

ON THE PHORESY AND MORPHOLOGY OF *PAVANIA CARABIDOPHILA* KHAUSTOV, 2005 (ACARI: DOLICHOCYBIDAE)

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ABSTRACT: The phoresy of *Pavania carabidophila* Khaustov, 2005 (Acari: Dolichocybidae) on the carabid beetle *Bembidion (Plataphus) lucillum* Bates, 1883 (Coleoptera: Carabidae) is documented for the first time with the aid of SEM microscopy. Supplementary description of previously unknown characters of *P. carabidophila* is provided. The attachment sites and type of attachments of dolichocybid mites are discussed.

KEY WORDS: Mites, carabid beetles, SEM microscopy, attachment.

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INTRODUCTION

The Dolichocybidae family is a small group of the early derivative heterostigmatic mites, which currently includes 2 subfamilies, 6 genera and 39 species in the world fauna (Hajiqaanbar and Khaustov 2010; Rahiminejad *et al.* 2011; Zhang *et al.* 2011; Loghmani *et al.* 2013; Bahramian *et al.* 2015). Little is known about the way of life of the dolichocybid mites, but, probably, all of them are fungivorous (Rack 1967; Magowski 1988; Kaliszewski *et al.* 1995). Adult females of dolichocybid mites utilize various insects for phoresy. Most dolichocybids are phoretic on various beetles of the families Bostrichidae, Carabidae, Curculionidae, Prostomidae, Scarabaeidae, Silvaniidae and Tenebrionidae (Sevastianov 1980; Magowski and Moser 1993; Khaustov 2005; Hajiqaanbar and Khaustov 2010; Rahiminejad *et al.* 2011; Loghmani *et al.* 2013; Katlav *et al.* 2014, 2015; Bahramian *et al.* 2015; Mortazavi *et al.* 2015); mites of the genus *Formicomotes* Sevastianov, 1980 are phoretic on ants (Sevastianov 1980). Mites from the genera *Acanthomastix*, *Dolichocybe*, *Dolichomotes* and the carabid-associated *Pavania* are phoretic under the elytra of beetles (Rahiminejad *et al.* 2011; Katlav *et al.* 2014; Mortazavi *et al.* 2015); scarabaeid-associated species usually attach to a soft membrane between head and pronotum, or between pronotum and elytra (Loghmani *et al.* 2013; Katlav *et al.* 2015). The mechanism of attachment of dolichocybid female to host beetle is unknown.

Pavania carabidophila Khaustov, 2005 was described from under elytra of the carabid beetle *Bembidion* sp. from southern part of the European Russia (Khaustov 2005). We collected one specimen of the beetle *Bembidion (Plataphus) lucillum* Bates, 1883 (Coleoptera: Carabidae) from the Far

East of Russia containing numerous females of *Pavania carabidophila* under its elytra. We studied the attachment of *P. carabidophila* females with the aid of SEM microscope for the first time. During the study of *P. carabidophila* using the compound microscope, some new morphological characters were revealed and their supplementary description is provided in this article.

MATERIAL AND METHODS

A living beetle *Bembidion (Plataphus) lucillum* Bates, 1883 was collected by junior author in Primorsky Krai, Lazo, 43° 30' 04.6" N, 133° 34' 47.1" E, 902 m a.s.l., 28 August 2016, on the bank of the stream. The beetle was brought to the laboratory at Tyumen State University and inspected for phoretic mites with the aid of the stereomicroscope Discovery V8 (Carl Zeiss, Germany). Alive mites attached to beetle without right elytron were scanned without dusting with the aid of a JEOL–JSM-6510LV SEM microscope. After that, mites were removed from the beetle and mounted on slides in Hoyer's medium. The identification of mites was provided with the aid of an AxioImager A2 compound microscope (Carl Zeiss, Germany) with phase contrast and DIC objectives. Micrographs were taken with the aid of digital camera Hitachi KP-HD20A. Terminology follows mostly that of Lindquist (1986). All mites are deposited in the mite collection of the Tyumen State University Museum of Zoology, Tyumen, Russia.

RESULTS

All phoretic mites were located in two large groups on tergites covered by left and right elytra

