A NEW SPECIES OF THE FEATHER MITE GENUS AMERODECTES VALIM ET HERNANDES, 2010 (ACARIFORMES: PROCTOPHYLLODIDAE) FROM XANTHOCEPHALUS XANTHOCEPHALUS (PASSERIFORMES: ICTERIDAE) IN OREGON

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ABSTRACT: A new feather mite species, *Amerodectes xanthocephali* sp. n. (Astigmata: Proctophyllodidae: Pterodectinae), is described from the *Xanthocephalus xanthocephalus* (Bonaparte) (Passeriformes: Icteridae) in Oregon, USA. The new species *A. xanthocephali* is most close to *A. sicalis* Mironov et González-Acuña 2011, *A. phrygilus* Mironov et González-Acuña 2011 and *A. zonotrichiae* Mironov et González-Acuña 2014. Males of *A. xanthocephali* differ from these species in having the aedeagus extending to the posterior margin of anal suckers (vs. to the midlevel of these suckers in *A. phrygilus* and *A. zonotrichiae*, and to the anterior end of terminal cleft in *A. sicalis*). Females of *A. xanthocephali* most clearly differ from these species in having the apodemes of oviporus separated from epimerites IIIa (vs. fused), the anterior hysteronotal and the lobar shields completely separated from each other (vs. connected ventro-laterally), and the posterior end of fused epimerites I tridentate (vs. with a short median extension). A brief review of taxonomic works on the *Amerodectes* is provided.

KEY WORDS: Feather mites, Acariformes, Proctophyllodidae, Amerodectes, systematics, Passeriformes

INTRODUCTION

The feather mite genus *Amerodectes* Valim et Hernandes, 2010 (Proctophyllodidae: Pterodectinae) was established in the course of taxonomic and biodiversity investigations of proctophyllodids associated with passerines (Passeriformes) of South America (Valim and Hernandes 2010), and to date it has included 25 species (Mironov et al. 2008; Mironov and González-Acuña 2011, 2014; Mironov and OConnor 2014). As for most proctophyllodids, representatives of this genus live the wing feathers (primaries, secondaries, and tertialis) and the tail feather (retrices) of their avian hosts, where they are located in corridors formed by barbs on the ventral side of vanes.

Within the subfamily Pterodectinae, the genus Amerodectes belongs the Pterodectes generic complex, also referred as derived pterodectines of the New World (Mironov 2009; Valim and Hernandes 2009, 2010; Hernandes 2013; Mironov and González-Acuña 2014). Valim and Hernandes (2010) summarized host associations of all Amerodectes species known up to that time. Modern redescriptions and illustrations of all species described in the 19th and 20th Centuries and presently belonging to the genus Amerodectes were given in the papers of Valim and Hernandes (2006, 2008, 2010). Species of this genus described or redescribed before 2010 were treated in the content of the genus Pterodectes Robin, 1877. A key to the majority of presently known Amerodectes species

was provided by Mironov and González-Acuña (2011).

The genus *Amerodectes* is the species-richest and most widely distributed genus of the Pterodectes complex (Mironov 2009; Valim and Hernandes 2010). Its representatives are known from birds of 10 families. The majority of previously known species (19 species) are associated with oscine passerines of the infraorder Passerida. Among them, 14 species were described from birds of the superfamily Passeroidea (Cardinalidae, Emberizidae, Icteridae, Parulidae and Thraupidae), four species are known from hosts of the superfamily Muscicapoidea (Turdidae and Mimidae), and one species is known from wrens, Troglodytidae (Certhioidea). Four species are known so far from suboscine passerines: three from birds of the family Tyrannidae, and one species from hosts of the family Furnariidae. Most Amerodectes species are known from South America, and only three species, A. dumetellae Mironov et OConnor, 2014, A. molothrus (Mironov, 2008), and Amerodectes sialiarum (Stoll, 1893), were recorded from North America (Park and Atyeo 1971; Hernandes and Valim 2005, 2006; OConnor et al. 2005; Valim and Hernandes 2010; Mironov and González-Acuña 2011, 2014; Mironov and OConnor 2014; Galloway et al. 2014).

In the present report we describe a new *Amerodectes* species from the Yellow-headed Blackbird, *Xanthocephalus xanthocephalus* (Bonaparte) (Passeriformes: Icteridae), captured in Oregon (USA).

