivieso, 1974	<i>Stenoderma rufum</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus carolliae</i> Fain, 1970	<i>Carollia perspicillata</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus surinamensis</i> Fain, 1970	<i>Saccopteryx canescens</i>	Emballonuridae	Chiroptera	South America
“	<i>Saccopteryx leptura</i>	Emballonuridae	Chiroptera	South America
“	<i>Saccopteryx bilineata</i>	Emballonuridae	Chiroptera	South America
“	<i>Peropteryx</i> sp.	Emballonuridae	Chiroptera	South America
“	<i>Carollia perspicillata</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus tonatae</i> Fain, 1970	<i>Lophostoma brasiliense</i>	Phyllostomidae	Chiroptera	South America
“	<i>Lophostoma silvicolum</i>	Phyllostomidae	Chiroptera	South America
“	<i>Sturnira lilium</i>	Phyllostomidae	Chiroptera	South America
“	<i>Trachops cirrhosus</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus guineensis</i> Fain, 1970	<i>Nycteris gambiensis</i>	Nycteridae	Chiroptera	Africa
<i>Paralabidocarpus trachops</i> Fain, 1972	<i>Trachops cirrhosus</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus trachops mimon</i> Fain, 1976	<i>Mimon crenulatum</i>	Phyllostomidae	Chiroptera	South America
“	<i>Mimon bennetti</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus macrophyllum</i> Fain, 1972	<i>Macrophyllum macrophyllum</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus desmodus</i> Fain, 1972	<i>Desmodus rotundus</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus dentatus</i> Fain, 1972	<i>Megaderma spasma</i>	Megadermatidae	Chiroptera	Asia
<i>Paralabidocarpus pilosus</i> Fain, 1979	<i>Megaderma spasma</i>	Megadermatidae	Chiroptera	Asia
<i>Paralabidocarpus megaderma</i> Fain, 1982	<i>Megaderma spasma</i>	Megadermatidae	Chiroptera	Asia

A review of mammal-associated Psoroptidia

<i>Paralabidocarpus anthorhinae</i> Fain, 1973	<i>Dobsonia crenulata</i>	Phyllostomidae	Chiroptera	South America
<i>Paralabidocarpus furipterus</i> Fain, 1976	<i>Furipterus horrens</i>	Furipteridae	Chiroptera	South America
<i>Paralabidocarpus brewsterae</i> Fain, 1976	<i>Furipterus horrens</i>	Furipteridae	Chiroptera	South America
<i>Paralabidocarpus aselliscus</i> Fain, 1982	<i>Aselliscus tricuspis</i>	Hipposideridae	Chiroptera	Australia
<i>Paralabidocarpus hipposideros</i> Fain, 1976	<i>Hipposideros lankadiva</i>	Hipposideridae	Chiroptera	Asia
“	<i>Hipposideros diadema</i>	Hipposideridae	Chiroptera	Asia
<i>Paralabidocarpus coxatus</i> Fain, 1979	<i>Hipposideros pratti</i>	Hipposideridae	Chiroptera	Asia
“	<i>Rhinolophus affinis</i>	Rhinolophidae	Chiroptera	Asia
<i>Paralawrenceocarpus</i> Guerrero, 1992				
<i>Paralawrenceocarpus dossuarius</i> (McDaniel, 1972)	<i>Chaerephon major</i>	Molossidae	Chiroptera	South America
<i>Pseudoalabidocarpus</i> McDaniel, 1972				
<i>Pseudoalabidocarpus phyllostomi</i> (Fain, 1972) (= <i>secus</i> McDaniel, 1972)	<i>Phyllostomus hastatus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Phyllostomus elongatus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Phyllostomus discolor</i>	Phyllostomidae	Chiroptera	South America
“	<i>Monophyllus plethodon</i>	Phyllostomidae	Chiroptera	South America
<i>Pseudoalabidocarpus macrotus</i> (Cruz, 1974)	<i>Macrotus waterhousii</i>	Phyllostomidae	Chiroptera	South America
<i>Pseudoalabidocarpus noctilio</i> (Fain, 1970)	<i>Noctilio leporinus</i>	Noctilionidae	Chiroptera	South America
<i>Pteropiella</i> Fain, 1970				
<i>Pteropiella pteropi</i> Fain, 1970	<i>Pteropus rufus</i>	Pteropodidae	Chiroptera	Africa (Madagascar)
“	<i>Pteropus giganteus</i>	Pteropodidae	Chiroptera	Asia
<i>Pteropiella papuana</i> Fain, 1975	<i>Pteropus</i> sp.	Pteropodidae	Chiroptera	Australia (New Guinea)
<i>Trilabidocarpus</i> Fain, 1970				
<i>Trilabidocarpus kasaii</i> Fain, 1970	<i>Mops thersites</i>	Molossidae	Chiroptera	Africa
<i>Rynconyssus</i> Fain, 1967				
<i>Rynconyssus galagoensis</i> Fain, 1967	<i>Otolemur crassicaudatus</i>	Galagidae	Primates	Africa
“	<i>Perodicticus potto</i>	Lorisidae	Primates	Africa
<i>Eulabidocarpus</i> Lawrence, 1948				
<i>Eulabidocarpus compressus</i> (Ewans, 1911)	<i>Pteropus giganteus</i>	Pteropodidae	Chiroptera	Asia
Schizocarpini Fain, 1971				
<i>Lutrilichus</i> Fain, 1970				
<i>Lutrilichus schoutedeni</i> Fain, 1970	<i>Hydrictis maculicollis</i>	Mustelidae	Carnivora	Africa
<i>Lutrilichus javanicus</i> Fain, 1970	<i>Melogale orientalis</i>	Mustelidae	Carnivora	Asia
<i>Lutrilichus linsang</i> Fain, 1970	<i>Prionodon linsang</i>	Viverridae	Carnivora	Asia
<i>Lutrilichus nivalis</i> Beron, 1973	<i>Mustela nivalis</i>	Mustelidae	Carnivora	Europe
<i>Lutrilichus canadensis</i> Fain, Lukoschus et Clulow, 1974	<i>Mustela erminea</i>	Mustelidae	Carnivora	North America
<i>Schizocarpus</i> Trouessart, 1896				
<i>Schizocarpus mingaudi</i> Trouessart, 1896	<i>Castor canadensis</i>	Castoridae	Rodentia	North America

<i>Schizocarpus spinifer</i> Fain, Whitaker et Smith, 1984	<i>Castor canadensis</i>	Castoridae	Rodentia	Europe, North America
<i>Schizocarpus indianensis</i> Fain, Whitaker et Smith, 1984	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus tetrapilis</i> Fain, Whitaker et Smith, 1984	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus virgulatus</i> Fain, Whitaker et Smith, 1984	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus subvirgulatus</i> Fain, Whitaker et Smith, 1984	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus inversus</i> Fain, Whitaker et Smith, 1984	<i>Castor canadensis</i>	Castoridae	Rodentia	North America, Europe
<i>Schizocarpus furcatus</i> Fain, Whitaker et Smith, 1984	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus paramingaudi</i> Fain et Whitaker, 1988	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus alaskensis</i> Fain et Whitaker, 1988	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus reductus</i> Fain et Whitaker, 1988	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus postannulatus</i> Fain et Whitaker, 1988	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus centralis</i> Fain et Whitaker, 1988	<i>Castor canadensis</i>	Castoridae	Rodentia	North America, Europe
<i>Schizocarpus distinctus</i> Fain et Whitaker, 1988	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus contrarius</i> Fain et Whitaker, 1988	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus protinus</i> Fain et Whitaker, 1988	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus posticus</i> Fain et Whitaker, 1988	<i>Castor canadensis</i>	Castoridae	Rodentia	North America
<i>Schizocarpus anomalis</i> Bochkov, 1993	<i>Castor canadensis</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus numerosus</i> (Dubinina, 1964)	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
“	* <i>Castor canadensis</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus fedjushini</i> (Dubinina, 1964)	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus capitis</i> (Dubinina, 1964)	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus brachyurus</i> (Dubinina, 1964)	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus grandis</i> (Dubinina, 1964)	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
“	* <i>Castor canadensis</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus latus</i> (Dubinina, 1964)	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
“	* <i>Castor canadensis</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus parvus</i> (Dubinina, 1964)	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
“	<i>Castor canadensis</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus subparvus</i> (Dubinina, 1964)	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
“	* <i>Castor canadensis</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus minor</i> (Dubinina, 1964)	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
“	* <i>Castor canadensis</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus subminor</i> (Dubinina, 1964)	<i>Castor fiber</i>	Castoridae	Rodentia	Europe

A review of mammal-associated Psoroptidia

<i>Schizocarpus parabrachyurus</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus similis</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus diebzigensis</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus modestus</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus humilis</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus pusillus</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus hexapilis</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus insignis</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus intercalatus</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus pygidialis</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus ornatus</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus subornatus</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus dubininae</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus subdiebzigensis</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus brevis</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus curtus</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus exiguous</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus subhexapilis</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
“	* <i>Castor canadensis</i>	Castoridae	Rodentia	Europe
<i>Schizocarpus parahexapilis</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus radiatus</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Schizocarpus ventricosus</i> Fain et Lukoschus, 1985	<i>Castor fiber</i>	Castoridae	Rodentia	Eurasia
<i>Soricilichus</i> Fain, 1970				
<i>Soricilichus scutisorex</i> Fain, 1970	<i>Scutisorex somereni</i>	Soricidae	Soricomorpha	Africa
<i>Soricolichus kivuensis</i> Fain, 1972	Soricomorpha sp.		Soricomorpha	Africa
“	<i>Crocidura</i> sp.	Soricidae	Soricomorpha	Africa
Schizocoptinae Fain, 1970				
<i>Schizocoptes</i> Lawrence, 1944				
<i>Schizocoptes conjugatus</i> Lawrence, 1944	<i>Chrysospalax villosus</i>	Chrysochloridae	Afrosoricida	Africa
“	<i>Chrysospalax trevelyani</i>	Chrysochloridae	Afrosoricida	Africa
“	<i>Amblysomus hottentotus</i>	Chrysochloridae	Afrosoricida	Africa

“	<i>Chrysochloris stuhlmanni</i>	Chrysochloridae	Afrosoricida	Africa
“	<i>Calcochloris leucorhinus</i>	Chrysochloridae	Afrosoricida	Africa
<i>Schizocoptes chrysochloris</i> Fain, 1970	<i>Calcochloris leucorhinus</i>	Chrysochloridae	Afrosoricida	Africa
Lemuroeciinae Fain, 1968 <i>Lemuroecius</i> Fain, 1968				
<i>Lemuroecius cheirogalei</i> Fain, 1968	<i>Cheirogaleus major</i>	Cheirogaleidae	Primates	Africa (Madagascar)

Table 10. Host distribution of the family Listrophoridae Megnin et Trouessart, 1884