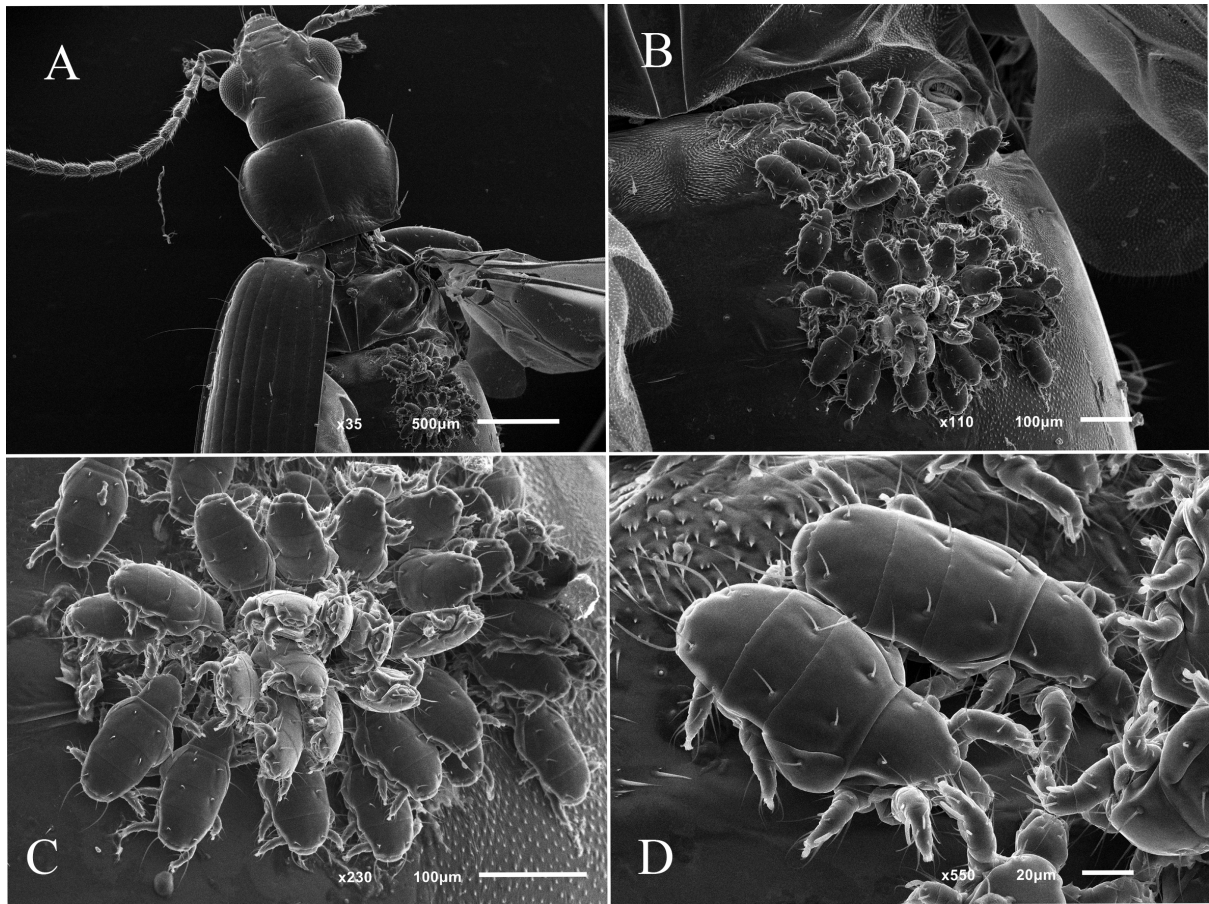


Fig. 1. SEM photos of females of *Pavana carabidophila* Khaustov, 2005 under elytra of *Bembidion lucillum* Bates, 1883 (right elytron removed): A—general view; B–D—detailed view.

(Fig. 1). Each group of mites consists of approximately 60 individuals. Most of the mite specimens were attached to the soft membrane of lateral parts of anterior abdominal tergites (Figs. 1A, B). Mites in each large group were oriented to particular small area; individuals above this area oriented vertically and tightly to each other (Fig. 1C). All individuals of mites were attached by their mouthparts to beetle host (Fig. 1D). The mechanism of attachment of mite gnathosoma to beetle membrane is not clear. Potentially, it could involve insertion of cheliceral stylets and pinching with palptibial claws.

Supplementary description of previously unknown characters of *P. carabidophila*

During this study, we found some morphological characters which were not illustrated in the original description of *Pavana carabidophila*. The most remarkable character is the presence of spine-like processes on trochanters I and II (Fig. 2A). Trochanter I with 5–6 dorsal processes; trochanter II with two processes. Such spine-like processes previously have never been recorded in any species

of *Pavana*. Another unusual character found in *P. carabidophila* is the presence of cone-like subcuticular structure located laterally to setae 1b (Fig. 2C). This structure has never been found in any dolichocybid mite. Tarsus I of *P. carabidophila* with ventrodistal membranous flange (Fig. 2D). This structure has never been found in any dolichocybid mite. Our examination of other available species of *Pavana* revealed the presence of this flange in all species. We also discern pharynx of *P. carabidophila*. It is narrow, with weak lateral projections (Fig. 2B). The pharynx of *P. carabidophila* considerably differs from scarabaeid-associated species of *Pavana* (see Fig. 3 in Bahramian *et al.* 2015), which have more short and wide pharynx with relatively long lateral projections.