MATERIAL AND METHODS

The material used in the present work was collected by RO in the course of parasitological investigation of various birds in The Summer Lake Wildlife Area (also known as Summer Lake State Game Management Area), Oregon, USA, in 2002 and 2014. Bird individuals were collected under the Federal Fish and Wildlife Permit and corresponding Oregon Scientific Taking Permits. Feather mites detected on birds were removed from feathers using fine forceps or a fine red sable spotting brush and placed into tubes with 70% ethanol. Then mite specimens were mounted on microslides in Hoyer's medium according to the standard technique for this group of mites (Krantz and Walter 2009).

The description is given according to the format elaborated for species of pterodectines (Hernandes and Valim 2006; Mironov et al. 2008, 2012; Mironov and González-Acuña 2011). General morphological terms and leg chaetotaxy follow Gaud and Atyeo (1996); idiosomal chaetotaxy also follows these authors with corrections for coxal setae proposed by Norton (1998). All measurements are in micrometers (µm). The measuring techniques used for particular structures are given in the following papers: Mironov et al. (2008, 2012), Mironov and González-Acuña (2011). Distance between setae of the same pair is the direct distance between their bases, and distance between different pairs of setae is the shortest distance between the transverse levels formed by the setae of respective pairs.

The taxonomic system and scientific names of birds follow Clements et al. (2014).

Type material depositories: BMOC — Museum of Zoology, University of Michigan (Ann Arbor, MI, USA), ZISP — Zoological Institute of the Russian Academy of Sciences (Saint-Petersburg, Russia).

SYSTEMATICS

Family Proctophyllodidae Mégnin et Trouessart, 1884 Subfamily Pterodectinae Park et Atyeo, 1971

Genus *Amerodectes* Valim et Hernandes, 2010

Amerodectes xanthocephali sp. n. Figs. 1–3

Type material. Male holotype (BMOC 15-0521-001), 10 male and 14 female paratypes from

the Yellow-headed Blackbird *Xanthocephalus xanthocephalus* (Bonaparte, 1826) (Passeriformes: Icteridae), USA, Oregon, Lake County, Summer Lake, 42°57'17.7"N, 120°42'43.6"W, July 2002, coll. R.M. Overstreet. Holotype, 5 male and 7 female paratypes — BMOC, 5 male and 7 female paratypes — ZISP.

Additional material. 7 male and 6 females, *X. xanthocephalus* (Passeriformes: Icteridae), USA, Oregon, Lake County, Summer Lake, 42°56′58.5″N, 120°45′54.2″W, July 2014, coll. R.M. Overstreet.

Description. Male (holotype, range for 10 paratypes in parentheses). Idiosoma, length \times width, 375 (360-385) × 175 (160-180), length of hysterosoma 230 (220-240). Prodorsal shield: entire, antero-lateral extensions with two unequal indentations, lateral margins slightly concave at level of scapular setae, posterior margin straight, posterior angles acute, length 106 (105-110), width 125 (120-130), surface with few minute lacunae in anterior half or without them (Fig. 1A); bases of scapular setae se separated by 62 (60-65). Setae ve present, rudimentary. Scapular shields narrow, scarcely developed dorsally. Humeral shields absent or represented by rudimentary sclerites antero-lateral to bases of setae cp. Setae cp and c2 situated on striated tegument. Subhumeral setae c3 lanceolate, $19(18-21) \times 8(7-8)$. Hysteronotal shield: greatest length 235 (230-245), width at anterior margin 125 (120-130), anterior margin straight, surface with minute lacunae, in some samples these lacunae in anterior half indistinct. Distance between prodorsal and hysteronotal shields about 20. Opisthosomal lobes approximately as long as wide at base; posterior margins of lobes roughly rounded, with small dent-like extensions at bases of setae h^2 and h^3 . Terminal cleft shaped as an inverted U with divergent branches, 30 (28-32) long. Supranal concavity semicircular. Setae f2 anterior to bases of setae ps2. Setae h1 situated at level of anterior end of terminal cleft. Setae h3 whip-like, 90 (80-95) long; setae ps2 88 (80–90) long; setae ps1 filiform, about 10 long, situated on margin of terminal cleft approximately at level of setae ps2. Distances between dorsal setae: c2:d2 98 (90-105), d2:e2 88 (85-95), e2:h3 44 (40-48), d1:d2 30 (28-33), e1:e2 32 (30-36), h1:ps2 22 (20-22), h2:h2 55 (53-60), h3:h3 40 (38-45), ps2:ps2 68 (65-70).

Epimerites I fused into a V, fused part with three small dent-like extensions (lateral ones in some specimens poorly expressed) (Fig. 1B).