Mite species	Host species	Host family	Host order	Distribution
Listrophorinae Megnin et Trouessart, 1884				
<i>Listrophorus</i> Pagenstecher, 1862				
<i>Listrophorus leuckarti</i> Pagenstecher, 1862	<i>Arvicola amphibious</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus arvalis</i>	Cricetidae	Rodentia	Europe
“	<i>Myodes rutilus</i>	Cricetidae	Rodentia	Asia
<i>Listrophorus occitanus</i> Fain et Portus, 1978	<i>Microtus</i> sp.	Cricetidae	Rodentia	Europe
“	<i>Microtus duodecimcostatus</i>	Cricetidae	Rodentia	Europe
“	<i>Arvicola scherman</i>	Cricetidae	Rodentia	Europe
“	<i>Arvicola amphibious</i>	Cricetidae	Rodentia	Europe
<i>Listrophorus brevipes</i> Dubinina, 1968	<i>Microtus arvalis</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus agrestis</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus oeconomicus</i>	Cricetidae	Rodentia	Europe
“	<i>Myodes glareolus</i>	Cricetidae	Rodentia	Europe
<i>Listrophonts mediterraneus</i> Portus, Fain et Lukoschus, 1980	<i>Myodes glareolus</i>	Cricetidae	Rodentia	Europe
“	<i>Eliomys quercinus</i>	Gliridae	Rodentia	Europe
<i>Listrophorus validus</i> Banks, 1910	<i>Ondatra zibethicus</i>	Cricetidae	Rodentia	Europe, North America
<i>Listrophorus ondatrae</i> Fain, Kok et Lukoschus, 1970	<i>Ondatra zibethicus</i>	Cricetidae	Rodentia	North America
<i>Listrophorus dozieri</i> Radford, 1944	<i>Ondatra zibethicus</i>	Cricetidae	Rodentia	North America
<i>Listrophorus americanus</i> Radford, 1944	<i>Ondatra zibethicus</i>	Cricetidae	Rodentia	North America
<i>Listrophorus faini</i> Dubinina, 1972	<i>Ondatra zibethicus</i>	Cricetidae	Rodentia	North America
<i>Listrophorus kingstownensis</i> Fain et Hyland, 1973	<i>Ondatra zibethicus</i>	Cricetidae	Rodentia	North America
<i>Listrophorus laynei</i> Fain, Smith et Whitaker, 1986	<i>Neofiber alleni</i>	Cricetidae	Rodentia	North America
<i>Listrophorus caudatus</i> Fain, Smith et Whitaker, 1986	<i>Neofiber alleni</i>	Cricetidae	Rodentia	North America
<i>Listrophorus meridionalis</i> Fain, 1970	<i>Arvicola amphibious</i>	Cricetidae	Rodentia	Europe
<i>Listrophorus mexicanus</i> Fain, 1970	<i>Microtus mexicanus</i>	Cricetidae	Rodentia	North America
<i>Listrophorus mexicanus squamiferus</i> Fain et Hyland, 1972	<i>Myodes gapperi</i>	Cricetidae	Rodentia	North America
“	<i>Microtus pennsylvanicus</i>	Cricetidae	Rodentia	North America

A review of mammal-associated Psoroptidia

<i>Listrophorus mexicanus squamiferus</i> Fain et Hyland, 1972	<i>Peromyscus leucopus</i>	Cricetidae	Rodentia	North America
<i>Listrophorus phenacomys</i> Fain et Hyland, 1972	<i>Phenacomys</i> sp.	Cricetidae	Rodentia	North America
“	<i>Phenacomys intermedius</i>	Cricetidae	Rodentia	North America
<i>Listrophorus pitymys</i> Fain et Hyland, 1972	<i>Microtus pinetorum</i>	Cricetidae	Rodentia	North America
“	<i>Microtus pennsylvanicus</i>	Cricetidae	Rodentia	North America
“	<i>Blarina brevicauda</i>	Soricidae	Soricomorpha	North America
<i>Listrophorus dicrostonyx</i> Fain et Hyland, 1972	<i>Dicrostonyx</i> sp.	Cricetidae	Rodentia	North America
<i>Listrophorus neotomae</i> Fain et Hyland, 1972	<i>Neotoma micropus</i>	Cricetidae	Rodentia	North America
<i>Listrophorus synaptomys</i> Fain, Whitaker, McDaniel et Lukoschus, 1974	<i>Synaptomys cooperi</i>	Cricetidae	Rodentia	North America
<i>Listrophorus synaptomys edleri</i> Fain, Whitaker, McDaniel et Lukoschus, 1974	<i>Lemmus lemmus</i>	Cricetidae	Rodentia	Europe
<i>Aeromychirus</i> Fain, 1972				
<i>Aeromychirus aeromys</i> (Fain, 1970)	<i>Aeromys tephromelas</i>	Sciuridae	Rodentia	Asia
<i>Aeromychirus fimbriatus</i> Bochkov et OConnor, 2005	<i>Eoglaucmys fimbriatus</i>	Sciuridae	Rodentia	Asia
<i>Aeromychirus petinomys</i> Bochkov et OConnor, 2006	<i>Petinomys crinitus</i>	Sciuridae	Rodentia	Asia
<i>Afrolistrophorus</i> Fain, 1970				
<i>Afrolistrophorus lophuromys</i> (Radford, 1940)	<i>Lophuromys sikapusi</i>	Muridae	Rodentia	Africa
“	<i>Hybomys univittatus</i>	Muridae	Rodentia	Africa
“	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Lemniscomys striatus</i>	Muridae	Rodentia	Africa
“	<i>Grammomys</i> sp.	Muridae	Rodentia	Africa
“	<i>Rattus rattus</i>	Muridae	Rodentia	Africa
“	<i>Mus musculoides</i>	Muridae	Rodentia	Africa
“	<i>Uranomys ruddi</i>	Muridae	Rodentia	Africa
“	<i>Crociodura occidentalis</i>	Soricidae	Soricomorpha	Africa
<i>Afrolistrophorus congoicola</i> Fain, 1971	<i>Lophuromys flavopunctatus</i>	Muridae	Rodentia	Africa
“	<i>Lophuromys aguilus</i>	Muridae	Rodentia	Africa
“	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
“	<i>Rattus rattus</i>	Muridae	Rodentia	Africa
“	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
“	<i>Gerbilliscus leucogaster</i>	Muridae	Rodentia	Africa
“	<i>Lemniscomys striatus</i>	Muridae	Rodentia	Africa
“	<i>Dendromus insignis</i>	Nesomyidae	Rodentia	Africa
<i>Afrolistrophorus muricola</i> Fain, 1970	<i>Mus musculoides</i>	Muridae	Rodentia	Africa
“	<i>Mus minutoides</i>	Muridae	Rodentia	Africa
“	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
“	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Rattus rattus</i>	Muridae	Rodentia	Africa

<i>Afrolistrophorus muricola</i> Fain, 1970	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
“	<i>Lemniscomys striatus</i>	Muridae	Rodentia	Africa
“	<i>Oenomys hypoxanthus</i>	Muridae	Rodentia	Africa
“	<i>Gerbilliscus leucogaster</i>	Muridae	Rodentia	Africa
“	<i>Lophuromys aguilus</i>	Muridae	Rodentia	Africa
“	<i>Crociodura occidentalis</i>	Soricidae	Soricomorpha	Africa
<i>Afrolistrophorus pakistanensis</i> Fain, 1976	<i>Mus</i> sp.	Muridae	Rodentia	Asia
“	<i>Mus musculus</i>	Muridae	Rodentia	Asia
“	<i>Millardia meltada</i>	Muridae	Rodentia	Asia
“	<i>Rattus rattus</i>	Muridae	Rodentia	Asia
“	<i>Meriones hurrianae</i>	Muridae	Rodentia	Asia
“	<i>Tatera indica</i>	Muridae	Rodentia	Asia
<i>Afrolistrophorus musculus</i> Wilson et Lawrence, 1967	<i>Mus musculus</i>	Muridae	Rodentia	Asia, Australia (Hawaii)
“	<i>Millardia meltada</i>	Muridae	Rodentia	Asia
<i>Afrolistrophorus maculatus</i> Fain, 1976	<i>Leopoldamys sabanus</i>	Muridae	Rodentia	Asia
“	<i>Maxomys musschenbroekii</i>	Muridae	Rodentia	Asia
“	<i>Niviventer niviventer</i>	Muridae	Rodentia	Asia
“	<i>Rattus exulans</i>	Muridae	Rodentia	Asia
“	<i>Apomys datae</i>	Muridae	Rodentia	Asia
“	<i>Apomys microdon</i>	Muridae	Rodentia	Asia
“	<i>Chrotomys silaceus</i>	Muridae	Rodentia	Asia
“	<i>Chrotomys whiteheadi</i>	Muridae	Rodentia	Asia
“	<i>Archboldomys musseri</i>	Muridae	Rodentia	Asia
“	<i>Rhynchomys sorcoides</i>	Muridae	Rodentia	Asia
<i>Afrolistrophorus maculatus rattus</i> Fain, 1976	<i>Rattus rattus</i>	Muridae	Rodentia	South America
“	<i>Rattus rattus</i>	Muridae	Rodentia	South America
<i>Afrolistrophorus apodemi</i> Fain, 1970	<i>Apodemus sylvaticus</i>	Muridae	Rodentia	Europe
“	<i>Apodemus flavicollis</i>	Muridae	Rodentia	Europe
“	<i>Myodes smithii</i>	Cricetidae	Rodentia	Asia
<i>Afrolistrophorus brevis</i> Fain, 1970	<i>Steatomys opimus</i>	Nesomyidae	Rodentia	Africa
“	<i>Steatomys pratensis</i>	Nesomyidae	Rodentia	Africa
<i>Afrolistrophorus dasymys</i> Fain, 1970	<i>Dasymys incomtus</i>	Muridae	Rodentia	Africa
“	<i>Dasymys rufulus</i>	Muridae	Rodentia	Africa
<i>Afrolistrophorus otomys</i> Fain, 1970	<i>Otomys tropicalis</i>	Muridae	Rodentia	Africa
“	<i>Taterillus emini</i>	Muridae	Rodentia	Africa
“	<i>Mylomys dybowskii</i>	Muridae	Rodentia	Africa
“	<i>Oenomys hypoxanthus</i>	Muridae	Rodentia	Africa
<i>Afrolistrophorus concinius</i> Fain, 1970	<i>Mus musculoides</i>	Muridae	Rodentia	Africa
“	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Lemniscomys striatus</i>	Muridae	Rodentia	Africa
“	<i>Lophuromys sikapusi</i>	Muridae	Rodentia	Africa
“	<i>Otomys tropicalis</i>	Muridae	Rodentia	Africa
<i>Afrolistrophorus steatomys</i> Fain, 1970	<i>Steatomys pratensis</i>	Nesomyidae	Rodentia	Africa
<i>Afrolistrophorus stochomys</i> Fain, 1971	<i>Stochomys longicaudatus</i>	Muridae	Rodentia	Africa
“	<i>Steatomys opimus</i>	Muridae	Rodentia	Africa