DISCUSSION

A similar type of location of phoretic dolichocybid mite on a beetle host was recently documented for *Acantomastix derivatus* Katlav and Hajiqanbar, 2014 (Katlav *et al.* 2014). Females of this species are also attached to soft cuticle between

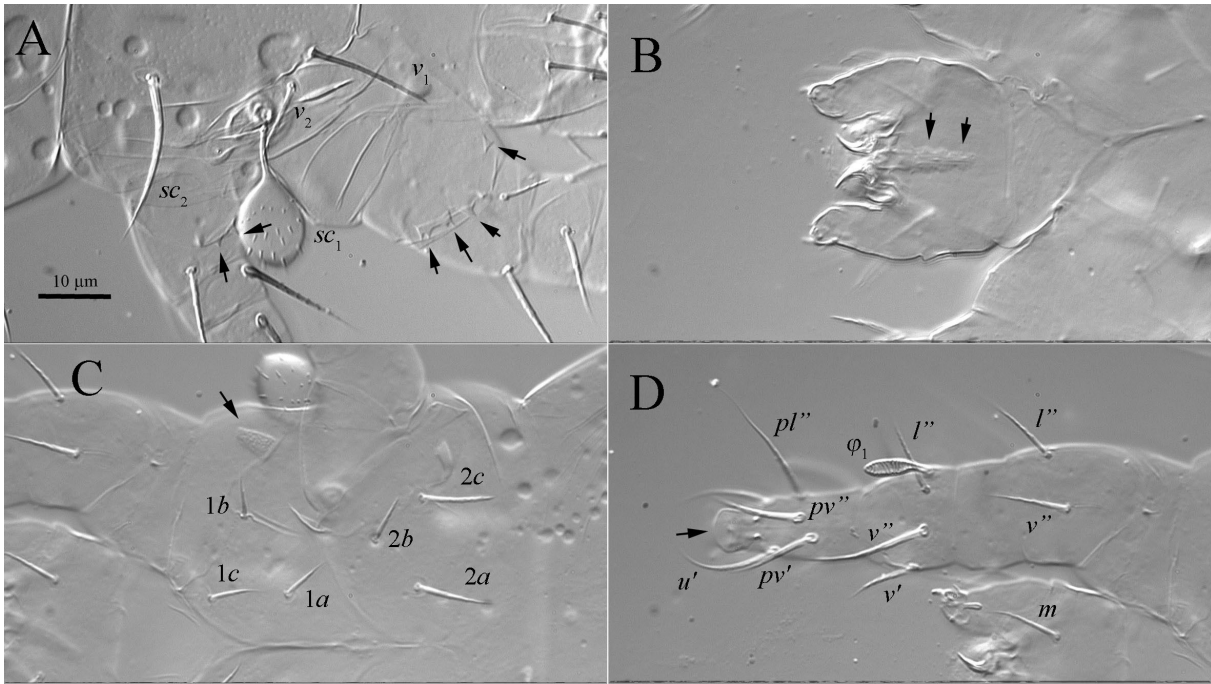


Fig. 2. DIC micrographs of females of *Pavania carabidophila* Khaustov, 2005: A—lateral part of prodorsum and bases of legs I and II; arrows point to spine-like processes; B—pharynx; C—lateral part of ventral propodosoma; arrow points to cone-like structure; D—leg I in ventral view; arrow points to membranous flange.

tergites on anterolateral corners of dorsal abdomen of host beetles *Uloma culinaris* (Linnaeus, 1758) and *Prostomis* sp. Dolichocybid mites located on anterolateral corners of dorsal abdomen beneath the elytra of beetles probably because it is the most protected place from the wind during the beetle's flight. Such attachment to the host by gnathosoma during phoresy is unique among Heterostigmata. Only mites of the family Carabaoacaridae exhibit a similar behavior. Nickel and Elzinga (1969) observed insertion of chelicerae of *Carabaoacarus karenae* Nickel and Elzinga, 1969 into hind wings of carabid beetles. However, carabaoacarid mites attached to the host mainly by modified sucker-like setae 1a and very large empodia on legs II-IV. The characteristic insertion of gnathosoma of dolichocybid mites into soft cuticle of host beetles suggests that they can acquire at least a liquid from the beetles. Kaliszewski *et al.* (1995) considered phoresy as preadaptation to permanent parasitism. In our opinion, phoresy in dolichocybid mites looks like the first step towards parasitism. The parasitic way of life is evident for the Crotalomorphidae, the second family of the superfamily Dolichocyboidea, the parasites of carabid beetles (Lindquist and Krantz 2002). On other hand, dolichocybid mites could suck a liquid from their hosts during phoresy. Such phenomenon was evident for heteromor-

phic deutonymphs of astigmatid mite *Hemisarcoptes cooremani* (Astigmata: Hemisarcoptidae), which at least acquired water from the hemolymph of host beetle *Chilocorus cacti* (Coleoptera: Coccinellidae) via caudal ventral suckers (Houck and Cohen 1995).

We do not know whether the dolichocybid mites acquire the material from beetle hosts or not. Further studies are required to solve this question.

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