Fig. 1. Amerodectes xanthocephali sp. n., male. A - dorsal view, B - ventral view.

Coxal fields I, II without extensively sclerotized areas. Rudimentary sclerites rEpIIa absent. Coxal fields I–III open. Coxal fields IV without sclerotized areas at bases of trochanters IV. Epimerites IVa absent. Genital arch of moderate size, 24 (22–26) × 51 (50–53); basal sclerite of genital apparatus with rounded posterior margin; aedeagus sword-shaped, 98 (95–105) long, extending to posterior margins of anal suckers or slightly beyond this level (Fig. 3E). Genital papillae not con-

nected at bases. Genital and adanal shields absent. Adanal suckers 13 (13–14) in diameter, corolla smooth, surrounding membrane with radial striae. Opisthoventral shields occupying lateral areas of opisthosoma and distal one third or half of opisthosomal lobes; inner margins of these shields at level of anal suckers with roughly trapezoidal or angular extensions, bearings setae ps3. Setae 4b situated slightly posterior to level of setae 3a. Distance between ventral setae: 4b:3a 9 (7-10), 4b:4a



Fig. 2. Amerodectes xanthocephali sp. n., female. A — dorsal view, B — ventral view. rh — rudimentary humeral shield.

38 (35–42), *4a*:*g* 44 (42–48), *g*:*ps3* 58 (55–60), *ps3*:*ps3* 69 (65–70), *ps3*:*h3* 30 (29–35).

Femora I, II with narrow ventral crests, other segments of legs I, II without processes (Figs 3A, B). Solenidion σI of genu I 10 (9–11) long, situated at midlevel of segment. Genual setae *cG*I, II and *mG*I filiform, setae *mG*II slightly thickened basally. Seta *d* of tarsi II subequal to corresponding seta *f*; seta *d* of tarsi III half as long as corresponding seta *f* (Fig. 3C). Solenidion φ of tibia IV extending to midlevel of ambulacral disc. Tarsus IV 31 (30–34) long, with apical process; seta *d* in basal half of segment (Fig. 3D). Length of solenidia: ω *I*I 11 (11–13), ω *I*II 8 (8–10), φ I 69 (65–70), φ II 53 (50–55), φ III 31 (28–33), φ IV 33 (32–36).

Female (range for 10 paratypes). Idiosoma, length × width, $500-530 \times 190-210$, length of hysterosoma 350-390. Prodorsal shield: shaped as in male, $125-135 \times 130-138$, surface with poorly distinct lacunae at posterior margin or without them, bases of setae *se* separated by 70–80 (Fig.



Fig. 3. *Amerodectes xanthocephali* sp. n., details. A — leg I of male, B — leg II of male, C — leg III of male, D — tibia and tarsus IV of male, E — opisthosoma of male, ventral view, F — spermatheca and spermaducts of female. co — copulatory opening, hs — head of spermathecal, pd — primary spermaduct, sd — secondary spermaduct. Scale bars: 50 μ m (A–D, F); 100 μ m (E).

2A). Setae *ve* present, rudimentary. Scapular shields narrow, not developed dorsally. Humeral shields absent or, represented by rudimentary sclerites antero-lateral to bases of setae *cp*. Setae *cp* and *c2* situated on striated tegument. Setae *c3* lanceolate, $22-24 \times 8-9$. Anterior and lobar parts of hysteronotal shield separated dorsally by narrow transverse band of soft tegument (Fig. 2B). Anterior hysteronotal shield nearly rectangular,

anterior margin straight, greatest length 270–290, width at anterior margin 130–145, posterior half with minute lacunae. Length of lobar region 82–90, greatest width 85–94. Terminal cleft narrow, with lateral margins parallel-sided and almost touching, 55–60 long. Supranal concavity poorly distinct; lobar shield with short posterior incision extending to area of supranal concavity; surface without ornamentation. Setae h1 at level of supra-

nal concavity; setae h1 and f2 arranged in a trapezium. Setae h2 spindle-like, $45-49 \times 8-9$. Setae ps1 situated near inner margins of opisthosomal lobes, close to lobar apices. Setae h3 18–22 long, about 1/8 the length of terminal appendages. Distances between dorsal setae: c2:d2 120–130, d2:e2118–130, e2:h2 60–65, h2:h3 35–40, d1:d2 55– 60, e1:e2 35–45, h1:h2 30–35, h1:h1 35–40, h2:h2 70–78, h2:ps1 25–30.