A review of mammal-associated Psoroptidia

<i>Afrolistophorus chiropodomys</i> Fain, 1970	<i>Chiropodomys major</i>	Muridae	Rodentia	Asia
<i>Afrolistophorus tachyoryctes</i> (Coffee, 1971)	<i>Tachyoryctes splendens</i>	Spalacidae	Rodentia	Africa
“	<i>Tachyoryctes daemon</i>	Spalacidae	Rodentia	Africa
“	<i>Tachyoryctes ruandae</i>	Spalacidae	Rodentia	Africa
“	<i>Deomys ferrugineus</i>	Muridae	Rodentia	Africa
<i>Afrolistophorus dipodicola dipodicola</i> (Tragardh, 1904)	<i>Gerbillus gerbillus</i>	Muridae	Rodentia	Africa
“	<i>Gerbillus pyramidum</i>	Muridae	Rodentia	Africa
“	<i>Ctenodactylus gundi</i>	Ctenodactylidae	Rodentia	Africa
<i>Afrolistophorus dipodicola theodori</i> (Radford, 1954)	<i>Meriones arimalius</i>	Muridae	Rodentia	Asia, Africa
<i>Afrolistophorus dipodicola taterae</i> Fain, 1971	<i>Gerbilliscus kempi</i>	Muridae	Rodentia	Africa
“	<i>Gerbilliscus validus</i>	Muridae	Rodentia	Africa
“	<i>Taterillus gracilis</i>	Muridae	Rodentia	Africa
<i>Afrolistophorus bothae</i> (Hirst, 1923)	<i>Gerbillinae</i> sp.	Muridae	Rodentia	Africa
“	<i>Gerbilliscus leucogaster</i>	Muridae	Rodentia	Africa
“	<i>Gerbilliscus leucogaster</i>	Muridae	Rodentia	Africa
“	<i>Lophuromys</i> sp.	Muridae	Rodentia	Africa
“	<i>Lophuromys aguilus</i>	Muridae	Rodentia	Africa
“	<i>Aethomys chrysophilus</i>	Muridae	Rodentia	Africa
“	<i>Oenomys</i> sp.	Muridae	Rodentia	Africa
“	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
“	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
“	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
“	<i>Grammomys dolichurus</i>	Muridae	Rodentia	Africa
“	<i>Lemniscomys striatus</i>	Muridae	Rodentia	Africa
“	! <i>Crocidura</i> sp.	Soricidae	Soricomorpha	Africa
<i>Afrolistophorus cannomys</i> Fain, 1980	<i>Cannomys badius</i>	Spalacidae	Rodentia	Asia
<i>Afrolistophorus sicista</i> Fain, 1970	<i>Sicista subtilis</i>	Dipodidae	Rodentia	Asia
<i>Afrolistophorus rhizomys</i> (Fain, 1970)	<i>Rhizomys pruinosus</i>	Spalacidae	Rodentia	Asia
<i>Afrolistophorus obesus</i> Fain et Lukoschus, 1983	<i>Spalax graecus</i>	Spalacidae	Rodentia	Europe
“	<i>Spalax microphthalmus</i>	Spalacidae	Rodentia	Asia
“	<i>Spalax leucodon</i>	Spalacidae	Rodentia	Europe
<i>Afrolistophorus punctatus</i> Fain et Lukoschus, 1983	<i>Spalax leucodon</i>	Spalacidae	Rodentia	Europe
<i>Afrolistophorus myospalaxis</i> Dubinina, 1973	<i>Myospalax myospalax</i>	Spalacidae	Rodentia	Asia
<i>Afrolistophorus sumatrensis</i> Fain, 1980	<i>Rhizomys sumatrensis</i>	Spalacidae	Rodentia	Asia
<i>Afrolistophorus neacomys</i> Fain et Lukoschus, 1973	<i>Neacomys tenuipes</i>	Cricetidae	Rodentia	South America
“	<i>Neacomys spinosus</i>	Cricetidae	Rodentia	South America
<i>Afrolistophorus medius</i> Fain et Lukoschus, 1983	<i>Tarsius pumilus</i>	Tarsiidae	Primates	Asia
<i>Afrolistophorus bradypus</i> Fain, 1981	<i>Bradypus tridactylus</i>	Bradypodidae	Pilosa	South America

<i>Afrolistrophorus (Spalacarus)</i> Fain, 1980 stat. nov.				
<i>Afrolistrophorus spalacis</i> (Fain, 1970)	<i>Spalax nehringi</i>	Spalacidae	Rodentia	Asia
“	<i>Spalax microphthalmus</i>	Spalacidae	Rodentia	Europe
<i>Afrolistrophorus costai</i> (Fain, 1981)	<i>Spalax microphthalmus</i>	Spalacidae	Rodentia	Asia
<i>Afrolistrophorus sinensis</i> (Fain, 1970)	<i>Myospalax psilurus</i>	Spalacidae	Rodentia	Asia
<i>Afrolistrophorus mediolineatus</i> Fain, 1976	<i>Bandicota bengalensis</i>	Muridae	Rodentia	Asia
<i>Afrolistrophorus mediolineatus nesokia</i> Fain et Hyland, 1980	<i>Nesokia indica</i>	Muridae	Rodentia	Asia
<i>Afrolistrophorus stubbei</i> (Fain et Lukoschus, 1979)	<i>Allactaga sibirica</i>	Dipodidae	Rodentia	Asia
<i>Afrolistrophorus elongatus</i> Fain et Lukoschus, 1983	<i>Ctenodactylus gundi</i>	Ctenodacylidae	Rodentia	Africa
<i>Amlistrophorus</i> Fain, 1981 stat. nov.				
<i>Amlistrophorus hornensis</i> (Fain, 1976)	<i>Euneomys chinchilloides</i>	Cricetidae	Rodentia	South America
<i>Amlistrophorus inca</i> (Fain, 1976)	<i>Lestoros inca</i>	Caenolestidae	Paucituberculata	South America
<i>Amlistrophorus laticoxa</i> (Fain et Lukoschus, 1982)	<i>Necomys urichi</i>	Cricetidae	Rodentia	South America
<i>Amlistrophorus venezuelensis</i> (Fain et Lukoschus, 1983) comb. nov.	<i>!Monodelphis brevicaudata</i>	Didelphidae	Didelphimorphia	South America
<i>Asiochirus</i> Fain, 1970				
<i>Asiochirus suncus</i> (Radford, 1940)	<i>Suncus murinus</i>	Soricidae	Soricomorpha	Asia
<i>Asiochirus chimarrogale</i> Fain, 1976	<i>Chimarrogale himalayica</i>	Soricidae	Soricomorpha	Asia
“	<i>Nectogale elegans</i>	Soricidae	Soricomorpha	Asia
<i>Asiochirus soriculus</i> Fain et Bochkov, 2003	<i>Episoriculus leucops</i>	Soricidae	Soricomorpha	Asia
<i>Asiochirus nepalensis</i> Fain et Bochkov, 2003	<i>Episoriculus caudatus</i>	Soricidae	Soricomorpha	Asia
<i>Carnilistrophorus</i> Fain, 1980				
<i>Carnilistrophorus poecilogalei</i> (Fain, 1970)	<i>Poecilogale albinucha</i>	Mustelidae	Carnivora	Africa
<i>Carnilistrophorus myospalacis</i> (Fain, 1970)	<i>Eospalax fontanierii</i>	Spalacidae	Rodentia	Asia
<i>Carnilistrophorus (Carnilistrophorellus)</i> Fain, 1980				
<i>Carnilistrophorus rhynchocyoni</i> (Fain, 1970)	<i>Rhynchocyon cirnei</i>	Macroscelidae	Macroscelidea	Africa
<i>Carnilistrophorus myonax</i> (Fain, 1970)	<i>Galerella sanguinea</i>	Viverridae	Carnivora	Africa
<i>Carnilistrophorus genettus</i> (Radford, 1944)	<i>Genetta</i> sp.	Viverridae	Carnivora	Africa
“	<i>Genetta tigrina</i>	Viverridae	Carnivora	Africa
<i>Dubininetta</i> Fain et Lukoschus, 1978				
<i>Dubininetta talpae</i> (Fain et Lukoschus, 1978)	<i>Euroscaptor longirostris</i>	Talpidae	Soricomorpha	Asia
<i>Dubininetta dubinini</i> (Dubinina, 1969)	<i>Desmana moschata</i>	Talpidae	Soricomorpha	Europe
“	<i>Galemys pyrenaicus</i>	Talpidae	Soricomorpha	Europe
“	<i>Mogera wogura</i>	Talpidae	Soricomorpha	Asia
<i>Dubininetta taiwanensis</i> (Fain et Lukoschus, 1978)	<i>Mogera insularis</i>	Talpidae	Soricomorpha	Asia
<i>Echinosorella</i> Fain, 1980				
<i>Echinosorella echinosorex</i> (Fain et Lukoschus, 1978)	<i>Echinosorex gymmura</i>	Erinaceidae	Erinaceomorpha	Asia

A review of mammal-associated Psoroptidia

<i>Geomylichus</i> Fain, 1970				
<i>Geomylichus dipodomys</i> (Radford, 1953)	<i>Dipodomys phillipsii</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus texanus</i> Fain, Whitaker, Schwan et Lukoschus, 1978	<i>Dipodomys ordii</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus utahensis</i> Fain et Whitaker, 1987	<i>Dipodomys microps</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus intercalatus</i> Fain, Whitaker et Thomas, 1991	<i>Dipodomys compactus</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus californicus</i> Fain, Whitaker et Thomas, 1988	<i>Dipodomys venustus</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus quasinudus</i> Fain, Whitaker et Thomas, 1991	<i>Dipodomys ingens</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus multistriatus</i> Fain, Whitaker et Thomas, 1988	<i>Dipodomys nitratoides</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus microdipodops</i> Fain et Whitaker, 1980	<i>Microdipodops megacephalus</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus perognathi</i> Fain et Whitaker, 1980	<i>Perognathus parvus</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus inaequalis</i> Fain, Whitaker, Schwan et Lukoschus, 1978	<i>Chaetodipus hispidus</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus hispidus</i> Vargas, Perez et Polaco, 1999	<i>Chaetodipus hispidus</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus formosus</i> Fain et Whitaker, 1987	<i>Chaetodipus formosus</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus brevispinosus</i> Fain, Whitaker, Schwan et Lukoschus, 1978	<i>Chaetodipus penicillatus</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus penicillatus</i> Vargas, Perez et Polaco, 1999	<i>Chaetodipus eremicus</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus nelsoni</i> Vargas, Perez et Polaco, 1999	<i>Chaetodipus nelsoni</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus durangoensis</i> Vargas, Perez et Polaco, 1999	<i>Chaetodipus nelsoni</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus comitanensis</i> Hoffman et Servin, 1990	<i>Chaetodipus arenarius</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus bassolsae</i> Servin, Aguilar et Alvaroz-Castaneda, 1992	<i>Chaetodipus arenarius</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus guaycurensis</i> Servin, Aguilar et Alvaroz-Castaneda, 1994	<i>Chaetodipus arenarius</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus thomomys</i> Fain, Whitaker, Schwan et Lukoschus, 1978	<i>Thomomys bottae</i>	Geomyidae	Rodentia	North America
<i>Geomylichus (Neogeomylichus)</i> Fain et Whitaker, 1987				
<i>Geomylichus postscutatus</i> Fain, 1976	<i>Dipodomys</i> sp.	Heteromyidae	Rodentia	North America
<i>Geomelichus (Whitakerobius)</i> Fain, 1981				
<i>Geomelichus floridanus</i> (Radford, 1949)	<i>Geomys pinetis</i>	Geomyidae	Rodentia	North America
<i>Geomelichus deserti</i> Fain et Whitaker, 1987	<i>Dipodomys deserti</i>	Heteromyidae	Rodentia	North America
<i>Geomelichus (Ageomylichus)</i> Fain, 1981				
<i>Geomylichus nectomys</i> Fain, Whitaker, Schwan et Lukoschus, 1978	<i>Nectomys</i> sp.	Cricetidae	Rodentia	South America
<i>Geomylichus mexicanus</i> Fain, 1976	<i>Neotoma phenax</i>	Cricetidae	Rodentia	North America
<i>Geomylichus neacomys</i> Fain, Whitaker, Schwan et Lukoschus, 1978	<i>Neacomys tenuipes</i>	Cricetidae	Rodentia	South America

<i>Geomylichus oaxacae</i> Fain et Estebanes, 2000	<i>Heteromys desmarestianus</i>	Heteromyidae	Rodentia	North America
<i>Geomylichus klebergi</i> (McDaniel, 1965)	<i>Sigmodon hispidus</i>	Cricetidae	Rodentia	North America
<i>Geomylichus incertae sedis</i>				
<i>Geomylichus sylvilagus</i> Fain, 1973	<i>Sylvilagus floridanus</i>	Leporidae	Lagomorpha	North America
<i>Hemigalichus</i> Fain, 1970				
<i>Hemigalichus baramensis</i> Fain, 1970	<i>Hemigalus derbyanus</i>	Viverridae	Carnivora	Asia
<i>Hemigalichus chrotogale</i> Bochkov et OConnor, 2005	<i>Chrotogale owstoni</i>	Viverridae	Carnivora	Asia
<i>Leporacarus</i> Fain, 1970				
<i>Leporacarus gibbus</i> (Pagenstecher, 1861)	<i>Oryctolagus cuniculus</i>	Leporidae	Lagomorpha	Cosmopolite
“	<i>Lepus capensis</i>	Leporidae	Lagomorpha	Africa
“	<i>Lepus europaeus</i>	Leporidae	Lagomorpha	Europe
“	<i>Lepus californicus</i>	Leporidae	Lagomorpha	North America
<i>Leporacarus (Leporacaroides)</i> Fain, 1971				
<i>Leporacarus leporicolus</i> (Lawrence, 1951)	<i>Lepus saxatilis</i>	Leporidae	Lagomorpha	Africa
“	* <i>Galerella sanguinea</i>	Viverridae	Carnivora	Africa
<i>Leporacarus sylvilagi</i> Fain, Lukoschus et Whitaker, 1981	<i>Sylvilagus bachmani</i>	Leporidae	Lagomorpha	North America
<i>Leporacarus brevicaudatus</i> (Ullrich, 1938)	<i>Lepus timidus</i>	Leporidae	Lagomorpha	Europe
“	<i>Lepus europaeus</i>	Leporidae	Lagomorpha	Europe
<i>Lynxacarus</i> Radford, 1951				
<i>Lynxacarus morlani</i> Radford, 1951	<i>Lynx rufus</i>	Felidae	Carnivora	North America
<i>Lynxacarus mustelae</i> (Megnin, 1885)	<i>Mustela nivalis</i>	Mustelidae	Carnivora	Europe
“	<i>Martes martes</i>	Mustelidae	Carnivora	Europe
“	<i>Mustela putorius</i>	Mustelidae	Carnivora	Europe
“	<i>Mustela erminea</i>	Mustelidae	Carnivora	Europe, North America
“	<i>Mustela nivalis</i>	Mustelidae	Carnivora	Europe
“	<i>Mustela frenata</i>	Mustelidae	Carnivora	North America
“	<i>Martes foina</i>	Mustelidae	Carnivora	Europe
“	<i>Meles meles</i>	Mustelidae	Carnivora	Europe
<i>Lynxacarus nearcticus</i> Fain et Hyland, 1973	<i>Mustela lutreola</i>	Mustelidae	Carnivora	North America
“	<i>Mustela frenata</i>	Mustelidae	Carnivora	North America
“	! <i>Eutamias</i> sp.	Sciuridae	Rodentia	North America
<i>Lynxacarus radovskyi</i> Tenorio, 1974	<i>Felis catus</i>	Felidae	Carnivora	Australia (New Zealand), South America
<i>Lynxacarus semnopithecii</i> Fain, 1970	! <i>Presbytis hosei</i>	Cercopithecidae	Primates	Asia
“	<i>Tupaia javanica</i>	Tupaiaidae	Scandentia	Asia
“	* <i>Paradoxurus hermaphroditus</i>	Viverridae	Carnivora	Asia
<i>Lynxacarus tupaiae</i> Fain, 1970	<i>Tupaia belangeri</i>	Tupaiaidae	Scandentia	Asia
“	<i>Tupaia montana</i>	Tupaiaidae	Scandentia	Asia
“	<i>Tupaia glis</i>	Tupaiaidae	Scandentia	Asia

A review of mammal-associated Psoroptidia

<i>Lynxacarus palawanensis</i> Fain, 1976	<i>Tupaia palawanensis</i>	Tupaiidae	Scandentia	Asia
<i>Lynxacarus lyncodon</i> Fain, 1970	<i>Lyncodon patagonicus</i>	Mustelidae	Carnivora	South America
<i>Lynxacarus grandior</i> Fain, 1976	! <i>Sundamys infraluteus</i>	Muridae	Rodentia	Asia
<i>Lynxacarus (Lutracarus)</i> Fain et Yunker, 1980				
<i>Lynxacarus canadensis</i> (Fain et Yunker, 1980)	<i>Lutra canadensis</i>	Mustelidae	Carnivora	North America
<i>Lynxacarus visoni</i> Fain et Bochkov, 2002	<i>Neovison vison</i>	Mustelidae	Carnivora	North America
<i>Metalistrophorus</i> Fain, 1970				
<i>Metalistrophorus pagenstecheri</i> (Halle, 1880)	<i>Sciurus vulgaris</i>	Sciuridae	Rodentia	Eurasia, North America
<i>Metalistrophorus sciuricola</i> Fain, 1970	<i>Tamias sibiricus</i>	Sciuridae	Rodentia	Asia
<i>Metalistrophorus laosensis</i> (Fain, 1978)	Sciuridae sp.	Sciuridae	Rodentia	Asia
<i>Metalistrophorus pahangensis</i> (Fain, 1976)	<i>Tamiops maclellandi</i>	Sciuridae	Rodentia	Asia
<i>Olistrophorus</i> McDaniel et Whitaker, 1972				
<i>Olistrophorus cryptotae</i> McDaniel et Whitaker, 1972	<i>Cryptotis parva</i>	Soricidae	Soricomorpha	North America
<i>Olistrophorus guatemalensis</i> (Fain, 1979)	<i>Cryptotis tropicalis</i>	Soricidae	Soricomorpha	South America
<i>Olistrophorus blarina</i> (Fain et Hyland, 1972)	<i>Blarina brevicauda</i>	Soricidae	Soricomorpha	North America
<i>Olistrophorus crocidurae</i> (Fain, 1976)	<i>Suncus etruscus</i>	Soricidae	Soricomorpha	Asia
<i>Olistrophorus platacanthomys</i> (Fain, 1970)	<i>Platacanthomys lasiurus</i>	Platacanthomyidae	Rodentia	Asia
“?”	<i>Dryomys nitedula</i>	Gliridae	Rodentia	Europe
<i>Olistrophorus (Mexicochirus)</i> Fain et Estibanes, 1996 comb. nov.				
<i>Olistrophorus bilobatus</i> (Fain et Estibanes, 1996)	<i>Sorex</i> sp.	Soricidae	Soricomorpha	North America
<i>Olistrophorus unilobatus</i> (Fain et Estibanes, 1996)	<i>Sorex</i> sp.	Soricidae	Soricomorpha	North America
<i>Prolistrophorus</i> Fain, 1970				
<i>Prolistrophorus argentinus</i> (Hirst, 1921)	<i>Kunsia tomentosus</i>	Cricetidae	Rodentia	South America
“	<i>Oryzomys subflavus</i>	Cricetidae	Rodentia	South America
<i>Prolistrophorus frontalis</i> (Hirst, 1921)	<i>Oligoryzomys delticola</i>	Cricetidae	Rodentia	South America
“	<i>Oligoryzomys fulvescens</i>	Cricetidae	Rodentia	North America
<i>Prolistrophorus grassii</i> (Radford, 1954)	<i>Oryzomys palustris</i>	Cricetidae	Rodentia	North America
“	<i>Synaptomys cooperi</i>	Cricetidae	Rodentia	North America
<i>Prolistrophorus paraguayensis</i> Fain, 1970	<i>Oryzomys angouya</i>	Cricetidae	Rodentia	South America
<i>Prolistrophorus amazonicus</i> Fain, 1970	<i>Oryzomys laticeps</i>	Cricetidae	Rodentia	South America
<i>Prolistrophorus amazonicus musculus</i> Fain, 1973	<i>Mus musculus</i>	Muridae	Rodentia	South America
<i>Prolistrophorus pernamboucensis</i> Fain, 1973	<i>Oryzomys</i> sp.	Cricetidae	Rodentia	South America
<i>Prolistrophorus nectomys</i> Fain, 1970	<i>Nectomys squamipes</i>	Cricetidae	Rodentia	South America
“	<i>Nectomys rattus</i>	Cricetidae	Rodentia	South America
“	<i>Baiomys musculus</i>	Cricetidae	Rodentia	North America

<i>Prolistrophorus nectomys</i> Fain, 1970	<i>Phyllomys brasiliensis</i>	Echimyidae	Rodentia	South America
“	<i>Mesomys hispidus</i>	Echimyidae	Rodentia	South America
“	<i>!Monodelphis brevicaudata</i>	Didelphidae	Didelphimorphia	South America
<i>Prolistrophorus surinamensis</i> Fain, 1973	<i>Nectomys rattus</i>	Cricetidae	Rodentia	South America
<i>Prolistrophorus striatus</i> Fain, 1973	<i>Zygodontomys brevicauda</i>	Cricetidae	Rodentia	South America
“	<i>Phyllomys brasiliensis</i>	Echimyidae	Rodentia	South America
<i>Prolistrophorus sclerobursatus</i> Fain et Estebanes, 1996	<i>Peromyscus</i> sp.	Cricetidae	Rodentia	North America
<i>Prolistrophorus cryptophallus</i> Fain, 1970	<i>Kannabateomys amblyonyx</i>	Echimyidae	Rodentia	South America
<i>Prolistrophorus (Aprolistrophorus)</i> Fain, 1980				
<i>Prolistrophorus sparsilineatus</i> Fain, 1970	<i>Peromyscus gossypinus</i>	Cricetidae	Rodentia	North America
<i>Prolistrophorus ctenomys</i> Fain, 1970	<i>Ctenomys talarum</i>	Ctenomyiidae	Rodentia	South America
<i>Prolistrophorus akodon</i> Fain et Lukoschus, 1984	<i>Necomys urichi</i>	Cricetidae	Rodentia	South America
“	<i>Necomys lasiurus</i>	Cricetidae	Rodentia	South America
<i>Prolistrophorus primitivus</i> Fain et Lukoschus, 1984	<i>Oryzomys albigularis</i>	Cricetidae	Rodentia	South America
<i>Prolistrophorus scotinomys</i> Fain et Lukoschus, 1982	<i>Scotinomys xerampelinus</i>	Cricetidae	Rodentia	South America
<i>Prolistrophorus reithrodontomys</i> Fain et Lukoschus, 1982	<i>Reithrodontomys creper</i>	Cricetidae	Rodentia	South America
<i>Prolistrophorus monilistriatus</i> Fain et Lukoschus, 1984	<i>Lestoros inca</i>	Caenolestidae	Paucituberculata	South America
<i>Prolistrophorus lestoros</i> Fain et Lukoschus, 1982	<i>Lestoros inca</i>	Caenolestidae	Paucituberculata	South America
<i>Prolistrophorus bidentatus</i> Fain et Lukoschus, 1982	<i>Lestoros inca</i>	Caenolestidae	Paucituberculata	South America
<i>Prolistrophorus postscutatus</i> Fain et Lukoschus; 1982	<i>Lestoros inca</i>	Caenolestidae	Paucituberculata	South America
<i>Prolistrophorus cuzcoensis</i> Fain et Lukoschus, 1982	<i>Lestoros inca</i>	Caenolestidae	Paucituberculata	South America
<i>Prolistrophorus curvistriatus</i> Fain et Lukoschus, 1984	<i>Lestoros inca</i>	Caenolestidae	Paucituberculata	South America
<i>Prolistrophorus birkenholzi</i> Fain, Smith et Whitaker, 1986	<i>Neofiber alleni</i>	Cricetidae	Rodentia	North America
<i>Prolistrophorus (Beprolistrophorus)</i> Fain, 1980				
<i>Prolistrophorus bakeri</i> (Radford, 1949)	<i>Sigmodon hispidus</i>	Cricetidae	Rodentia	North America
<i>Prolistrophorus hirstianus</i> Fain, 1973	<i>Kunsia tomentosus</i>	Cricetidae	Rodentia	South America
<i>Prolistrophorus incerae sedis</i>				
<i>Prolistrophorus inornatus</i> Lizaso, 1977	<i>Oryzomys subflavus</i>	Cricetidae	Rodentia	South America
<i>Prolistrophorus dolichus</i> Lizaso, 1975	Rodentia sp.		Rodentia	South America
<i>Pteromychirus</i> Fain, 1980				
<i>Pteromychirus lukoschusi</i> (Fain, 1978)	<i>Pteromys momonga</i>	Sciuridae	Rodentia	Asia
<i>Quasilistrophorus</i> Fain, Whitaker et Lukoschus, 1978				
<i>Quasilistrophorus microticulus</i> Fain, Whitaker et Lukoschus, 1978	<i>Arborimus albipes</i>	Cricetidae	Rodentia	North America