Epimerites I fused into a V, fused part with 3 small denticles (lateral ones in some specimens poorly expressed) (Fig. 2B). Lateral parts of coxal fields II without large sclerotized areas. Epimerites IVa absent. Translobar apodemes of opisthosomal lobes present, wide, not fused to each other anterior to terminal cleft. Epigynum with small lateral ledges, greatest width 70-78; apodemes of ovipore free from epimerites IIIa. Primary spermaduct with ampuliform enlargement near head of spermatheca; secondary spermaducts 30-35 long (Fig. 3F). Pseudanal setae filiform, setae ps2 situated at level of posterior half of anal opening and widely separated from each other; distance between pseudanal setae: ps2:ps2 40-46, ps3:ps3 18-21, ps2:ps3 24-30.

Femora II with ventral crest, other segments of legs I, II without processes. Solenidion $\sigma 1$ of genu I short, 9–11 long, situated at midlevel of segment. Genual setae *cG*I, II, *mG*I, II as in male. Seta *d* and *f* of tarsi II subequal, setae *d* of tarsi III, IV much shorter than corresponding setae *f*. Genu IV dorsally inflated, with narrow dorsal crest. Lengths of solenidia: ωII 11–13, ωIII 7–9, φI 68–72, φII 55–62, φIII 30–33, φIV 9–12.

Differential diagnosis. Among previously described species, the new species Amerodectes xanthocephali sp. n. is close to A. sicalis Mironov et González-Acuña 2011 from Sicalis luteola (Sparman), A. phrygilus Mironov et González-Acuña 2011 from Phrygilus patagonicus Lowe, and A. zonotrichiae Mironov et González-Acuña 2014 from Zonotrichia capensis (Statius Müller) (Emberizidae) in having the following combination of features: in both sexes, genual setae cGI, II are filiform; the anterior margin of the hysteronotal shield is straight or slightly concave, the humeral shields are absent or strongly reduced, and the hysteronotal shield is without ornamentation of large ovate lacunae; in males, setae h3 are long and exceed the distance between bases of setae se, and the end of fused epimerites I is tridentate and free from epimerites II; in females, the supranal concavity is poorly outlined.

The new species differs from these three species by the following features: in males of A. xanthocephali, the aedeagus extends to the posterior margin of adanal suckers (vs. to the midlevel of adanal suckers in A. phrygilus and A. zonotrichiae, and to the anterior end of terminal cleft in A. sicalis); in females of A. xanthocephali, the apodemes of oviporus are separated from epimerites IIIa (vs. fused to), the anterior hysteronotal and lobar shields are completely separated from each other (vs. connected ventro-laterally), the posterior end of fused epimerites I is tridentate (vs. with a short median extension), and setae ps2 are situated at level of anal opening (vs. posterior to anal opening). Additionally, A. xanthocephali differs from A. phrygilus by the following characters: in males of A. xanthocephali, setae d1 are noticeably closer to the level of setae d2 than to that of setae c2 (vs. equidistant from levels of c2 and d2in A. phrygilus), the distal half of tarsus IV extends beyond the level of lobar apices (vs. not extending to); in females, the humeral shields are rudimentary or absent (vs. represented by small sclerites touching bases of setae cp).

Etymology. The specific epithet is derived from the generic name of the type host and is a noun in the genitive case.

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REFERENCES

- Clements, J.F., Schulenberg, T.S., Iliff, M.J., Sullivan, B.L., Wood, C.L., and Roberson, D. 2014. The eBird/Clements checklist of birds of the world: Version 6.9. http://www.birds.cornell.edu/clementschecklist/download/ (Accessed 13 October 2014).
- Galloway, T.D., Proctor, H.C., and Mironov, S.V. 2014.
 Chapter 5. Chewing Lice (Insecta: Phthiraptera: Amblycera, Ischnocera) and Feather Mites (Acari: Astigmatina: Analgoidea, Pterolichoidea): Ectosymbionts of Grassland Birds in Canada. *In*: H.A. Cárcamo and D.J. Giberson (Eds.). Arthropods of

Canadian Grasslands (Volume 3): Biodiversity and Systematics, Part 1. Biological Survey of Canada, pp. 139–188.