A review of mammal-associated Psoroptidia

<i>Quasilistrophorus microticolus</i> Fain, Whitaker et Lukoschus, 1978	<i>Arborimus longicaudus</i>	Cricetidae	Rodentia	North America
“	<i>Synaptomys cooperi</i>	Cricetidae	Rodentia	North America
<i>Sciurochirus</i> Fain, 1972				
<i>Sciurochirus philippinensis</i> Fain, 1972	<i>Sundasciurus steerii</i>	Sciuridae	Rodentia	Asia
“	<i>Sundasciurus philippinensis</i>	Sciuridae	Rodentia	Asia
“	<i>Sundasciurus tenuis</i>	Sciuridae	Rodentia	Asia
“	<i>Callosciurus notatus</i>	Sciuridae	Rodentia	Asia
“	<i>Ratufa bicolor</i>	Sciuridae	Rodentia	Asia
“	<i>Ratufa affinis</i>	Sciuridae	Rodentia	Asia
“	<i>Petinomys crinitus</i>	Sciuridae	Rodentia	Asia
<i>Sciurochirus thailandiae</i> Fain, 1978	<i>Callosciurus canniceps</i>	Sciuridae	Rodentia	Asia
<i>Sciurochirus tupaiae</i> Fain, 1972	<i>Tupaia glis</i>	Tupaiaidae	Scandentia	Asia
<i>Sclerolistrophorus</i> Fain, 1976				
<i>Sclerolistrophorus oxymycterus</i> Fain, 1976	<i>Oxymycterus gvaestor</i>	Cricetidae	Rodentia	South America
“	<i>Oryzomys megacephalus</i>	Cricetidae	Rodentia	South America
“	<i>Oryzomys yunganus</i>	Cricetidae	Rodentia	South America
<i>Sclerolistrophorus oryzomys</i> Fain, 1976	<i>Oryzomys laticeps</i>	Cricetidae	Rodentia	South America
<i>Sclerolistrophorus neacomys</i> (Fain et Lukoschus, 1980)	<i>Neacomys tenuipes</i>	Cricetidae	Rodentia	South America
<i>Sclerolistrophorus peruviansis</i> Fain et Ritzi, 2001	<i>Oryzomys megacephalus</i>	Cricetidae	Rodentia	South America
<i>Sclerolistrophorus longipes</i> Fain et Ritzi, 2001	<i>Oryzomys megacephalus</i>	Cricetidae	Rodentia	South America
“	<i>Oryzomys yunganus</i>	Cricetidae	Rodentia	South America
Listrophorinae incertae sedis				
<i>Listrophorus arishi</i> Oyouun, Kamman et Kady, 1994	<i>Gerbillus pyramidum</i>	Muridae	Rodentia	Africa
Aplodontochirinae Fain et Hyland, 1972				
<i>Aplodontochirus</i> Fain et Hyland, 1972				
<i>Aplodontochirus borealis</i> Fain et Hyland, 1972	<i>Aplodontia rufa</i>	Aplodontiidae	Rodentia	North America

Table 11. Host distribution of the family Myocoptidae Gunther, 1942

The paper by Haitlinger (1986) contains too many obviously accidental records and is not considered here.

Mite species	Host species	Host family	Host order	Distribution
<i>Myocoptes</i> Claparede, 1869				
<i>Myocoptes musculus</i> (Koch, 1844)	<i>Mus musculus</i>	Muridae	Rodentia	Cosmopolite
“	<i>Apodemus sylvaticus</i>	Muridae	Rodentia	Europe
“	<i>Apodemus flavicollis</i>	Muridae	Rodentia	Europe
“	* <i>Peromyscus leucopus</i>	Cricetidae	Rodentia	North America
“	* <i>Dasyurus maculatus</i>	Dasyuridae	Dasyuromorphia	Australia
<i>Myocoptes hoogstraali</i> Fain, 1971	<i>Gerbillus gerbillus</i>	Muridae	Rodentia	Africa
<i>Myocoptes lepidotus</i> (Lawrence, 1956)	<i>Aethomys chrysophilus</i>	Muridae	Rodentia	Africa
<i>Myocoptes grammomys</i> Fain, 1970	<i>Grammomys dolichurus</i>	Muridae	Rodentia	Africa

<i>Myocoptes grammomys</i> Fain, 1970	<i>Grammomys dolichurus</i>	Muridae	Rodentia	Africa
“	<i>Oenomys hypoxanthus</i>	Muridae	Rodentia	Africa
<i>Myocoptes kivuensis</i> Fain, 1970	<i>Thomomys bottae</i>	Muridae	Rodentia	Africa
<i>Myocoptes malacomys</i> Fain, 1970	<i>Malacomys</i> sp.	Muridae	Rodentia	Africa
“	<i>Malacomys edwardsi</i>	Muridae	Rodentia	Africa
“	<i>Malacomys longipes</i>	Muridae	Rodentia	Africa
<i>Myocoptes rattus</i> Fain et Zumpt, 1977	<i>Rattus norvegicus</i>	Muridae	Rodentia	Africa
<i>Myocoptes hybomys</i> Fain, 1970	<i>Hybomys univittatus</i>	Muridae	Rodentia	Africa
<i>Myocoptes spinulatus</i> Fain, 1970	<i>Dendromus melanotis</i>	Nesomyidae	Rodentia	Africa
“	<i>Dendromus mesomelas</i>	Nesomyidae	Rodentia	Africa
<i>Myocoptes dendromus</i> Fain, 1970	<i>Dendromus mesomelas</i>	Nesomyidae	Rodentia	Africa
“	<i>Dendromus melanotis</i>	Nesomyidae	Rodentia	Africa
<i>Myocoptes japonensis japonensis</i> Radford, 1955	<i>Myodes smithii</i>	Cricetidae	Rodentia	Asia
“	<i>Myodes glareolus</i>	Cricetidae	Rodentia	Europe
“	<i>Myodes gapperi</i>	Cricetidae	Rodentia	North America
“	<i>Microtus agrestis</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus arvalis</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus subterraneus</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus pennsylvanicus</i>	Cricetidae	Rodentia	North America
“	<i>Microtus oeconomus</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus tatricus</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus miurus egorovi</i> (extinct subspecies)	Cricetidae	Rodentia	Asia
“	<i>Arvicola amphibious</i>	Cricetidae	Rodentia	Europe
“	<i>Chionomys nivalis</i>	Cricetidae	Rodentia	Europe
“	<i>Marmota monax</i>	Sciuridae	Rodentia	North America
<i>Myocoptes japonensis canadensis</i> Radford, 1955	<i>Dicrostonyx</i> sp.	Cricetidae	Rodentia	North America
“	<i>Dicrostonyx groenlandicus</i>	Cricetidae	Rodentia	North America
<i>Myocoptes squamosus</i> Fain, Munting et Lukoschus, 1969	<i>Microtus oeconomus</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus pennsylvanicus</i>	Cricetidae	Rodentia	North America
<i>Myocoptes pitymys</i> Fain et Bochkov, 2004	<i>Microtus pinetorum</i>	Cricetidae	Rodentia	North America
<i>Myocoptes ondatrae</i> Lukoschus et Rouwet, 1968	<i>Ondatra zibethicus</i>	Cricetidae	Rodentia	Europe
<i>Myocoptes neotomae</i> Fain, Lukoschus, Cudmore et Whitaker, 1984	<i>Neotoma floridana</i>	Cricetidae	Rodentia	North America
<i>Myocoptes persicus</i> Fain, 1970	<i>Cricetulus migratorius</i>	Cricetidae	Rodentia	Asia
<i>Myocoptes ictonyx</i> Fain, 1970	<i>Ictonyx striatus</i>	Mustelidae	Carnivora	Africa
<i>Myocoptes (Omyocoptes)</i> Fain, 1986				
<i>Myocoptes gerbillicola</i> Fain, 1970	<i>Taterillus</i> sp.	Muridae	Rodentia	Africa
“	<i>Taterillus congicus</i>	Muridae	Rodentia	Africa
“	<i>Gerbilliscus kempfi</i>	Muridae	Rodentia	Africa
<i>Myocoptes queenslandicus</i> Fain, 1986	<i>Mesembriomys gouldii</i>	Muridae	Rodentia	Australia

A review of mammal-associated Psoroptidia

<i>Myocoptes (Comyocoptes)</i> Fain, 1970				
<i>Myocoptes verrucosus</i> Fain, 1970	<i>Tatera</i> sp.	Muridae	Rodentia	Africa
<i>Myocoptes striatus</i> Fain, 1970	<i>Saccostomus campestris</i>	Nesomyidae	Rodentia	Africa
“	<i>Steatomys opimus</i>	Nesomyidae	Rodentia	Africa
<i>Myocoptes (Amyocoptes)</i> Fain, 1972				
<i>Myocoptes nudus</i> Fain, 1970	<i>Lophuromys flavopunctatus</i>	Muridae	Rodentia	Africa
“	<i>Aethomys nyikae</i>	Muridae	Rodentia	Africa
<i>Crinicaster</i> Poppe, 1889				
<i>Crinicaster criceti</i> Poppe, 1889	<i>Cricetus cricetus</i>	Cricetidae	Rodentia	Europe
<i>Crinicaster apodemi</i> Fain, Munting et Lukoschus, 1969	<i>Apodemus sylvaticus</i>	Muridae	Rodentia	Europe
“	<i>Apodemus agrarius</i>	Muridae	Rodentia	Europe
<i>Crinicaster congolensis</i> Fain, 1970	<i>Grammomys poensis</i>	Muridae	Rodentia	Africa
<i>Crinicaster deomys</i> Fain, 1970	<i>Deomys ferrugineus</i>	Muridae	Rodentia	Africa
<i>Gliricoptes</i> Lawrence, 1956				
<i>Gliricoptes glirinus</i> (Canestrini, 1895)	<i>Glis glis</i>	Gliridae	Rodentia	Eurasia
<i>Gliricoptes muscardinus</i> Kok, Lukoschus et Fain, 1971	<i>Muscardinus avallanarius</i>	Gliridae	Rodentia	Europe
<i>Gliricoptes nitidulus</i> Kok, Lukoschus et Fain, 1971	<i>Dryomys nitidula</i>	Gliridae	Rodentia	Europe
<i>Gliricoptes eliomyis</i> Kok, Lukoschus et Fain, 1971	<i>Eliomyis quercinus</i>	Gliridae	Rodentia	Europe, Africa
<i>Gliricoptes vulcanorum</i> Fain, 1970	<i>Graphiurus murinus</i>	Gliridae	Rodentia	Africa
<i>Gliricoptes graphiuri</i> Fain, 1970	<i>Graphiurus nagtglasii</i>	Gliridae	Rodentia	Africa
<i>Gliricoptes asiaticus</i> Fain, 1970	<i>Typhlomys cinereus</i>	Platacanthomyidae	Rodentia	Asia
<i>Gliricoptes betulinus</i> Kok, Lukoschus et Fain, 1971	<i>Sicista betulina</i>	Dipodidae	Rodentia	Europe
<i>Gliricoptes zapus</i> Fain et Whitaker, 1974	<i>Zapus trinotatus</i>	Dipodidae	Rodentia	North America
<i>Sciurocoptes</i> Fain, Munting et Lukoschus, 1970				
<i>Sciurocoptes sciurinus</i> (Hennemann, 1919)	<i>Sciurus vulgaris</i>	Sciuridae	Rodentia	Europe
<i>Sciurocoptes tamias</i> Fain et Hyland, 1970	<i>Tamias striatus</i>	Sciuridae	Rodentia	North America
<i>Trichoecius</i> Canestrini, 1899				
<i>Trichoecius tenax</i> (Michael, 1889)	<i>Microtus agrestis</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus arvalis</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus oeconomus</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus subterraneus</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus tatricus</i>	Cricetidae	Rodentia	Europe
“	<i>Microtus pennsylvanicus</i>	Cricetidae	Rodentia	North America
“	<i>Myodes smithii</i>	Cricetidae	Rodentia	Asia
<i>Trichoecius clethrionomyidis</i> Portus et Gallego, 1986	<i>Myodes glareolus</i>	Cricetidae	Rodentia	Europe
<i>Trichoecius pitymydis</i> Portus et Gallego, 1986	<i>Microtus duodecimcostatus</i>	Cricetidae	Rodentia	Europe
<i>Trichoecius tibetanus</i> Fain, 1970	<i>Cricetulus kamensis</i>	Cricetidae	Rodentia	Asia

<i>Trichoecius gettingeri</i> Fain, Lukoschus, Cudmore et Whitaker, 1984	<i>Peromyscus leucopus</i>	Cricetidae	Rodentia	North America
<i>Trichoecius calomysci</i> Bochkov, Malikov et Arbobi, 1999	<i>Calomyscus</i> sp.	Calomyscidae	Rodentia	Asia
<i>Trichoecius akodon</i> Fain, 1970	<i>Akodon albiventer</i>	Cricetidae	Rodentia	South America
<i>Trichoecius romboutsii</i> (Van Eyndhoven, 1946)	<i>Mus musculus</i>	Muridae	Rodentia	Cosmopolite
<i>Trichoecius muris</i> Fain, Munting et Lukoschus, 1969	<i>Rattus norvegicus</i>	Muridae	Rodentia	Europe
<i>Trichoecius micromys</i> Fain, Munting et Lukoschus, 1969	<i>Micromys minutus</i>	Muridae	Rodentia	Europe
<i>Trichoecius apodemi</i> Fain, Munting et Lukoschus, 1969	<i>Apodemus sylvaticus</i>	Muridae	Rodentia	Europe
<i>Trichoecius blaszaki</i> Labrzycka et Dabert, 2008	<i>Apodemus flavicollis</i>	Muridae	Rodentia	Europe
<i>Trichoecius widawaensis</i> Haitlinger, 1986	<i>Apodemus agrarius</i>	Muridae	Rodentia	Europe
<i>Trichoecius kalrai</i> (Radford, 1947)	<i>Gerbillus</i> sp.	Muridae	Rodentia	Africa
“	<i>Taterillus</i> sp.	Muridae	Rodentia	Africa
“	<i>Taterillus congicus</i>	Muridae	Rodentia	Africa
“	<i>Gerbilliscus kempii</i>	Muridae	Rodentia	Africa
<i>Trichoecius malacomys</i> Fain, 1970	<i>Malacomys edwardsi</i>	Muridae	Rodentia	Africa
<i>Trichoecius euphallas</i> Fain, 1972	<i>Mus minutoides</i>	Muridae	Rodentia	Africa
<i>Trichoecius lootensi</i> Fain, 1970	<i>Lophuromys flavopunctatus</i>	Muridae	Rodentia	Africa
<i>Trichoecius natalensis</i> Fain, 1972	<i>Mastomys natalensis</i>	Muridae	Rodentia	Africa
<i>Trichoecius otomys</i> Fain, 1970	<i>Otomys irroratus</i>	Muridae	Rodentia	Africa
<i>Trichoecius hollidayi</i> Lawrence, 1951	<i>Rhabdomys pumilio</i>	Muridae	Rodentia	Africa
<i>Trichoecius hauwaertsi</i> Fain, 1970	<i>Dendromus melanotis</i>	Nesomyidae	Rodentia	Africa
<i>Trichoecius idiuri</i> Fain, 1970	<i>Idiurus macrotis</i>	Anomaluridae	Rodentia	Africa
<i>Apocalypsis</i> Bochkov gen. nov.				
<i>Apocalypsis allactaga</i> (Fain et Lukoschus, 1979)	<i>Allactaga sibirica</i>	Dipodidae	Rodentia	Asia

Table 12. Host distribution of the family Gastronyssidae Fain, 1959

Mite species	Host species	Host family	Host order	Distribution
Gastronyssinae Fain, 1959				
Gastronyssini Fain, 1959				
<i>Gastronyssus</i> Fain, 1955				
<i>Gastronyssus bakeri</i> Fain, 1955	<i>Eidolon helvum</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Africa
”	<i>Cynopterus brachyotis</i>	Pteropodidae	Chiroptera	Asia
“	<i>Cynopterus horsfieldii</i>	Pteropodidae	Chiroptera	Asia
“	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
<i>Gastronyssus philippinensis</i> Bochkov et OConnor, 2008	<i>Harpyionycteris whiteheadi</i>	Pteropodidae	Chiroptera	Asia
Phyllostomyssini Bochkov et OConnor, 2008				
<i>Phyllostomyssus</i> Fain, 1970				
<i>Phyllostomyssus conradyunkeri</i> Fain, 1970	<i>Artibeus lituratus</i>	Phyllostomidae	Chiroptera	South America
“	<i>Artibeus jamaicensis</i>	Phyllostomidae	Chiroptera	South America

A review of mammal-associated Psoroptidia

<i>Phyllostomonyssus. conradyunkeri</i> Fain, 1970	<i>Platyrrhinus helleri</i>	Phyllostomidae	Chiroptera	South America
“	<i>Uroderma bilobatum</i>	Phyllostomidae	Chiroptera	South America
Rodhainyssini Fain, 1964				
<i>Rodhainyssus</i> Fain, 1956				
<i>Rodhainyssus yunkeri</i> Fain, 1956	<i>Lavia frons</i>	Megadermatidae	Chiroptera	Africa
<i>Rodhainyssus nycteris</i> Fain, 1967	<i>Nycteris</i> sp.	Nycteridae	Chiroptera	Africa
<i>Rodhainyssus longipilis</i> Fain, 1959	<i>Cynomops planirostris</i>	Molossidae	Chiroptera	South America
“	<i>Molossus molossus</i>	Molossidae	Chiroptera	South America
<i>Rodhainyssus myotis</i> Fain, 1967	<i>Myotis myotis</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Myotis velifer</i>	Vespertilionidae	Chiroptera	North America
“	<i>Nyctalus noctula</i>	Vespertilionidae	Chiroptera	Europe
“	<i>Otonycteris hemprichii</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Kerivoula smithii</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Histiotus velatus</i>	Vespertilionidae	Chiroptera	South America
<i>Rodhainyssus surinamensis</i> Fain et Lukoschus, 1972	<i>Eptesicus</i> sp. [<i>mel-</i> <i>anopterus</i>]	Vespertilionidae	Chiroptera	South America
<i>Rodhainyssus miniopteri</i> Fain, 1967	<i>Miniopterus shreibersii</i>	Vespertilionidae	Chiroptera	Africa
<i>Rodhainyssus eptesicus</i> Fain, 1967	<i>Neoromicia rendalli</i>	Vespertilionidae	Chiroptera	Africa
“	<i>Eptesicus serotinus</i>	Vespertilionidae	Chiroptera	Europe
<i>Rodhainyssus nyctophili</i> Fain, 1967	<i>Nyctophilus</i> sp.	Vespertilionidae	Chiroptera	Australia
<i>Rodhainyssus taphozous</i> Fain, 1967	<i>Saccolaimus peli</i>	Emballonuridae	Chiroptera	Africa
<i>Rodhainyssus emballonurae</i> Fain, 1967	<i>Emballonura monticola</i>	Emballonuridae	Chiroptera	Asia
<i>Rodhainyssus balantiopteryx</i> Fain, 1967	<i>Balantiopteryx plicata</i>	Emballonuridae	Chiroptera	North America
<i>Rodhainyssus saccopteryx</i> Bochkov et OConnor, 2008	<i>Saccopteryx bilineata</i>	Emballonuridae	Chiroptera	South America
<i>Mycteronyssus</i> Fain, 1959				
<i>Mycteronyssus polli</i> Fain, 1959	<i>Hypsignathus monstrosus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Epomophorus wahlbergi</i>	Pteropodidae	Chiroptera	Africa
<i>Mycteronyssus haplonycteris</i> Bochkov et OConnor, 2008	<i>Haplonycteris fischeri</i>	Pteropodidae	Chiroptera	Asia
“	<i>Cynopterus brachyotis</i>	Pteropodidae	Chiroptera	Asia
<i>Opsonyssus</i> Fain, 1959				
<i>Opsonyssus brutsaerti</i> Fain, 1959	<i>Epomophorus labiatus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Epomophorus wahlbergi</i>	Pteropodidae	Chiroptera	Africa
“	<i>Micropteropus pusillus</i>	Pteropodidae	Chiroptera	Africa
“	<i>Rousettus aegyptiacus</i>	Pteropodidae	Chiroptera	Africa
<i>Opsonyssus brutsaerti indica</i> Fain, 1959	<i>Eonycteris spelaea</i>	Pteropodidae	Chiroptera	Asia
<i>Opsonyssus pseudoindicus</i> Bochkov et OConnor, 2008	<i>Ptenochirus jagori</i>	Pteropodidae	Chiroptera	Asia
“	<i>Ptenochirus minor</i>	Pteropodidae	Chiroptera	Asia
“	<i>Macroglossus minimus</i>	Pteropodidae	Chiroptera	Asia
<i>Opsonyssus eidoloni</i> Fain, 1959	<i>Eidolon helvum</i>	Pteropodidae	Chiroptera	Africa

<i>Opsonyssus pteropi</i> Bochkov et OConnor, 2008	<i>Pteropus hypomelanus</i>	Pteropodidae	Chiroptera	Asia
<i>Opsonyssus asiaticus</i> Fain, 1959	<i>Pteropus giganteus</i>	Pteropodidae	Chiroptera	Asia
“	<i>Pteropus punius</i>	Pteropodidae	Chiroptera	Asia
“	<i>Pteropus melanopogon</i>	Pteropodidae	Chiroptera	Asia
<i>Opsonyssus klompeni</i> Bochkov et OConnor, 2008	<i>Pteropus leucopterus</i>	Pteropodidae	Chiroptera	Asia
<i>Opsonyssus macroglossus</i> Bochkov et OConnor, 2008	<i>Macroglossus minimus</i>	Pteropodidae	Chiroptera	Asia
<i>Pseudoopsonyssus</i> Bochkov et OConnor, 2008				
<i>Pseudoopsonyssus phyllorhinae</i> (Fain, 1959)	<i>Hipposideros diadema</i>	Hipposideridae	Chiroptera	Australia (New Guinea)
“	<i>Hipposideros armiger</i>	Hipposideridae	Chiroptera	Asia
<i>Pseudoopsonyssus zumpti</i> (Fain, 1959)	<i>Rhinolophus capensis</i>	Rhinolophidae	Chiroptera	Africa
“	<i>Rhinolophus affinis</i>	Rhinolophidae	Chiroptera	Asia
<i>Pseudoopsonyssus striatus</i> Fain, 1967	<i>Rhinolophus hipposideros</i>	Rhinolophidae	Chiroptera	Europe
“	<i>Rhinolophus ferrumequinum</i>	Rhinolophidae	Chiroptera	Europe
<i>Eidolonyssus</i> Fain, 1967				
<i>Eidolonyssus ugandae</i> (Fain, 1964)	<i>Eidolon helvum</i>	Pteropodidae	Chiroptera	Africa
<i>Eidolonyssus myersi</i> Bochkov et OConnor, 2008	<i>Haplonycteris fischeri</i>	Pteropodidae	Chiroptera	Asia
“	<i>Cynopterus sphinx</i>	Pteropodidae	Chiroptera	Asia
Yunkeracarinae Fain, 1964				
<i>Yunkeracarus</i> Fain, 1957				
<i>Yunkeracarus muris</i> Fain, 1957	<i>Arvicanthis niloticus</i>	Muridae	Rodentia	Africa
“	<i>Pelomys fallax</i>	Muridae	Rodentia	Africa
“	<i>Lemniscomys striatus</i>	Muridae	Rodentia	Africa
“	<i>Rhabdomys pumilio</i>	Muridae	Rodentia	Africa
<i>Yunkeracarus microti</i> Smith, Whitaker et Giesen, 1985	<i>Microtus pennsylvanicus</i>	Cricetidae	Rodentia	North America
“	<i>Microtus arvalis</i>	Cricetidae	Rodentia	Europe
<i>Yunkeracarus ascanicus</i> Zabludovskaya, 1989	<i>Microtus socialis</i>	Cricetidae	Rodentia	Europe
<i>Yunkeracarus alticola</i> Zabludovskaya, 2008	<i>Alticola argentatus</i>	Cricetidae	Rodentia	Asia
<i>Yunkeracarus faini</i> Hyland et Clark, 1959	<i>Peromyscus leucopus</i>	Cricetidae	Rodentia	North America
“	<i>Peromyscus mexicanus</i>	Cricetidae	Rodentia	South America
<i>Yunkeracarus apodemi</i> Fain, Lukoschus, Jadin et Ah, 1967	<i>Apodemus sylvaticus</i>	Muridae	Rodentia	Europe
“	<i>Apodemus agrarius</i>	Muridae	Rodentia	Asia
“	<i>Apodemus pallipes</i>	Muridae	Rodentia	Asia
“?”	<i>Mus musculus</i>	Muridae	Rodentia	Europe
“?”	<i>Microtus socialis</i>	Cricetidae	Rodentia	Europe
<i>Yunkeracarus lophuromys</i> Bochkov et OConnor, 2008	<i>Lophuromys flavopunctatus</i>	Muridae	Rodentia	Africa
<i>Yunkeracarus otomys</i> Bochkov et OConnor, 2008	<i>Otomys typus</i>	Muridae	Rodentia	Africa
<i>Yunkeracarus hylomyscus</i> Bochkov et OConnor, 2008	<i>Hylomyscus arcimontensis</i>	Muridae	Rodentia	Africa