- Gaud, J. and Atyeo, W.T. 1996. Feather mites of the World (Acarina, Astigmata): the supraspecific taxa. Annales du Musée Royal de l'Afrique Centrale, Sciences Zoologiques, 277, 1–193 (Pt. 1, text), 1–436 (Pt. 2, illustrations).
- Hernandes, F.A. 2013. A new genus and species of pterodectine feather mite (Acari: Proctophyllodidae) from Tod-Tyrants in Brazil (Passeriformes: Tyrannidae). *Acta Parasitologica*, 58: 309–316.
- Hernandes, F.A. and Valim, M.P. 2005. A new species of *Pterodectes* Robin, 1877 (Proctophyllodidae: Pterodectinae) from the pale-breasted thrush, *Turdus leucomelas* (Passeriformes: Turdidae). *Zootaxa*, 1081: 61–68.
- Hernandes, F.A. and Valim, M.P. 2006. Two new species of the feather mite subfamily Pterodectinae (Acari: Astigmata: Proctophyllodidae) from Brazil. *Zootaxa*, 1235: 49–61.
- Krantz, G. and Walter, D. (Eds). 2009. A Manual of Acarology. 3rd Edition. Texas Tech University Press, Lubbock, TX, USA, 807 pp.
- Mironov, S.V. 2009. Phylogeny of feather mites of the subfamily Pterodectinae (Astigmata: Proctophyllodidae) and their host associations with passerines (Aves: Passeriformes). Proceedings of the Zoological Institute of the Russian Academy of Sciences, 313: 97–118.
- Mironov, S.V. and González-Acuña, D.A. 2011. New feather mites of the subfamily Pterodectinae (Astigmata: Proctophyllodidae) from passerines (Aves: Passeriformes) from Chile and Cuba. *Zoo-taxa*, 3057: 1–48.
- Mironov, S.V. and González-Acuña, D.A. 2014. Two new species of the feather mite genus *Amerodectes* Valim et Hernandes, 2010 (Acariformes: Proctophyllodidae) from passerines (Aves: Passeriformes) of the New World. *Acarina*, 22 (1): 3–13.
- Mironov, S.V. and OConnor, B.M. 2014. New species of the feather mite family Proctophyllodidae (Acariformes: Astigmata) from two species of mockingbirds (Passeriformes: Mimidae) in East-

ern North America. *Journal of Medical Entomol-ogy*, 51: 529–546.

- Mironov, S.V., Literák, I., and Čapek, M. 2008. New feather mites of the subfamily Pterodectinae (Acari: Astigmata: Proctophyllodidae) from passerines (Aves: Passeriformes) in Mato Grosso do Sul, Brazil. *Zootaxa*, 1947: 1–38.
- Mironov, S.V., Literák, I., Nguen, M.H., and Čapek, M. 2012. New feather mites of the subfamily Pterodectinae (Acari: Proctophyllodidae) from passerines and woodpeckers (Aves: Passeriformes, Piciformes) from Vietnam. *Zootaxa*, 3440: 1–49.
- Norton, R. 1998. Morphological evidence for the evolutionary origin of Astigmata (Acari: Acariformes). *Experimental and Applied Acarology*, 22: 559–594.
- OConnor, B.M., Foufopoulos, J., Lipton, D., and Lindström, K. 2005. Mites associated with the small ground finch, *Geospiza fuliginosa* (Passeriformes: Emberizidae), from the Galapagos Islands. *Journal of Parasitology*, 91: 1304–1313.
- Park, C.K. and Atyeo, W.T. 1971. A generic revision of the Pterodectinae, a new subfamily of feather mites (Sarcoptiformes: Analgoidea). Bulletin of the University of Nebraska State Museum, 9: 39–88.
- Valim, M.P. and Hernandes, F.A. 2006. Redescription of four species of the feather mite genus *Pterodectes* Robin, 1877 (Acari: Proctophyllodidae: Pterodectinae) described by Herbert F. Berla. *Acarina*, 14 (1): 41–55.
- Valim, M.P. and Hernandes, F.A. 2008. Redescription of five species of the feather mite genus *Pterodectes* Robin, 1877 (Acari: Proctophyllodidae: Pterodectinae) with the proposal of a new genus and a new species. *Acarina*, 16 (2): 131–158.
- Valim, M.P. and Hernandes, F.A. 2009. A new species of the feather mite genus *Cotingodectes* and a new genus of Pterodectinae (Astigmata: Proctophyllodidae). *International Journal of Acarology*, 35: 265–272.
- Valim, M.P. and Hernandes, F.A. 2010. A systematic review of feather mites of the *Pterodectes* generic complex (Acari: Proctophyllodidae) with redescriptions of species described by Vladimir Černý. *Acarina*, 18 (1): 3–35.