A review of mammal-associated Psoroptidia

<i>Yunkeracarus hylomyscus</i> Bochkov et OConnor, 2008	<i>Praomys jacksoni</i>	Muridae	Rodentia	Africa
<i>Yunkeracarus apomys</i> Bochkov et OConnor, 2008	<i>Apomys</i> sp. C	Muridae	Rodentia	Asia
<i>Yunkeracarus rattus</i> Bochkov et OConnor, 2008	<i>Rattus rattus</i>	Muridae	Rodentia	Africa (Madagascar)
<i>Yunkeracarus limnomys</i> Bochkov et OConnor, 2008	<i>Limnomys bryophilus</i>	Muridae	Rodentia	Asia
“	<i>Tarsomys apoensis</i>	Muridae	Rodentia	Asia
“	<i>Batomys salomonseni</i>	Muridae	Rodentia	Asia
<i>Sciuracarus</i> Fain, 1964				
<i>Sciuracarus paraxeri</i> Fain, 1964	<i>Paraxerus cepapi</i>	Sciuridae	Rodentia	Africa

Table 13. Host distribution of the family Lemurnyssidae Fain, 1957

Mite species	Host species	Host family	Host order	Distribution
<i>Lemurnyssus</i> Fain, 1957				
<i>Lemurnyssus galagoensis</i> Fain, 1957	<i>Galago moholi</i>	Galagidae	Primates	Africa
<i>Mortelmansia</i> Fain, 1959				
<i>Mortelmansia longus</i> Fain, 1959	<i>Saimiri sciureus</i>	Cebidae	Primates	South America
<i>Mortelmansia brevis</i> Fain, 1959	<i>Saimiri sciureus</i>	Cebidae	Primates	South America
<i>Mortelmansia duboisi</i> Fain, 1964	<i>Callithrix jacchus</i>	Callitrichidae	Primates	South America

Table 14. Host distribution of the family Pneumocoptidae Baker, Camin, Cunliffe, Woolley et Yunker, 1958

Mite species	Host species	Host family	Host order	Distribution
<i>Pneumocoptes</i> Baker, 1951				
<i>Pneumocoptes penrosei</i> (Weidman, 1917)	<i>Cynomys ludovicianus</i>	Sciuridae	Rodentia	North America
<i>Pneumocoptes tiollaisi</i> Doby, 1963	<i>Myodes glareolus</i>	Cricetidae	Rodentia	Europe
<i>Pneumocoptes jellisoni</i> Baker, 1951	<i>Peromyscus</i> sp.	Cricetidae	Rodentia	North America
“	<i>Onychomys leucogaster</i>	Cricetidae	Rodentia	North America
<i>Pneumocoptes banksi</i> (Wellman et Wherry, 1910)	<i>Spermophilus beecheyi</i>	Sciuridae	Rodentia	North America

Table 15. Host distribution of the subfamily Listropsoralginae Fain, 1965

Mite species	Host species	Host family	Host order	Distribution
<i>Listropsoralges</i> Fain, 1965				
<i>Listropsoralges marmosae</i> Fain, 1965	<i>Marmosa robinsoni</i>	Didelphidae	Didelphimorphia	South America
“	<i>Caluromys derbianus</i>	Didelphidae	Didelphimorphia	South America
<i>Listropsoralges caluromys</i> Bochkov et Wauthy, 2009	<i>Caluromys lanatus</i>	Didelphidae	Didelphimorphia	South America
“	<i>Caluromys philander</i>	Didelphidae	Didelphimorphia	South America
<i>Listropsoralges faini</i> Bochkov et Wauthy, 2009	<i>Marmosa murina</i>	Didelphidae	Didelphimorphia	South America
<i>Listropsoralges monodelphis</i> Fain, 1965	<i>Monodelphis dimidiata</i>	Didelphidae	Didelphimorphia	South America
<i>Listropsoralgoides</i> Fain et Lukoschus, 1970				
<i>Listropsoralgoides surinamensis</i> Fain et Lukoschus, 1970	<i>Proechimys guyannensis</i>	Echimyidae	Rodentia	South America
<i>Petauralges</i> Fain et Lukoschus, 1979				
<i>Petauralges rackae</i> Fain et Lukoschus, 1979	<i>Petaurus breviceps</i>	Petauridae	Diprotodontia	Australia
<i>Petauralges mordax</i> Domrow, 1988	<i>Phascogale tapoatafa</i>	Dasyuridae	Dasyuromorphia	Australia

Table 16. Host distribution of the subfamily Dromiciocoptinae Fain, 1970

Mite species	Host species	Host family	Host order	Distribution
<i>Dromiciocoptes</i> Fain, 1970				
<i>Dromiciocoptes brieni</i> Fain, 1970	<i>Dromiciops gliroides</i>	Didelphidae	Didelphimorphia	South America
<i>Dromiciocoptes marmosops</i> Bochkov et OConnor, 2008	<i>Marmosops parvidens</i>	Didelphidae	Didelphimorphia	South America
<i>Dromiciocoptes caenolestes</i> Bochkov et OConnor, 2008	<i>Caenolestes fuliginosus</i>	Caenolestidae	Paucituberculata	South America

Table 17. Host distribution of the family Psoroptidae Canestrini, 1892

Hosts species of some Psoroptinae are not listed — they are too numerous and infestation of many of them is not natural. In the same way some curious records of psoroptin species, like finding *Otodectes cynotis* in human ears (Heyning and Thienpont 1977) are not listed.

Mite species	Host species	Host family	Host order	Distribution
Psoroptinae Canestrini, 1892				
<i>Psoroptes</i> Gervais, 1841				
<i>Psoroptes ovis</i> (von Hering, 1838) [= <i>Psoroptes equi</i> (von Hering, 1838), <i>Psoroptes cuniculi</i> (Delafond, 1859), <i>Psoroptes cervinus</i> (Ward, 1915)]	<i>Ovis aries</i>	Bovidae	Artiodactyla	Cosmopolite
“	Perissodactyla spp.		Perissodactyla	Cosmopolite
“	Artiodactyla spp.		Perissodactyla	Cosmopolite
	* <i>Oryctolagus cuniculus</i>	Leporidae	Lagomorpha	Cosmopolite
“	* <i>Lepus europaeus</i>	Leporidae	Lagomorpha	Europe
“	<i>Ovis aries</i>	Bovidae	Artiodactyla	Cosmopolite
<i>Psoroptes natalensis</i> (Hirst, 1919)	<i>Bos primigenius</i>	Bovidae	Artiodactyla	Africa
“	<i>Bubalus bubalis</i> or <i>Bubalus arnee</i>	Bovidae	Artiodactyla	Africa
“	<i>Equus caballus</i>	Equidae	Perissodactyla	Cosmopolite
<i>Psoroptes pienaari</i> Fain, 1970	<i>Syncerus caffer</i>	Bovidae	Artiodactyla	Africa
<i>Chorioptes</i> Gervais et van Beneden, 1859				
<i>Chorioptes bovis</i> (von Hering, 1845) (= <i>Chorioptes japonensis</i> Takahashi et Nogami, 2001 syn. nov.)	<i>Bos primigenius</i>	Bovidae	Artiodactyla	Cosmopolite
“	Artiodactyla spp.		Artiodactyla	Cosmopolite
“	Perissodactyla spp.		Perissodactyla	Cosmopolite
<i>Chorioptes texanus</i> Hirst, 1924	<i>Capra hircus</i>	Bovidae	Artiodactyla	Cosmopolite
<i>Chorioptes crewei</i> (Lavoipierre, 1958)	<i>Cephalophus rufilatus</i>	Bovidae	Artiodactyla	Africa
<i>Chorioptes mydaus</i> Fain, 1975	<i>Mydaus javanensis</i>	Mephitidae	Carnivora	Asia
<i>Chorioptes panda</i> Fain et Leclerc, 1975	<i>Ailuropoda melano-leuca</i>	Ursidae	Carnivora	Asia
“	<i>Ursus thibetanus</i>	Ursidae	Carnivora	Asia
<i>Choriotodectes</i> Fain, 1975				
<i>Choriotodectes impala</i> Fain, 1975	<i>Aepyceros melampus</i>	Bovidae	Artiodactyla	Africa
<i>Choriopsoroptes</i> Sweatman, Walker et Bindernagel, 1964				
<i>Choriopsoroptes kenyensis</i> Sweatman, Walker and Bindernagel, 1964	<i>Syncerus caffer</i>	Bovidae	Artiodactyla	Africa
<i>Choriopsoroptes syncercus</i> Fain et Mortelmans, 1975	<i>Syncerus caffer</i>	Bovidae	Artiodactyla	Africa

A review of mammal-associated Psoroptidia

<i>Psorochorioptes</i> Fain, 1963				
<i>Psorochorioptes zumpti</i> Fain, 1963	<i>Connochaetes taurinus</i>	Bovidae	Artiodactyla	Africa
<i>Otodectes</i> Canestrini, 1894				
<i>Otodectes cynotis</i> (von Hering, 1838)	<i>Canis familiaris</i>	Canidae	Carnivora	Cosmopolite
“	Carnivora spp.		Carnivora	Cosmopolite
<i>Caparinia</i> Canestrini, 1894				
<i>Caparinia setifera</i> (Megnin, 1880)	<i>Hyaena hyaena</i>	Hyaenidae	Carnivora	Africa
<i>Caparinia erinacei</i> Fain, 1962	<i>Atelerix frontalis</i>	Erinaceidae	Erinaceomorpha	Africa
“	<i>Atelerix albiventris</i>	Erinaceidae	Erinaceomorpha	Africa
<i>Caparinia algirus</i> Fain et Portus, 1979	<i>Atelerix algirus</i>	Erinaceidae	Erinaceomorpha	Europe
<i>Caparinia tripilis</i> (Michael, 1889)	<i>Erinaceus europaeus</i>	Erinaceidae	Erinaceomorpha	Europe
“	* <i>Ictonyx striatus</i>	Mustelidae	Carnivora	Africa
<i>Caparinia lophiomys</i> Fain, 1975	<i>Lophiomys imhausi</i>	Cricetidae	Rodentia	Africa
“	<i>Jaculus jaculus</i>	Dipodidae	Rodentia	Africa
<i>Echimyalgae</i> Fain, 1967				
<i>Echimyalgae belluominii</i> Fain, 1967	<i>Kannabateomys amblyonyx</i>	Echimyidae	Rodentia	South America
<i>Hyracoptes</i> Fain et Lukoschus, 1981				
<i>Hyracoptes emersoni</i> Fain et Lukoschus, 1981	<i>Dendrohyrax dorsalis</i>	Procaviidae	Hyracoidea	Africa
<i>Trouessalgae</i> Fonseca, 1954				
<i>Trouessalgae peccari</i> Fonseca, 1954	<i>Pecari tajacu</i>	Tayassuidae	Artiodactyla	South America
Psoralginae Oudemans, 1908				
<i>Psoralges</i> Trouessart, 1896				
<i>Psoralges libertus</i> Trouessart, 1896	<i>Tamandua tetradactyla</i>	Myrmecophagidae	Pilosa	South America
<i>Psoralges andrei</i> Fain et Johnston, 1964	<i>Bradypus tridactylus</i>	Bradypodidae	Pilosa	South America
<i>Acaroptes</i> Womersley, 1953				
<i>Acaroptes vombatus</i> Womersley, 1953	<i>Vombatus ursinus</i>	Vombatidae	Diprotodontia	Australia
<i>Acaroptes womersleyi</i> Fain, 1965	<i>Vombatus ursinus</i>	Vombatidae	Diprotodontia	Australia
<i>Edentalges</i> Fonseca, 1954				
<i>Edentalges bradypus</i> Fonseca, 1954	<i>Bradypus tridactylus</i>	Bradypodidae	Pilosa	South America
<i>Edentalges choloepi</i> Fain, 1964	<i>Choloepus didactylus</i>	Megalonychidae	Pilosa	South America
<i>Edentalges quadrilobatus</i> Fonseca, 1954	<i>Myrmecophaga tridactyla</i>	Myrmecophagidae	Pilosa	South America
<i>Myoproctalges</i> Fain et Lukoschus, 1974				
<i>Myoproctalges surinamensis</i> Fain et Lukoschus, 1974	<i>Myoprocta acouchy</i>	Dasyproctidae	Rodentia	South America
<i>Coendalges</i> Fain et Mendez, 1979				
<i>Coendalges panamensis</i> Fain et Mendez, 1979	<i>Coendou rothschildi</i>	Erethizontidae	Rodentia	South America
Marsupialginae Fain, 1963				
<i>Marsupialges</i> Fain, 1963				
<i>Marsupialges misonnei</i> Fain, 1963	<i>Caluromys philander</i>	Didelphidae	Didelphimorphia	South America
“	<i>Didelphis marsupialis</i>	Didelphidae	Didelphimorphia	South America
“	<i>Marmosa murina</i>	Didelphidae	Didelphimorphia	South America
“	<i>Holochilus chacarius</i>	Cricetidae	Rodentia	South America

Makialginae Gaud et Mouchet, 1959				
<i>Makialges</i> Gaud et Till, 1957				
<i>Makialges lepilemuri</i> Gaud et Till, 1957	<i>Lepilemur mustelinus</i>	Lepilemuridae	Primates	Africa (Madagascar)
“	<i>Lepilemur ruficaudatus</i>	Lepilemuridae	Primates	Africa (Madagascar)
<i>Makialges lobatus</i> Fain, 1966	<i>Lepilemur ruficaudatus</i>	Lepilemuridae	Primates	Africa (Madagascar)
<i>Makialges sternodons</i> Gaud et Till, 1957	<i>Lepilemur</i> sp.	Lepilemuridae	Primates	Africa (Madagascar)
<i>Galagalges</i> Fain, 1963				
<i>Galagalges congolensis</i> Fain, 1963	<i>Galago moholi</i>	Galagidae	Primates	Africa
<i>Cheirogalalges</i> Fain, 1963				
<i>Cheirogalalges evansi</i> Fain, 1963	<i>Cheirogaleus medius</i>	Cheirogaleidae	Primates	Africa (Madagascar)
“	<i>Cheirogaleus major</i>	Cheirogaleidae	Primates	Africa (Madagascar)
<i>Daubentoniales</i> Fain, 1972				
<i>Daubentoniales brygooi</i> Fain, 1972	<i>Daubentonia madagascariensis</i>	Daubentoniidae	Primates	Africa (Madagascar)
<i>Gaudalges</i> Fain, 1963				
<i>Gaudalges propithecii</i> (Gaud et Till, 1957)	<i>Propithecus verreauxi</i>	Indriidae	Primates	Africa (Madagascar)
<i>Gaudalges brevisetosus</i> Bochkov et OConnor, 2006	<i>Eulemur coronatus</i>	Lemuridae	Primates	Africa (Madagascar)
<i>Gaudalges caparti</i> Fain, 1963	<i>Eulemur coronatus</i>	Lemuridae	Primates	Africa (Madagascar)
“	<i>Hapalemur griseus</i>	Lemuridae	Primates	Africa (Madagascar)
<i>Gaudalges haymani</i> Fain, 1963	<i>Eulemur fulvus</i>	Lemuridae	Primates	Africa (Madagascar)
“	<i>Eulemur coronatus</i>	Lemuridae	Primates	Africa (Madagascar)
<i>Lemuralges</i> Fain, 1963				
<i>Lemuralges intermedius</i> Fain, 1963	<i>Lepilemur ruficaudatus</i>	Lepilemuridae	Primates	Africa (Madagascar)
“	<i>Eulemur fulvus</i>	Lemuridae	Primates	Africa (Madagascar)
“	<i>Hapalemur griseus</i>	Lemuridae	Primates	Africa (Madagascar)
“	<i>Propithecus verreauxi</i>	Indriidae	Primates	Africa (Madagascar)
Cebalinae Fain, 1962				
<i>Cebalges</i> Fain, 1962				
<i>Cebalges gaudi</i> Fain, 1962	<i>Cebus capucinus</i>	Cebidae	Primates	South America
“	<i>Cebus apella</i>	Cebidae	Primates	South America
<i>Alouattalges</i> Fain, 1963				
<i>Alouattalges corbeti</i> Fain, 1963	<i>Alouatta seniculus</i>	Atelidae	Primates	South America
<i>Fonsecalges</i> Fain, 1962				
<i>Fonsecalges johnjadini</i> Fain, 1962	<i>Callithrix jacchus</i>	Cebidae	Primates	South America
<i>Fonsecalges johnjadini</i> Fain, 1962	<i>Callithrix geoffroyi</i>	Cebidae	Primates	South America
<i>Fonsecalges saimiri</i> Fain, 1963	<i>Saimiri sciureus</i>	Cebidae	Primates	South America
“	<i>Saguinus</i> sp.	Cebidae	Primates	South America

A review of mammal-associated Psoroptidia

<i>Cebalgoides</i> Fain, 1963				
<i>Cebalgoides cebi</i> Fain, 1963	<i>Cebus albifrons</i>	Cebidae	Primates	South America
"	<i>Cebus apella</i>	Cebidae	Primates	South America
"	<i>Saguinus oedipus</i>	Cebidae	Primates	South America
<i>Procebalges</i> Fain, 1963				
<i>Procebalges pitheciae</i> Fain, 1963	<i>Pithecia monachus</i>	Pitheciidae	Primates	South America
<i>Schizopodalges</i> Fain, 1963				
<i>Schizopodalges lagothricola</i> Fain, 1963	<i>Lagothrix lagotricha</i>	Atelidae	Primates	South America
Nasaliinae Fain et Nadchatram, 1979				
<i>Nasaliages</i> Fain et Nadchatram, 1979				
<i>Nasaliages borneensis</i> Fain et Nadchatram, 1979	<i>Nasalis larvatus</i>	Cercopithecidae	Primates	Asia

Table 18. Host distribution of the family Lobalgidae Fain, 1965

Mite species	Host species	Host family	Host order	Distribution
Lobalinae Fain, 1965				
<i>Lobalges</i> Fonseca, 1954				
<i>Lobalges trouessarti</i> Fonseca, 1954	<i>Bradypus variegatus</i>	Bradypodidae	Pilosa	South America
"	<i>Bradypus tridactylus</i>	Bradypodidae	Pilosa	South America
"	<i>Choloepus didactylus</i>	Megalonychidae	Pilosa	South America
Echimytricalginae Fain, 1970				
<i>Echimytricalges</i> Fain, 1970				
<i>Echimytricalges brasiliensis</i> Fain et Lukoschus, 1970	<i>Phyllomys pattoni</i>	Echimyidae	Rodentia	South America
"	<i>Trinomys albispinus</i>	Echimyidae	Rodentia	South America
"	<i>Trinomys iheringi</i>	Echimyidae	Rodentia	South America
<i>Echimytricalges surinamensis</i> Fain et Lukoschus, 1970	<i>Proechimys guyannensis</i>	Echimyidae	Rodentia	South America
"	<i>Proechimys oconnelli</i>	Echimyidae	Rodentia	South America
"	<i>Proechimys quadruplicatus</i>	Echimyidae	Rodentia	South America
"	<i>Proechimys cuvieri</i>	Echimyidae	Rodentia	South America
<i>Echimytricalges hoplomys</i> Fain, Lukoschus et Mendez, 1982	<i>Hoplomys gymnurus</i>	Echimyidae	Rodentia	South America
"	<i>Proechimys semispinosus</i>	Echimyidae	Rodentia	South America
<i>Echimytricalges mesomys</i> Fain, Lukoschus et Mendez, 1982	<i>Mesomys hispidus</i>	Echimyidae	Rodentia	South America
<i>Echimytricalges whitakeri</i> Fain et Ritzi, 2010	<i>Proechimys brevicauda</i>	Echimyidae	Rodentia	South America
"	<i>Proechimys cuvieri</i>	Echimyidae	Rodentia	South America
<i>Echimytricalges guyanensis</i> Fain, 1970	<i>!Caluromys philander</i>	Didelphidae	Didelphimorphia	South America
"	<i>Proechimys simonsi</i>	Echimyidae	Rodentia	South America

Table 19. Host distribution of the subfamily Paracoroptinae Lavoipierre, 1955

Mite species	Host species	Host family	Host order	Distribution
<i>Paracoroptes</i> Lavoipierre, 1955				
<i>Paracoroptes gordonii</i> Lavoipierre, 1955	<i>Cercopithecus mona</i>	Cercopithecidae	Primates	Africa
“	<i>Cercopithecus nictitans</i>	Cercopithecidae	Primates	Africa
<i>Paracoroptes allenopitheci</i> Fain, 1963	<i>Allenopithecus nigroviridis</i>	Cercopithecidae	Primates	Africa
<i>Paracoroptes colobi</i> Fain, 1963	<i>Colobus polykomos</i>	Cercopithecidae	Primates	Africa
<i>Paracoroptes natalensis</i> Fain et Segerman, 1978	<i>Thryonomys swinderianus</i>	Thryonomyidae	Rodentia	Africa
<i>Pangorillalges</i> Fain, 1962				
<i>Pangorillalges pani</i> Fain, 1962	<i>Pan troglodytes</i>	Hominidae	Primates	Africa
<i>Pangorillalges gorillae</i> (Gaud et Till, 1957)	<i>Gorilla gorilla</i>	Hominidae	Primates	Africa